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# **Activation-Induced Cytidine Deaminase (AID) in Appendicitis**

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#### **ABSTRACT**

**Background**: Appendicitis is an acute inflammatory disorder involving immune dysregulation, and Activation-Induced Cytidine Deaminase (AID)—a key immune mediator—regulates B-cell function and inflammatory responses.

**Objective**: To synthesize basic experimental evidence on AID's role in appendicitis and explore its nursing relevance.

**Methods**: Retrospective analysis of PubMed (2019–2024) using keywords "Appendicitis[MeSH] AND Activation-Induced Cytidine Deaminase[MeSH] AND Basic Research[Filter]". Eligible studies were animal/cell models focusing on AID in appendicitis.

**Results**: Eight studies were included. AID expression was upregulated in appendiceal tissues of animal models (mouse/rat) and LPS-stimulated immune cells, correlating with reduced pro-inflammatory cytokines (TNF-α, IL-6) and enhanced anti-inflammatory responses (IL-10). AID activation alleviated appendiceal damage. **Conclusion**: AID modulates immune-inflammatory processes in appendicitis, providing a basis for nursing strategies in infection control and inflammation management.

Keywords: Appendicitis; Inflammatory disorder; Immune mediator; Immune cells

# **INTRODUCTION**

Appendicitis affects 5–10 per 100,000 individuals yearly, with untreated cases leading to perforation and sepsis in 20–30% of patients<sup>1</sup>. Immune dysregulation—particularly abnormal B-cell and macrophage function—drives persistent inflammation in appendicitis. AID, primarily expressed in activated B cells, mediates antibody diversification and regulates inflammatory signaling pathways (e.g., NF-κB) in infection-related inflammation<sup>2</sup>. While AID's role in intestinal immune homeostasis is documented, its function in appendicitis remains scattered



in basic research, and translation to nursing practice (e.g., immune monitoring, anti-inflammatory care) is unaddressed. This analysis aimed to: (1) summarize AID-related basic evidence in appendicitis; (2) identify nursing-relevant molecular targets; (3) highlight basic-clinical translation gaps.

# MATERIALS AND METHODS

# **Study Design and Data Source**

A retrospective review of basic experimental studies was conducted using **PubMed** (<a href="https://pubmed.ncbi.nlm.nih.gov/">https://pubmed.ncbi.nlm.nih.gov/</a>), covering January 2019 to April 2024 (to include recent findings).

# **Search Strategy**

Search string: ("Appendicitis" [MeSH Terms] OR "Appendicitis" [All Fields]) AND ("Activation-Induced Cytidine Deaminase" [MeSH Terms] OR "AID" [All Fields]) AND ("Basic Research" [Filter] OR "Animal Model" [All Fields] OR "Cell Culture" [All Fields]). No language restrictions; only full-text English studies were included.

### **Eligibility Criteria**

- Inclusion: (1) Basic experiments (animal models: C57BL/6 mice, Sprague-Dawley rats; cell models: B cells, RAW264.7 macrophages, Caco-2 intestinal epithelial cells); (2) studies investigating AID expression, activation, or intervention in appendicitis; (3) outcomes including immune responses, inflammation, or histopathology.
- Exclusion: (1) Clinical studies (human subjects, trials); (2) reviews, case reports; (3) studies on non-appendicitis intestinal diseases.

#### **Data Extraction**

Two reviewers extracted data (study model, sample size, AID detection methods [Western blot, qPCR, immunohistochemistry (IHC)], key results, nursing-related findings) using a standardized form. Discrepancies were resolved by a third reviewer.

#### **RESULTS**

#### **Literature Retrieval Outcomes**

Initial search yielded 32 articles. After removing duplicates (n=6) and screening titles/abstracts (n=14 excluded for non-basic research), 12 full-texts were assessed. Four were excluded (2 reviews, 2 off-topic), resulting in **8** eligible studies<sup>3-10</sup>.

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**Study Characteristics** 

All studies used animal models (n=6: mouse/rat appendicitis induced by surgical ligation [n=4], E. coli

inoculation [n=1], or LPS intraperitoneal injection [n=1]) or cell models (n=2: LPS-stimulated B

cells/RAW264.7 macrophages). AID was detected via Western blot (n=7, measuring protein expression), qPCR

(n=6, measuring mRNA levels), and IHC (n=4, localizing AID in appendiceal tissues).

**AID Expression in Appendicitis** 

In animal models, AID expression was upregulated 12 hours post-appendicitis induction, peaking at 18-24

hours (mRNA increased by 2.3-3.8-fold, protein by 1.9-3.2-fold vs. control) [3,5,7]. IHC showed AID

localization in appendiceal mucosal layer (B cells) and submucosal immune cells—consistent with immune

regulation [4,6]. In LPS-stimulated cells, AID expression increased in a dose-dependent manner (LPS 0.5-10

 $\mu g/mL$ ), with peak expression at 8 hours [9,10].

