Clinical Guideline



Oscar Clinical Guideline: Bioengineered Skin and Soft Tissue Substitutes (CG030, Ver.10)

Bioengineered Skin and Soft Tissue Substitutes

Disclaimer

Clinical guidelines are developed and adopted to establish evidence-based clinical criteria for utilization management decisions. Clinical guidelines are applicable according to policy and plan type. The Plan may delegate utilization management decisions of certain services to third parties who may develop and adopt their own clinical criteria.

Coverage of services is subject to the terms, conditions, and limitations of a member's policy, as well as applicable state and federal law. Clinical guidelines are also subject to in-force criteria such as the Centers for Medicare & Medicaid Services (CMS) national coverage determination (NCD) or local coverage determination (LCD) for Medicare Advantage plans. Please refer to the member's policy documents (e.g., Certificate/Evidence of Coverage, Schedule of Benefits, Plan Formulary) or contact the Plan to confirm coverage.

Summary

Injuries and damage to the skin or underlying soft tissue can occur through a number of different mechanisms. Common causes may include severe burns, accidents, surgical procedures, and ulcers (e.g., diabetic or venous stasis ulcers). In most cases, conservative treatment, such as optimizing blood flow, preventing or treating infections, keeping the tissue moist, and clearing any dead tissue is adequate. However, some wounds require more advanced techniques to promote wound healing in the presence of other comorbidities or due to the extent of the damage. In such cases, biologic or synthetic skin/tissue products may be grafted to the wound. These skin/tissue products can provide living cells and/or a scaffold to encourage wound closure, promote tissue regeneration, or provide structural support. They can be "donated" from another site on the member's body or bioengineered depending on the specific indication. Such procedures are often part of a multidisciplinary wound care treatment plan.

This guideline outlines the clinical criteria, indications, and exclusions for bioengineered skin and soft tissue substitutes. This document does *not* address cosmetic or reconstructive skin procedures, blood-derived products for chronic wound healing, hyperbaric oxygen therapy, negative pressure wound therapy, suction devices, infrared or electrical stimulation, or other types of wound management.

Certain skin and soft tissue substitutes may be considered as part of medically necessary breast reconstruction as per the Plan Clinical Guideline: Breast Procedures (CG036).

Definitions

"Allograft (Allogeneic transplant)" is the transfer of tissue from a genetically non-identical donor of the same species.

"Bioengineered Skin Substitutes," also referred to as "Human Skin Equivalents (HSE)," are engineered, "artificial skin" products or combinations of skin products and other materials. They can be acellular or cellular, as defined below, and can be contained within a framework called a matrix, which can be synthetic or natural.

"Cellular Dressing" refers to a bioengineered skin substitute containing at least one layer of live cells (e.g., fibroblasts, keratinocytes, epidermal cells, dermal cells, etc). The cells can be derived from the member's own body (i.e., autologous), a human donor (i.e., allogeneic), or from another species (i.e., xenographic). They can also be derived from different tissues other than skin, such as placenta, intestine, or synthetic/composite materials. Cellular dressings provide a framework for the member's own body to begin wound healing as well as other growth factors and cells to facilitate this process. Some examples are, but not limited to:

- AmnioBand/Guardian (Q4151) an allograft placental matrix comprised of donated human amnion and chorion.
- Apligraf (Q4101) a living, bilayered (epidermal and dermal) skin substitute derived from human keratinocytes and fibroblasts
- Dermagraft (Q4106) a human fibroblast-derived dermal substitute
- Epicel (Q4100) a cultured epidermal autograft
- Epifix (Q4186) a multilayer biologic allograft derived from human amniotic membrane
- Grafix Core and Grafix PRIME (Q4132-Q4133) extracellular matrix containing growth factors
 designed as allografts with endogenous mesenchymal stem cells; Grafix Core is derived from
 chorionic placental tissue and Grafix PRIME is derived from the amniotic membrane of placental
 tissue
- OrCel (Q4100) a bilayered skin substitute consisting of epidermal keratinocytes and dermal fibroblasts in two layers of bovine collagen
- Theraskin (Q4121) a biologically active epidermal and dermal skin substitute consisting of cryopreserved human skin allograft, fibroblasts, keratinocytes, and extracellular matrix
- TransCyte (Q4100) a biosynthetic dressing consisting of allogeneic human dermal fibroblasts

"Acellular Dressing" refers to a bioengineered skin substitute containing matrix or scaffold materials (e.g., collagen, hyaluronic acid) but without any living cells. Acellular matrix provides a foundation for the member's cells to begin building upon to aid in wound healing. Some examples are, but not limited to:

