DEMONSTRATED PROTOCOL

Nuclei Isolation from Mouse Brain Tissue for Single Cell ATAC Sequencing

Overview

This protocol outlines how to isolate, wash, and count single nuclei from fresh, cryopreserved, and flash frozen mouse brain tissue samples for use with the Chromium Single Cell ATAC Solution. Tissue triturated into a nearly single cell suspension and subsequently frozen in media containing 10% DMSO produced metrics comparable to fresh tissue. High quality data can also be obtained using flash frozen tissue. Optimization of some protocol steps (e.g. homogenization, lysis reagent/time, centrifugation speed/time and filtration steps) may be needed when working with mouse brain tissues from different sources.

The recommended buffer compositions, final nuclei suspension concentration, and the wash step guidelines presented in this protocol for nuclei sample preparation are critical for optimal Single Cell ATAC assay performance. Failure to adhere to these guidelines may result in compromised microfluidics chip operation.

Additional Guidance

Consult Demonstrated Protocol Cell Preparation Guide (Document CG00053) for Tips & Best Practices.

Cells carry potentially hazardous pathogens. Follow material supplier recommendations and local laboratory procedures and regulations for the safe handling, storage and disposal of biological materials.

Cell Sourcing

Cell Type	Supplier	Part Number
E18 Mouse Combined Cortex, Hippocampus & Ventricular Zone	BrainBits, LLC	C57EHCV
Adult Mouse Cortex	BrainBits, LLC	C57ACX

Preparation – Buffers

Prepare all buffers fresh and maintain at 4°C

Wash Buffer	Stock	Final	2 ml
Tris-HCl (pH 7.4)	1 M	10 mM	20 µl
NaCl	5 M	10 mM	4 µl
$MgCl_2$	1 M	3 mM	6 µl
BSA	10%	1%	200 µl
Tween-20	10%	0.1%	20 µl
Nuclease-free Water	-	-	1.75 ml

Prepare all buffers fresh and maintain at 4°C

1X Lysis Buffer	Stock	Final	2 ml
Tris-HCl (pH 7.4)	1 M	10 mM	20 µl
NaCl	5 M	10 mM	4 µl
$MgCl_2$	1 M	3 mM	6 µl
Tween-20	10%	0.1%	20 µl
Nonidet P40 Substitute (alternatively, use IGEPAL CA-630) (If using Sigma 74385 or i8896, prepare a 10% stock)	10%	0.1%	20 µl
Digitonin (incubate at 65°C to dissolve precipitate)	5%	0.01%	4 µl
BSA	10%	1%	200 µl
Nuclease-free Water	-	-	1.732 ml
Lysis Dilution Buffer (may be prepared ahead)	**Stock**	**Final**	**2 ml**
Tris-HCl (pH 7.4)	1 M	10 mM	20 µl
NaCl	5 M	10 mM	4 µl
$MgCl_2$	1 M	3 mM	6 µl
BSA	10%	1%	200 µl
Nuclease-free Water	-	-	1.77 ml
0.1X Lysis Buffer	**Stock**	**Final**	**2 ml**
1X Lysis Buffer*	1X	0.1X	200 µl
Lysis Dilution Buffer	-	-	1.8 ml
Diluted Nuclei Buffer	**Stock**	**Final**	**1 ml**
Nuclei Buffer** (20X) (10x Genomics, PN 2000153/2000207)	20X	1X	50 µl
Nuclease-free Water	-	-	950 µl
Additional Buffers			
NbActiv-1 + 10% DMSO			