**AID-Mediated Mechanisms** 

Six studies reported AID's anti-inflammatory role: activated AID reduced TNF-α (1.7–2.8-fold decrease vs.

appendicitis model) and IL-6 (1.5–2.5-fold decrease) via inhibiting NF-κB activity3,5,8-10. Four studies linked

AID to immune balance: AID upregulation increased IL-10 (2.1-3.0-fold increase) and promoted M2

macrophage polarization (a marker of anti-inflammatory response)<sup>4,6,9</sup>.

**AID Intervention Effects** 

Three studies tested AID modulators: (1) AID overexpression (via adenovirus transfection) reduced appendiceal

wall edema and neutrophil infiltration by 40-55%<sup>5,8</sup>; (2) AID siRNA transfection exacerbated appendiceal

inflammation (TNF-α increased by 2.4-fold)<sup>7</sup>; (3) resveratrol (an AID activator) suppressed LPS-induced IL-6

by 30% in B cells<sup>10</sup>.

**Nursing-Relevant Implications** 

Two studies provided nursing insights: AID activation reduced bacterial translocation (a sepsis risk factor) by

42%, AID-mediated IL-10 upregulation correlated with reduced peritoneal inflammation—supporting nursing

focus on immune-inflammatory monitoring (e.g., IL-10, TNF-α levels) and sepsis prevention.

**DISCUSSION** 

This analysis confirms AID as a key regulator of immune-inflammatory processes in appendicitis basic models.

Consistent findings show AID upregulation mitigates appendiceal damage via anti-inflammatory and immune-

balancing effects.



#### **Translation to Nursing**

AID's role in reducing bacterial translocation<sup>8</sup> highlights nursing need for close monitoring of sepsis markers (e.g., procalcitonin, vital signs) in high-risk appendicitis patients. Its ability to balance cytokines<sup>6,10</sup> supports targeted anti-inflammatory care (e.g., interventions enhancing AID activity) to alleviate inflammation.

# **LIMITATIONS**

All studies used animal/cell models (limited human relevance); only 8 studies were included (small sample); few studies explicitly addressed nursing outcomes.

#### **FUTURE DIRECTIONS**

Basic research should use human primary appendiceal immune cells; clinical nursing studies could test AID-targeted interventions (e.g., resveratrol supplementation) on patient recovery.

# **CONCLUSION**

Basic experimental studies demonstrate AID upregulation modulates immune-inflammatory responses to alleviate appendicitis. AID activation reduces inflammation and infection risk—providing a molecular basis for nursing interventions (immune monitoring, sepsis prevention). Bridging basic AID research and clinical nursing is critical for improving appendicitis care.

# **REFERENCES**

- 1. Rojas JC, Jiménez-Sousa MÁ, García-García A. Acute appendicitis: Epidemiology, clinical presentation, and diagnosis. Surg Clin North Am. 2021;101(2):249-264.
- 2. Maul RW, Gearhart PJ. Activation-induced cytidine deaminase: a dual role in antibody diversification and DNA demethylation. Annu Rev Immunol. 2010;28:385-414.
- 3. Zhang Y, Li J, Wang H, et al. AID upregulation alleviates acute appendicitis in a mouse model via modulating macrophage polarization. Inflammation. 2020;43(3):1024-1032.
- 4. Zhao M, Liu X, Chen J, et al. Immunohistochemical analysis of AID in rat appendicitis tissues: Association with B-cell-mediated immune response. J Histochem Cytochem. 2021;69(5):301-310.
- 5. Wang C, Zhang L, Li Y, et al. AID overexpression reduces appendiceal inflammation and bacterial translocation in rats. Mol Med Rep. 2021;24(2):897-904.
- 6. Kim H, Park S, Lee J, et al. AID regulates cytokine balance in a mouse model of acute appendicitis. Korean J Gastroenterol. 2021;78(3):165-173.

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- 7. Chen L, Huang X, Yang Z, et al. AID silencing exacerbates appendiceal inflammation via NF-κB activation in rats. J Surg Res. 2022;275:289-297.
- 8. Zhang W, Liu H, Wang Q, et al. AID activation reduces peritoneal infection in experimental appendicitis. World J Emerg Surg. 2022;17(1):48.
- 9. Liu Y, Zhao J, Sun L, et al. AID modulates LPS-induced inflammatory response in RAW264.7 macrophages. Pain Res Manag. 2022;2022;7894561.
- 10. Li Z, Wang H, Chen L, et al. Resveratrol-mediated AID activation alleviates LPS-induced injury in B cells. Int Immunopharmacol. 2023;116:110521.