

## Factors involved in the progression of CKD



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Chronic kidney disease (CKD) is common in both the dog and cat. The traditional view is that an underlying primary insult initiates renal disease and once established, intrinsic progression is then likely to follow, often due to mal-adaptive mechanisms that in the short-term act to try to normalise glomerular filtration rate (GFR). These mechanisms serve as attractive therapeutic targets for delaying progression of renal disease. However, given that the underlying cause of kidney disease in dogs and cats often cannot be identified, even when histopathology is performed,<sup>1</sup> it remains difficult to determine the relative importance of these putative mechanisms of progression, as opposed to further, uncharacterised, primary renal insults. Nonetheless, the success of certain treatments, such as ACE-inhibitors in humans with diabetic nephropathy,<sup>2</sup> and feeding renal diets to dogs and cats with CKD,<sup>3-5</sup> do confirm that delaying progression of renal disease is possible in certain circumstances.

CKD is an important health problem in humans, affecting up to 10% of adults.<sup>6</sup> There is evidence for considerable clustering of CKD within families and the heritability of GFR has been estimated at up to 36–75% in population-based studies.<sup>7</sup> The disease has huge societal cost, particularly when it progresses to end-stage renal disease (ESRD) necessitating dialysis, resulting in high morbidity and mortality and great financial burden. Efforts have been made to identify the factors responsible for progressive decline in renal function in humans. For example a recent genome-wide association study (GWAS) of annual decline in estimated GFR had more than 63000 participants.<sup>8</sup> Even with such a large sample size, only two novel loci were identified in that study. This is in spite of the fact that GFR is known to have high heritability and CKD clusters within families.<sup>7</sup>

Many studies of the pathophysiological processes underlying CKD progression in humans have relied on experiments performed in animals; primarily rodents, but sometimes dogs. Some of the most commonly employed research

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techniques are outlined in Table 1. It is evident that these studies have significant limitations as models of what happens in naturally occurring disease, not least because the onset of the renal functional impairment is usually slow in spontaneous disease and abrupt in the experimental models. Nevertheless, they have

contributed to our understanding of renal disease. In particular, sub-total nephrectomy models have elucidated the role of glomerular hypertension in disease progression and the potential value of haemodynamic interventions, such as treatment with ACE-inhibitors. Unilateral ureteral obstruction has allowed the study of factors involved in the development of tubulointerstitial fibrosis.<sup>9</sup> Currently, nephrology research interest is shifting slightly towards the role that proximal tubular injury plays in progressive renal disease.<sup>10</sup> In part, this stems from the recent recognition that in addition to CKD being a risk factor for acute kidney injury (AKI), patients that have had an episode of AKI, even one from which they have appeared to have fully recovered, are at increased risk of subsequently developing CKD.<sup>11</sup> It is suggested in humans that CKD is actually a 'slow-moving AKI' and this theory has been extended to dogs and cats.<sup>12</sup>

#### Risk Factors for Progressive CKD in Dogs and Cats

Azotaemia is not always progressive in dogs and cats. It is often said that while most dogs with established azotaemia will die of their renal disease, usually within a year, about half of all azotaemic cats will live long enough (often 2-3 years) to die of something else. Although the data to support these clinical observations is sketchy, there is some support for this from comparing the results of prospective diet studies in dogs and cats.<sup>3,4</sup>

One difficulty in comparing rates of disease progression in dogs and cats is that primary glomerular disease is rare in cats, and while it is uncommon in dogs too, it is encountered more frequently and as a sweeping generalisation, progression of this form of renal disease tends to be more rapid than with tubulo-interstitial disease.<sup>13</sup>

In one study of 213 azotaemic cats, progression (defined as an increase in creatinine of >25% from baseline) was documented to occur in 47% of cats within one year, 29% of cats with IRIS stage 2 disease and 63% of IRIS stage 3 CKD cats progressed to IRIS stage 4 before death.<sup>14</sup> Interestingly, the risk-factors for disease progression appeared to be slightly different in cats with differing severity of disease, in cats with stage 2 disease PCV and urine protein-creatinine ratio (UPC) predicted progression, whereas in cats with stage 3 disease only plasma phosphate concentration was significantly associated. This observation may be a result of the small group sizes however, and needs to be verified in independent studies.