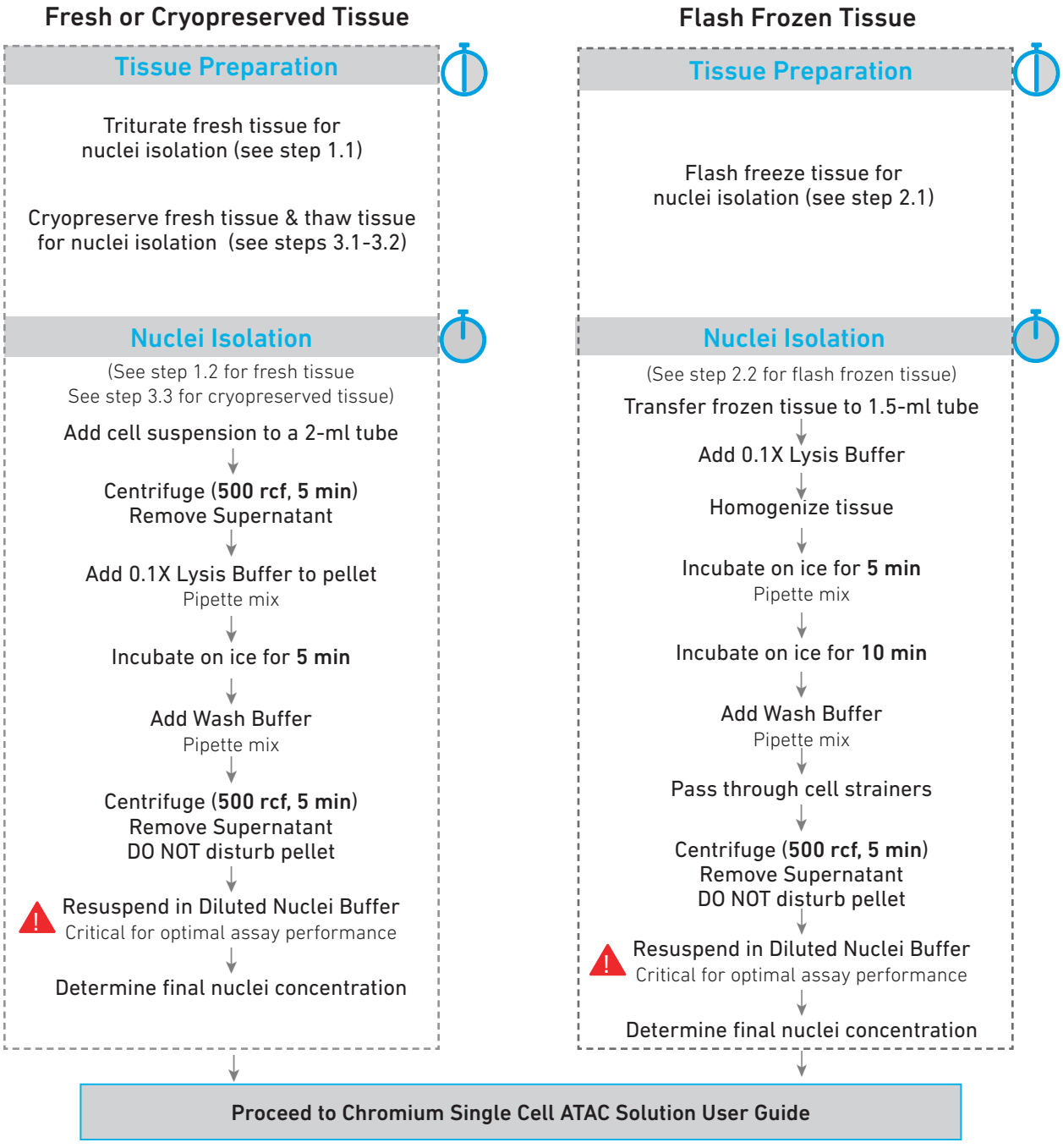
Specific Reagents & Consumables

Vendor	Item	Part Number
Thermo Fisher Scientific	Digitonin	BN2006
	LIVE/DEAD Viability/Cytotoxicity Kit	L3224
	Tween 20 Surfact-Amps Detergent	28320
VWR	Air-Tite All-Plastic Norm-Ject Syringes	53548-003
Miltenyi Biotec	MACS SmartStrainers (30 µm)	130-098-458
	MACS BSA Stock Solution	130-091-376
Millipore Sigma	Nonidet P40 (NP40) Substitute	74385
	IGEPAL CA-630	i8896
	Trizma Hydrochloride Solution, pH 7.4	T2194
	Sodium Chloride Solution, 5 M	59222C
	Magnesium Chloride Solution, 1 M	M1028
Fisher Scientific	RNase-Free Disposable Pellet Pestles	12-141-368
Bel-Art	Flowmi Cell Strainer, 40 µm	H13680-0040
	Flowmi Cell Strainer, 70 µm	H13680-0070

*DO NOT dilute 1X Lysis Buffer with Water. Dilute with Lysis Dilution Buffer.
**Included in the 10x Genomics Single Cell ATAC Library Kits.