• Allopatch (Q4128)- a hydrated allograft acellular dermal matrix uniquely derived from human tissue processed to remove epidermal and dermal cells

- Biobrane/Biobrane-L (Q4100) a biosynthetic wound dressing constructed from a silicon film with nylon impregnated into the dressing
- Cortiva (Allomax/NeoForm) (Q4100), Alloderm (Q4116), FlexHD (Q4128)
- DermACELL (Q4122) human acellular dermal matrix with ≥97% of donor DNA removed.
- Integra Bilayer Matrix Wound Dressing (Q4104), Integra Meshed Bilayer Wound Matrix (C9363), and Integra Dermal Regeneration Template (Q4105) cross-linked bovine tendon collagen and glycosaminoglycan and a semi-permeable polysiloxane (silicone) layers
- Graftjacket Regenerative Tissue Matrix (Q4107) a cadaveric dermal and epidermal skin substitute
- Oasis Wound Matrix (Q4102) an extracellular matrix material derived from the submucosal layer of porcine small intestine

"Grafting" is when a material (whether organic or synthetic) is transplanted to cover an injury or wound. There are several graft types:

- "Autologous" grafting (i.e., autograft) refers to a graft derived from the patient's own skin/tissue, and can be referred to as partial- or split-thickness depending on how it is harvested.
- "Allogeneic" grafting (i.e., allograft) refers to a graft derived from another human other than the member (e.g., cadaver).
- "Xenographic" grafting refers to a graft derived from non-human organisms (e.g., cows, pigs, etc.).
- "Synthetic" grafting refers to a graft that is derived from man-made materials.
- "Composite" grafting refers to a graft derived from various materials or organisms that can be interlinked to create a final product, such as a combination of human cells and synthetic matrix.

"Epidermolysis Bullosa (EB)" is a rare disease characterized by fragile skin and recurrent blisters resulting from minor irritation or trauma, typically presenting in early childhood. The resulting blisters may form large, painful wounds that resemble severe burns, and may require extensive grafting. Epidermolysis Bullosa can be categorized as Epidermolysis Bullosa Simplex (most common type with majority of mutations in the keratin genes KRT5 and KRT14), Junctional Epidermolysis Bullosa (majority of mutations in the laminin-332 genes), and Dystrophic Epidermolysis Bullosa (mutations in the COL7A1 gene).

Clinical Indications

General Criteria

In addition to the product-specific criteria outlined below, ALL of the following criteria must be met:

- 1. The procedure is performed by a licensed practitioner per state and federal law; and
- 2. Members using tobacco products have received cessation counseling and been informed of the impact of smoking on surgical outcomes prior to the procedure; *and*
- 3. Medical records document ALL of the following (unless the review of request is upon prior auth and wound characteristics are not available until after surgery, the surgery must meet medical necessity and the planned brand of tissue must be identified in request):
 - a. Medical necessity for the skin/tissue substitute; and

- b. Wound characteristics, including the size, location, depth, underlying conditions; and
- c. Previous methods, response, and duration of conservative therapy; and
- d. Full treatment plan that accompanies the bioengineered skin/tissue substitute.
- 4. The duration and frequency of the treatment plan is ordered as follows:
 - a. Treatment is limited to one initial application; and
 - b. Additional applications may occur at a minimum of 1 week intervals up to a maximum of 12 weeks (specific number of treatments may vary) *and* only when there is evidence of wound healing (e.g., reduced ulcer size, increasing epithelialization).

Product-Specific Criteria

Alginate or other fiber gelling dressing (A6196 - A6199)

Alginate or other fiber gelling dressing (A6196 - A6199) is considered medically necessary when the following criteria is met:

1. For moderate to heavy exudating partial and full thickness wounds (e.g., pressure ulcers, venous ulcers, diabetic ulcers, donor and graft sites, traumatic and surgical wounds, and 1st and 2nd degree burns).

AmnioBand/Guardian (Q4151), AlloPatch (AllopatchHD) (Q4128)

AmnioBand/Guardian (Q4151) or AlloPatchHD (Q4128) are considered medically necessary when the following criteria are met:

- 2. For diabetic foot ulcers, when the following characteristics are present:
 - a. The ulcer has no evidence of infection: and
 - b. Full thickness (i.e., extends through the dermis) neuropathic diabetic foot ulcer; and
 - c. The ulcer does not expose bone, tendon, muscle, or joint capsule; and
- A minimum of 4 weeks of appropriate conventional therapy has failed or been ineffective, as
 defined by standard dressing changes, debridement as necessary, and off-loading of
 pressure/weight: and
- 4. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%; and
- 5. The member must have adequate circulation to the affected extremity, and may be defined by one or more of the following:
 - a. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - b. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - c. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - d. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).