Proteinuria has been identified as a risk factor for progressive renal disease and/or all-cause mortality in studies of both dogs and cats with naturally occurring disease.<sup>15-18</sup> This is in accordance with the findings of studies in humans.<sup>19</sup> The role that systemic hypertension plays in progressive renal injury in dogs and cats is difficult to characterise, due to a high degree of confounding with the severity of proteinuria. Hypertension has been associated with shortened survival in dogs with CKD,<sup>20</sup> however this effect was only evident when proteinuria was not included in the model. In cats with CKD and/or hypertension, when proteinuria is included in the analysis, blood pressure at diagnosis,<sup>17,18</sup> or averaged over time during follow-up,<sup>17</sup> does not remain as an independent predictor of survival. However, the relationship is complex since in hypertensive cats treated with amlodipine the cats in which hypertension was least well controlled (i.e. the cats with the highest blood pressure over time values) were also the cats that were most proteinuric.<sup>17</sup>

### Renal Pathology

The histological lesions found in the kidneys of most cats with CKD are predominantly found within the tubulointerstitium; glomerular lesions tend to be mild and are presumed to be a result of the glomerular hyperfiltration that occurs as a consequence of nephron loss and/or systemic hypertension.<sup>21</sup> The tubulointerstitial changes include tubular degeneration and atrophy, interstitial inflammation consisting of infiltration of lymphocytes, plasma cells and macrophages, and lipid accumulation.<sup>22</sup> These lesions are notably multifocal to segmental with the cellular and lipid infiltrate typically surrounding the degenerating tubules. Interstitial fibrosis accompanies these changes and is correlated with the severity of azotaemia.<sup>1,23</sup> In dogs, the main focus of renal pathology studies has been specific breed-related conditions and in the characterisation of primary glomerular disease.<sup>24</sup> Glomerulosclerosis, interstitial fibrosis, and tubular atrophy are reportedly common findings in aged dogs,<sup>25,26</sup> but there is relatively little information available to correlate histopathological findings with clinical diagnoses or measures of GFR.

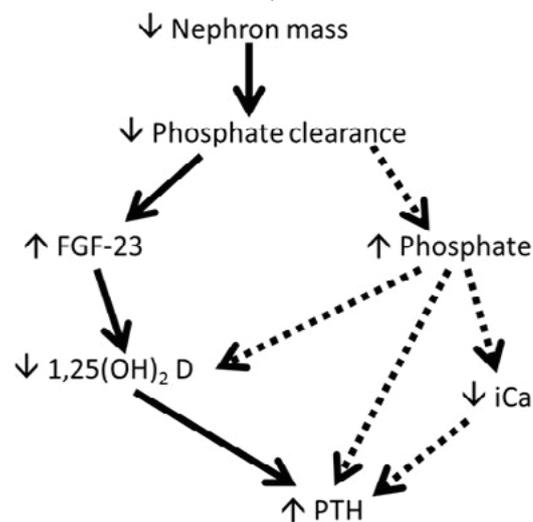
### Treatment Targets to Delay Progression of Renal Disease Hyperphosphataemia

In patients with CKD, as GFR declines, the amount of phosphate that is filtered by the kidney decreases. Initially, if the decline in GFR is not that great, this effect can be overcome by reducing the amount of phosphate that is reabsorbed in the proximal tubule. This reabsorption of the filtered phosphate load is inhibited by the actions of parathyroid hormone (PTH) and fibroblast growth factor-23 (FGF-23). The excessive secretion of PTH in patients with CKD is referred to as renal secondary hyperparathyroidism.