10x GENOMICS

Protocol Overview



Protocol 1: Nuclei Isolation from Fresh Tissue

1.1 Tissue Preparation

Triturate Tissue:

a. Triturate the tissue in **1 ml** NbActiv1 with a wide-bore pipette tip to break the tissue into small pieces. If using embryonic brain, triturate with a regular-bore pipette tip.

b. Continue triturating with a regular-bore pipette tip until the tissue is completely broken up.

c. Pass the triturated tissue through a 30 µm MACS SmartStrainer into a 15-ml conical tube.

d. Use the back of a syringe to gently press the tissue through the strainer. DO NOT grind the tissue when passing through the strainer as this may disrupt the chromatin structure.

e. Pass **1 ml** NbActiv1through the strainer and collect the flowthrough in the same conical tube.

f. Determine the concentration using a Countess II FL Automated Cell Counter (see Appendix).

1.2 Nuclei Isolation

Nuclei may be isolated from 100,000-1,000,000 cells using this protocol.

a. Add cells to a 2-ml microcentrifuge tube and centrifuge at **500 rcf** for **5 min** at **4°C**.

b. Remove the supernatant without disrupting the cell pellet.

c. Add **100 µl** chilled 0.1X Lysis Buffer. Pipette mix 5x.

d. Incubate for **5 min** on ice.

e. Add **1ml** chilled Wash Buffer to the lysed cells. Pipette mix 5x.

f. Centrifuge cells at **500 rcf** for **5 min** at **4°C**.

g. Remove the supernatant without disrupting the nuclei pellet.

h. Based on the cell concentration at step 1.1f and assuming ~50% nuclei loss during cell lysis, resuspend in chilled Diluted Nuclei Buffer. See Nuclei Stock Concentration Table and Example Calculation in the User Guide. Maintain on ice.

The resuspension in Diluted Nuclei Buffer is critical for optimal Single Cell ATAC assay performance. The composition of the Tris-based Diluted Nuclei Buffer, including Magnesium concentration, has been optimized for the Transposition and Barcoding steps in the Single Cell ATAC protocol. Suspension of nuclei in a different buffer may not be compatible.

i. **OPTIONAL** If cell debris and large clumps are observed, pass through a cell strainer. For low volume, use a 40 µm Flowmi Cell Strainer to minimize volume loss.

j. Determine the nuclei concentration using a Countess II FL Automated Cell Counter (see Appendix).

k. Proceed **immediately** to the Single Cell ATAC Solution User Guide (see References).

Protocol 2: Nuclei Isolation from Flash Frozen Tissue

2.1 Tissue Preparation

Freeze Tissue:

a. Cut the tissue into small pieces (size of rice grain).

b. Flash freeze the tissue pieces in liquid nitrogen and transfer to a cryovial.

c. Transfer the cryovials to vapor-phase nitrogen for long-term storage. Tissue storage in -80°C for long-term use is not recommended.

2.2 Nuclei Isolation

DO NOT thaw the tissue prior to lysis.

a. Using forceps, transfer the frozen tissue to a 1.5-ml microcentrifuge tube.

b. Add **500 µl** chilled 0.1X Lysis Buffer. **Immediately** homogenize 15x using a Pellet Pestle.

c. Incubate for **5 min** on ice.

d. Pipette mix 10x with a wide-bore pipette tip (regular-bore pipette tip may be used if tissue disintegrates easily).

e. Incubate for **10 min** on ice.

f. Using a regular-bore pipette tip, add **500 µl** chilled Wash Buffer to the lysed cells. Pipette mix 5x.

g. Pass the suspension through a 70 µm Flowmi Cell Strainer into a 2-ml tube. Filter ~**300 µl** suspension at a time, each time through a new 70 µm strainer.

h. Pass the collected flowthrough through a 40 µm Flowmi Cell Strainer.

i. Determine the nuclei concentration using a Countess II FL Automated Cell Counter (see Appendix).

j. Centrifuge at **500 rcf** for **5 min** at **4°C**.

k. Remove the supernatant without disrupting the nuclei pellet.

l. Based on the cell concentration at step 2.2i and assuming ~50% nuclei loss during cell lysis, resuspend in chilled Diluted Nuclei Buffer. See Nuclei Stock Concentration Table and Example Calculation in the User Guide. Maintain on ice.

The resuspension in Diluted Nuclei Buffer is critical for optimal Single Cell ATAC assay performance. The composition of the Tris-based Diluted Nuclei Buffer, including Magnesium concentration, has been optimized for the Transposition and Barcoding steps in the Single Cell ATAC protocol. Suspension of nuclei in a different buffer may not be compatible.

m. **OPTIONAL** If cell debris and large clumps are observed, pass through a cell strainer. For low volume, use a 40 µm Flowmi Cell Strainer to minimize volume loss.

n. Determine the nuclei concentration using a Countess II FL Automated Cell Counter (see Appendix).

o. Proceed **immediately** to the Chromium Single Cell ATAC Solution User Guide (see References).

Protocol 3: Nuclei Isolation from Cryopreserved Tissue-Derived Cells

3.1 Tissue Derived Cell Freezing

a. Triturate the tissue in at least **1 ml** NbActiv1 with a wide-bore pipette tip to break the tissue into small pieces. If using embryonic brain, triturate with a regular-bore pipette tip.

b. Continue triturating with a regular-bore pipette tip until the tissue is completely broken up.

c. Centrifuge at **500 rcf** for **5 min** at **4°C**.

d. Remove the supernatant and resuspend the pellet in **1 ml** NbActiv1 media.

e. Centrifuge at **500 rcf** for **5 min** at **4°C**.

f. Remove the supernatant and resuspend the pellet in **1 ml** chilled NbActiv1 containing 10% DMSO.

g. Dispense **1 ml** cell suspension into pre-cooled cryovials and place the cryovials inside a pre-cooled cell freezing container e.g., CoolCell FTS30.

h. Place the cell freezing container in a **–80°C** freezer for **≥4 h**. Transfer the cryovials to vapor-phase nitrogen for long-term storage.

3.2 Tissue Derived Cell Thawing

a. Thaw the cryovial at a **37°C water bath** for **1-2 min**. Remove from the water bath when a tiny crystal remains in the cryovial.

b. Add **1 ml** warm NbActiv1. Pipette mix 3x.

c. Transfer the suspension to a 15-ml conical tube containing **10 ml** warm NbActiv1.

d. Place a 30 µm MACS SmartStrainers over a new 15-ml conical and pass the entire suspension through it.

e. Pass **1-2 ml** NbActiv1through the strainer and collect the flowthrough in the same conical tube.

f. Centrifuge at **500 rcf** for **5 min**.

g. Resuspend in **1-2 ml** NbActiv1.

h. Determine the concentration using a Countess II FL Automated Cell Counter (see Appendix for Nuclei Count).

3.3 Nuclei Isolation

Nuclei may be isolated from 100,000-1,000,000 cells using this protocol.

a. Add cells to a 2-ml microcentrifuge tube and centrifuge at **500 rcf** for **5 min** at **4°C**.

b. Remove all the supernatant without disrupting the cell pellet.

c. Add **100 µl** chilled 0.1X Lysis Buffer. Pipette mix 5x.

d. Incubate for **5 min** on **4°C**.

e. Add **1 ml** chilled Wash Buffer to the lysed cells. Pipette mix 5x.

f. Centrifuge at **500 rcf** for **5 min** at **4°C**.

g. Remove the supernatant without disrupting the nuclei pellet.

h. Based on the cell concentration at step 3.2h and assuming ~50% nuclei loss during cell lysis, resuspend in chilled Diluted Nuclei Buffer. See Nuclei Stock Concentration Table and Example Calculation in the User Guide. Maintain on ice.