Traditionally this has been a focus for monitoring and treatment of patients with CKD. However, in human medicine attention has widened in the last decade to encompass changes in the concentrations of the other hormones as well as PTH. The preferred term for this syndrome is now CKD – mineral and bone disorder (MBD); this is defined as a systemic disorder of mineral and bone metabolism manifested by either one or a combination of the following:

- Abnormalities of calcium, phosphorus, PTH or vitamin D metabolism
- Abnormalities in bone turnover, mineralisation, volume, linear growth, or strength
- Vascular or other soft tissue calcification

The relative importance of the various humoral factors that are responsible for CKD – MBD varies according to the stage of renal disease. This syndrome is of great interest in human medicine because increases in FGF-23 are correlated not only with reduced survival of patients with advanced CKD requiring dialysis,<sup>27</sup> but also with much earlier stages of CKD.<sup>28</sup> Additionally FGF-23 predicts cardiovascular mortality and this is true in patients without CKD, although the effect is more marked in those with poor renal function.<sup>29</sup>



**Figure 1:** Simplified schematic illustrating the major adaptations/ maladaptations that occur as a result of decreased phosphate clearance in patients with CKD. Changes occurring with early/mild CKD. Changes occurring in the late stages/advanced CKD.

It has been known for some time that PTH concentrations are increased in cats with CKD, in many instances even when plasma phosphate remains within the laboratory reference range.<sup>30</sup> Similarly, PTH concentrations have also been shown to increase with severity of renal disease in dogs.<sup>31,32</sup> More recently it has been shown that FGF-23 concentrations are increased in cats,<sup>33</sup> and dogs,<sup>31</sup> with renal disease. As has been reported in human medicine, plasma FGF-23 is increased in early stage CKD in cats, prior to the onset of azotaemia.<sup>34</sup> The fact that these hormones are increased in patients with non-azotaemic renal disease suggests that, even with mild decreases in GFR, there are physiological adaptations occurring to maintain phosphate homeostasis. The importance of FGF-23 in handling the cat's phosphate load is supported by the observation that once the patient is azotaemic, if plasma phosphate is above the IRIS target range for that stage, FGF-23 is higher compared to those

cases where plasma phosphate is within the target range.<sup>33</sup> FGF-23 concentrations at initial presentation and diagnosis of CKD are predictive of progression of feline CKD and of all-cause mortality,<sup>35</sup> similar to the findings reported in human medicine. A previous study by our group shown that plasma phosphate was predictive of progression of CKD,<sup>14</sup> but when FGF-23 is included in the model it comes out as the strongest independent predictor of progression and displaces phosphate and PTH from the multivariate analysis.<sup>35</sup>

Dietary phosphate restriction, in the form of a diet designed for feeding to patients with CKD, has been associated with improved survival times in both cats,<sup>3,5,36</sup> and dogs.<sup>4</sup> Reductions in dietary phosphate intake have been shown to reduce both PTH,<sup>37</sup> and FGF-23,<sup>38</sup> concentrations in cats. In the future it will be interesting to see if setting particular targets for FGF-23 (and/or PTH) can further slow the progression of renal disease.

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### Renin-Angiotensin-Aldosterone System (RAAS)

Activation of both systemic and tissue-specific RAAS is widely accepted to occur in patients with CKD. Treatments targeting RAAS, including ACE-inhibitors, angiotensin receptor blockers (ARBs) and spironolactone have been extensively studied in animal models of CKD, and these drugs are widely used in both human and veterinary patients. This will be the subject of an accompanying lecture and is not discussed further here.

### Renal Fibrosis

Regardless of the underlying aetiology of CKD, renal fibrosis, characterised by accumulation of extracellular matrix proteins including collagens and fibronectin, is thought to represent the final common pathway in progression of renal disease. As a result, a great deal of research has been directed at identifying the factors that are important in driving this process. Much of this has been performed using the model of unilateral ureteral obstruction in mice, which allows the insult (ligation of a ureter, either completely or

partially) to be performed on transgenic animals and so to investigate the role of specific gene products on the ensuing inflammatory and fibrotic process. It is notable, however, that in spite of this research model being widely employed for over 20 years it has not, thus far, yielded any treatments for CKD that have made it to late-stage clinical trials.<sup>39</sup> This has led some authors to argue that fibrosis may not be

detrimental, but in fact could provide a scaffold for tissue repair, or else the fibrosis could simply be a marker for loss of proximal tubular cells that in health contribute half of the volume of the normal kidney.<sup>10</sup>

Transglutaminase-2 is an enzyme that cross-links collagen, stabilising the extracellular matrix. In the cat this enzyme has been shown to be up-regulated in renal tissue in patients with azotaemia and fibrosis.<sup>40</sup> Inhibitors of transglutaminase, selective for the extracellular space, are effective in reducing renal fibrosis in rat models of diabetic nephropathy,<sup>41</sup> and could in the future be used to slow the progression of renal disease.

### Hypoxia

Hypoxia is a proposed mechanism for progression of CKD. It may cause renal injury through activation of renal fibroblasts to produce matrix and/or cause mitochondrial derangements resulting in cellular apoptosis. Hypoxia may occur in diseased kidneys through a number of different mechanisms. Patients with CKD are often anaemic due to a relative lack of erythropoietin production, increased blood loss and iron deficiency. Angiotensin-II may increase in patients with CKD, constricting the efferent arterioles, and so increasing glomerular capillary pressure, but actually decreasing blood delivery to the peritubular capillary network. Finally, in the presence of tubular inflammation and fibrosis the diffusion distance from the peritubular capillaries to the tubular epithelial cells may increase, contributing to hypoxia. In cats, epidemiological studies have identified anaemia (or reduced PCV) as a risk factor for reduced survival time and progression of renal disease.<sup>14,42</sup> In humans with CKD anaemia worsens quality of life and increases

**Table 1:** Research methods used in the study of renal disease progression

Model	Species	Advantages/Successes	Disadvantages/Limitations
Subtotal nephrectomy	Rats, dogs & cats	Characterisation of glomerular hyperfiltration	Limited use in transgenic mice models due to small renal size Acute insult rather than chronic change
Unilateral ureteric obstruction	Theoretically any, but commonly mouse	Easily applied to transgenic mice Contralateral (control) kidney for comparison	No urine produced for biomarker discovery Reduced renal blood flow Limited systemic consequences (not azotaemic)
Genetic models	Theoretically any, but commonly mouse	Good models of specific diseases (e.g. Alport disease, polycystic kidney disease) but results may not be generalisable	Do not always re-capitulate the phenotype of other species (e.g. cystinosis)
Toxic nephropathies (e.g. cisplatin, doxorubicin)	Any (usually rats & mice)	Widely used as models of AKI, increasing interest in mini-AKI episodes as cause of CKD	Difficulty of injection (doxorubicin)
Streptozocin (Diabetic nephropathy)	Rats (usually)	Identified potential for ACE-inhibitors in preventing diabetic nephropathy	Mice become diabetic but have limited renal pathology, limiting genetic studies
Ischaemia-reperfusion injury	Most commonly rats. Recent work on cats	Renal fibrosis is demonstrated as an outcome	High morbidity Variable and often mild tubule- interstitial changes
Hypertensive models	Spontaneously hypertensive rat (& other strains) Two kidney, one-clip model (many species, including dogs)	Development of focal segmental glomerulosclerosis	Lack other components of metabolic syndrome

mortality.<sup>43</sup> However, while treatment with erythropoiesis stimulating agents (ESAs) is recommended when anaemia is severe, it is not usually recommended that haemoglobin concentrations are normalised, due to an increased risk of cardiovascular events. Treatment with ESAs has not been shown to delay progression of renal disease.<sup>44</sup>