The resuspension in Diluted Nuclei Buffer is critical for optimal Single Cell ATAC assay performance. The composition of the Tris-based Diluted Nuclei Buffer, including Magnesium concentration, has been optimized for the Transposition and Barcoding steps in the Single Cell ATAC protocol. Suspension of nuclei in a different buffer may not be compatible.

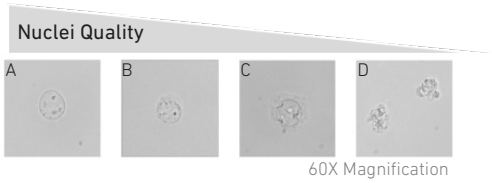
i. **OPTIONAL** If cell debris and large clumps are observed, pass through a cell strainer. For low volume, use a 40 µm Flowmi Cell Strainer to minimize volume loss.

j. Determine the nuclei concentration using a Countess II FL Automated Cell Counter (see Appendix for Nuclei Count).

k. Proceed **immediately** to the Chromium Single Cell ATAC Solution User Guide (see References).

Troubleshooting

Nuclei Quality - Representative Images (Panel A: recommended quality)



Appendix

Nuclei Count

This protocol provides instructions for counting nuclei using a fluorescent dye – Ethidium Homodimer-1 (part of LIVE/DEAD Viability/Cytotoxicity Kit) and the Countess II FL Automated Cell Counter (with RFP light cube) to enable accurate quantification even in the presence of sub-cellular debris. The optimal cell concentration for the Cell Counter is **1,000-4,000 cells/µl**. Refer to manufacturer's instructions for details on operations.

a. Vortex Ethidium Homodimer-1, centrifuge briefly, and dilute the concentrated stock as per manufacturer's instructions (~1:100 dilution) and aliquot **10 µl** Nuclear Stain in each tube.

b. Using a **regular-bore** pipette tip, gently mix the nuclei suspension. I**mmediately** add **10 µl** nuclei suspension to **10 µl** Nuclear Stain aliquot. Gently pipette mix 10x.

c. Transfer **10 µl** stained nuclei to a Countess II Cell Counting Slide chamber.

d. Insert the slide into the Countess II FL Cell Counter. Image the nuclei using the RFP setting for fluorescent illumination and filtering. Optimize focus and exposure settings. Confirm the absence of large clumps using the bright-field mode.
Note the RFP-positive concentration. Multiply by dilution factor 2 (from step b) to determine nuclei concentration.

Nuclei Stock Concentration Table

Based on the Targeted Nuclei Recovery, prepare the nuclei suspension in Diluted Nuclei Buffer to achieve the corresponding Nuclei Stock concentrations.

Targeted Nuclei Recovery	Nuclei Stock Concentration (nuclei/µl)
500	155-390
1,000	310-780
2,000	610-1,540
3,000	925-2,300
4,000	1,230-3,075
5,000	1,540-3,850
6,000	1,850-4,600
7,000	2,150-5,400
8,000	2,460-6,150
9,000	2,770-6,900
10,000	3,080-7,700

Example Calculation

Cell number for lysis in step 1.2a: **200,000**
Estimated total nuclei recovered based on count at step 1.2j (assuming ~50% loss): **100,000**
If targeting 5,000 Nuclei Recovery, nuclei pellet at step 1.2j may be resuspended in **30 µl** Diluted Nuclei Buffer for Nuclei Stock Concentration of 1,540-3,850 nuclei/µl (see Table above)

References

- Chromium Single Cell ATAC Reagent Kits User Guide (CG000168)
- Chromium Next GEM Single Cell ATAC Reagent Kits v1.1 User Guide (CG000209)
- Chromium Next GEM Single Cell ATAC Reagent Kits v2 User Guide (CG000496)

Contact:
support@10xgenomics.com
10x Genomics
7068 Koll Center Parkway
Suite 401
Pleasanton, CA 94566 USA