#### **Oxidative Stress**

Oxidative stress increases in human patients with CKD.<sup>45</sup> This is proposed to be because of the high metabolic activity of renal tubular cells resulting in production of reactive oxygen species (ROS) together with a relative deficiency of anti-oxidant defence mechanisms. As a result it is proposed that damage to DNA and lipids may occur, resulting in cellular injury and ultimately stimulating inflammation and fibrosis through activation of NFκβ. However, treatment with different antioxidants has yielded variable results, leading to a Cochrane review to conclude that there was no evidence for their benefit.<sup>46</sup>

#### **Indoxyl sulphate**

Indoxyl sulphate is a product of tryptophan metabolism that accumulates in patients with reduced renal function and is incriminated in causing progression of CKD. It is classified as a uraemic toxin.<sup>47</sup> Tryptophan from protein in the diet is cleaved to indole by the gut microbiota, primarily in the distal colon. Synthesis of indole may be increased in patients with CKD due to dysbiosis. Indole is then sequentially oxidised and sulphated within the liver, yielding indoxyl sulphate. Indoxyl sulphate causes oxidative stress, increases fibrosis and activates the local RAAS. Indoxyl sulphate concentrations increase with IRIS stage of CKD in both dogs and cats.<sup>48</sup> Furthermore, a recent study found that indoxyl sulphate concentrations were higher in dogs and cats with subsequent progression of renal disease.<sup>49</sup>

#### **References**

1. Chakrabarti S, Syme HM, Brown CA et al. Histomorphometry of Feline Chronic Kidney Disease and Correlation With Markers of Renal Dysfunction. *Vet Pathol Online* 2013;50:147-155.
2. Wu H-Y, Huang J-W, Lin H-J et al. Comparative effectiveness of renin-angiotensin system blockers and other antihypertensive drugs in patients with diabetes: systematic review and bayesian network meta-analysis. *The BMJ* 2013;347:f6008.
3. Ross SJ, Osborne CA, Kirk CA, et al. Clinical evaluation of dietary modification for treatment of spontaneous chronic kidney disease in cats. *J Am Vet Med Assoc* 2006;229:949-957.
4. Jacob F, Polzin DJ, Osborne CA et al. Clinical evaluation of dietary modification for treatment of spontaneous chronic renal failure in dogs. *J Am Vet Med Assoc* 2002;220:1163-1170.
5. Elliott J, Rawlings JM, Markwell PJ et al. Survival of cats with naturally occurring chronic renal failure: effect of dietary management. *J Small Anim Pract* 2000;41:235-242.
6. Coresh J, Selvin E, Stevens LA et al. Prevalence of chronic kidney disease in the United States. *Jama* 2007;298:2038-2047.
7. O'Seaghda CM, Fox CS. Genome-wide association studies of chronic kidney disease: what have we learned? *Nat Rev Nephrol* 2011;8:89-99.
8. Gorski M, Tin A, Garnaas M et al. Genome-wide association study of kidney function decline in individuals of European descent. *Kidney Int* 2015;87:1017-1029.
9. Chevalier RL, Forbes MS, Thornhill BA. Ureteral obstruction as a model of renal interstitial fibrosis and obstructive nephropathy. *Kidney Int* 2009;75:1145-1152.
10. Chevalier RL. The proximal tubule is the primary target of injury and progression of kidney disease: role of the glomerulotubular junction. *Am J Physiol-Renal Physiol* 2016;311:F145-F161.
11. Heung M, Chawla LS. Acute Kidney Injury: Gateway to Chronic Kidney Disease. *Nephron Clin Pract* 2014;127:30-34.
12. Cowgill LD, Polzin DJ, Elliott J et al. Is Progressive Chronic Kidney Disease a Slow Acute Kidney Injury? *Vet Clin North Am Small Anim Pract* 2016;46:995-1013.
13. Cook AK, Cowgill LD. Clinical and pathological features of protein-losing glomerular disease in the dog: a review of 137 cases (1985-1992). *J Am Anim Hosp Assoc* 1996;32:313-322.
14. Chakrabarti S, Syme HM, Elliott J. Clinicopathological Variables Predicting Progression of Azotemia in Cats with Chronic Kidney Disease. *J Vet Intern Med* 2012;26:275-281.
15. Jacob F, Polzin DJ, Osborne CA et al. Evaluation of the association between initial proteinuria and morbidity rate or death in dogs with naturally occurring chronic renal failure. *J Am Vet Med Assoc* 2005;226:393-400.
16. Jepson RE, Brodbelt D, Vallance C et al. Evaluation of predictors of the development of azotemia in cats. *J Vet Intern Med* 2009;23:806-813.
17. Jepson RE, Elliott J, Brodbelt D et al. Effect of control of systolic blood pressure on survival in cats with systemic hypertension. *J Vet Intern Med* 2007;21:402-409.
18. Syme HM, Markwell PJ, Pfeiffer D et al. Survival of cats with naturally occurring chronic renal failure is related to severity of proteinuria. *J Vet Intern Med* 2006;20:528-535.
19. Peterson JC, Adler S, Burkart JM et al. Blood pressure control, proteinuria, and the progression of renal disease. The Modification of Diet in Renal Disease Study. *Ann Intern Med* 1995;123:754-762.
20. Jacob F, Polzin DJ, Osborne CA et al. Association between initial systolic blood pressure and risk of developing a uremic crisis or of dying in dogs with chronic renal failure. *J Am Vet Med Assoc* 2003;222:322-329.
21. Brown CA, Elliott J, Schmiedt CW et al. Chronic Kidney Disease in Aged Cats: Clinical Features, Morphology, and Proposed Pathogeneses. *Vet Pathol* 2016;53:309-326.
22. Martino-Costa AL, Malhão F, Lopes C et al. Renal Interstitial Lipid Accumulation in Cats with Chronic Kidney Disease. *J Comp Pathol* 2017;157:75-79.
23. McLeland SM, Cianciolo RE, Duncan CG et al. A comparison of biochemical and histopathologic staging in cats with chronic kidney disease. *Vet Pathol* 2015;52:524-534.

24. Schneider SM, Cianciolo RE, Nabity MB et al. Prevalence of Immune-Complex Glomerulonephritides in Dogs Biopsied for Suspected Glomerular Disease: 501 Cases (2007–2012). *J Vet Intern Med* 2013;27:S67-S75.
25. Cianciolo RE, Benali SL, Aresu L. Aging in the Canine Kidney. *Vet Pathol* 2015.
26. Chase K, Lawler DF, McGill LD et al. Age relationships of postmortem observations in Portuguese Water Dogs. *Age (Dordr)* 2011;33:461-473.
27. Gutiérrez OM, Mannstadt M, Isakova T et al. Fibroblast Growth Factor 23 and Mortality among Patients Undergoing Hemodialysis. *N Engl J Med* 2008;359:584-592.
28. Semba RD, Fink JC, Sun K et al. Serum fibroblast growth factor-23 and risk of incident chronic kidney disease in older community-dwelling women. *Clin J Am Soc Nephrol* 2012;7:85-91.
29. Ix JH, Katz R, Kestenbaum BR et al. Fibroblast growth factor-23 and death, heart failure, and cardiovascular events in community-living individuals: CHS (Cardiovascular Health Study). *J Am Coll Cardiol* 2012;60:200-207.
30. Barber PJ, Elliott J. Feline chronic renal failure: calcium homeostasis in 80 cases diagnosed between 1992 and 1995. *J Small Anim Pract* 1998;39:108-116.
31. Harjes LM, Parker VJ, Dembek K et al. Fibroblast Growth Factor-23 Concentration in Dogs with Chronic Kidney Disease. *J Vet Intern Med* 2017;31:784-790.
32. Cortadellas O, Fernández del Palacio MJ, Talavera J et al. Calcium and Phosphorus Homeostasis in Dogs with Spontaneous Chronic Kidney Disease at Different Stages of Severity. *J Vet Intern Med* 2010;24:73-79.
33. Geddes RF, Finch NC, Elliott J et al. Fibroblast Growth Factor 23 in Feline Chronic Kidney Disease. *J Vet Intern Med* 2013;27:234-241.
34. Finch NC, Geddes RF, Syme HM et al. Fibroblast Growth Factor 23 (FGF-23) Concentrations in Cats with Early Nonazotemic Chronic Kidney Disease (CKD) and in Healthy Geriatric Cats. *J Vet Intern Med* 2013;27:227-233.
35. Geddes RF, Elliott J, Syme HM. Relationship between Plasma Fibroblast Growth Factor-23 Concentration and Survival Time in Cats with Chronic Kidney Disease. *J Vet Intern Med* 2015;29:1494-1501.
36. Plantinga EA, Everts H, Kastelein AMC et al. Retrospective study of the survival of cats with acquired chronic renal insufficiency offered different commercial diets. *Vet Rec* 2005;157:185-187.
37. Barber PJ, Rawlings JM, Markwell PJ et al. Effect of dietary phosphate restriction on renal secondary hyperparathyroidism in the cat. *J Small Anim Pract* 1999;40:62-70.
38. Geddes RF, Elliott J, Syme HM. The Effect of Feeding a Renal Diet on Plasma Fibroblast Growth Factor 23 Concentrations in Cats with Stable Azotemic Chronic Kidney Disease. *J Vet Intern Med* 2013;27:1354-1361.
39. Kaissling B, LeHir M, Kriz W. Renal epithelial injury and fibrosis. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* 2013;1832:931-939.
40. Sanchez-Lara AC, Elliott J, Syme HM et al. Feline chronic kidney disease is associated with upregulation of transglutaminase 2: a collagen cross-linking enzyme. *Vet Pathol* 2015;52:513-523.
41. Huang L, Haylor JL, Hau Z et al. Transglutaminase inhibition ameliorates experimental diabetic nephropathy. *Kidney Int* 2009;76:383-394.
42. Boyd LM, Langston C, Thompson K et al. Survival in cats with naturally occurring chronic kidney disease (2000-2002). *J Vet Intern Med* 2008;22:1111-1117.
43. Fishbane S, Spinowitz B. Update on Anemia in ESRD and Earlier Stages of CKD: Core Curriculum 2018. *Am J Kidney Dis* 2018.
44. Elliott S, Tomita D, Endre Z. Erythropoiesis stimulating agents and reno-protection: a meta-analysis. *BMC Nephrol* 2017;18:14.
45. Dounousi E, Papavasiliou E, Makedou A et al. Oxidative stress is progressively enhanced with advancing stages of CKD. *Am J Kidney Dis* 2006;48:752-760.
46. Jun M, Venkataraman V, Razavian M et al. Antioxidants for chronic kidney disease. *Cochrane Database Syst Rev* 2012;10:Cd008176.
47. Ellis RJ, Small DM, Vesey DA et al. Indoxyl sulphate and kidney disease: Causes, consequences and interventions. *Nephrology (Carlton)* 2016;21:170-177.
48. Cheng FP, Hsieh MJ, Chou CC et al. Detection of indoxyl sulfate levels in dogs and cats suffering from naturally occurring kidney diseases. *Vet J* 2015;205:399-403.
49. Chen CN, Chou CC, Tsai PSJ, et al. Plasma indoxyl sulfate concentration predicts progression of chronic kidney disease in dogs and cats. *Vet J* 2018;232:33-39.