Artiss (C9250)

Artiss (C9250) is considered medically necessary when the following criteria are met:

- 1. Members age 1 or older with severe burns undergoing autologous skin grafting; and
- 2. Body surface area of burn(s) is not more than 40%

Breast Reconstructive Surgery

Cortiva (AlloMax/NeoForm) (Q4100), Alloderm/Alloderm RTU (Q4116), DermACELL (Q4122), or FlexHD (Q4128) are considered medically necessary when the following criteria are met:

- 1. For use in members when in conjunction with a medically necessary breast reconstructive surgery; and
- 2. The member had completed discussion with the provider about the risks and benefits of implant-based breast reconstruction with or without these acellular dermal matrices. (In general, the FDA has provided a safety communication that acellular dermal matrix products have not been approved/cleared for breast reconstruction).

Biobrane/Biobrane-L (Q4100)

Biobrane/Biobrane-L (Q4100) is considered medically necessary when ONE of the following criteria are met:

- 1. Biobrane: For the temporary covering of superficial, partial-thickness thermal injury; or
- 2. Biobrane-L: Covered when the criteria for Biobrane are met AND the dressing is used as an adjunct to a meshed autograft.

Dermagraft (Q4106)

Dermagraft (Q4106) is considered medically necessary when the following criteria are met:

- 1. Dermagraft is not being used in conjunction with Dakin's solution, chlorhexidine, polymyxin/nystatin, or any other cytotoxic solution due to breakdown and loss of effectiveness as demonstrated in clinical studies; *and*
- 2. Dermagraft is not ordered for members under the age of 18, pregnant women, members receiving immunosuppressive therapy (e.g., steroids, chemotherapy), or members with ulcers overlying Charcot's deformity; *and*
- 3. Treatment strictly adheres to FDA labeling guidelines, as documented by the 24-step procedure; and
- 4. For dystrophic epidermolysis bullosa wounds; or
- 5. For diabetic foot ulcers, when the following characteristics are present:
 - a. Full thickness (i.e., extends through the dermis) neuropathic diabetic foot ulcer; and
 - b. The ulcer does not expose bone, tendon, muscle, or joint capsule; and
 - c. The ulcer has no evidence of infection or a sinus tract; and
 - d. The member must have adequate circulation to the affected extremity, and may be defined by one or more of the following:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).

- e. A minimum of 6 weeks of appropriate conventional therapy has failed or been ineffective, as defined by standard dressing changes, debridement as necessary, and off-loading of pressure/weight; *and*
- f. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%.

DermACELL (Q4122)

DermACELL (Q4122) is considered medically necessary when the following criteria are met:

- 1. For diabetic foot ulcers, when ALL the following characteristics are present:
 - a. Full thickness (i.e., extends through the dermis) diabetic foot ulcer; and
 - b. The ulcer does not expose bone, tendon, muscle, or joint capsule; and
 - c. The ulcer has no evidence of infection or a sinus tract; and
 - d. The member must have adequate circulation to the affected extremity, and may be defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.8 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).
 - e. A minimum of 4 weeks of appropriate conventional therapy has failed or been ineffective, as defined by standard dressing changes, debridement as necessary, and off-loading of pressure/weight; *and*
 - f. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%.

Epicel (Q4100)

Epicel (Q4100) is considered medically necessary per FDA-approved Humanitarian Device Exemption (HDE) when ONE of the following criteria are met:

- 1. For deep-dermal or full-thickness burns reaching a total body surface area (BSA) \geq 30%; or
- 2. For use in conjunction with split-thickness autografts or alone in members who are not candidates for autografting given the severity and/or extent of the thermal injury or the instability of the current overall condition.

Epifix (Q4186) Dehydrated Human Amnion/Chorion Membrane (dHACM)

Epifix (Q4186) is considered medically necessary when the following criteria are met:

- 1. The ulcer has no evidence of infection or malignancy; and
- 2. The member does not have an autoimmune connective tissue disease; and
- 3. The patient is not being treated with radiation, chemotherapy, or COX-2 inhibitors; and
- 4. For diabetic foot ulcers, when the following characteristics are present:
 - a. Full thickness (e.g. extends through the dermis) neuropathic diabetic foot ulcer; and
 - b. The ulcer does not expose bone, tendon, muscle, or joint capsule; and

- c. A minimum of 4 weeks, and a maximum of 52 weeks, of appropriate conventional therapy has failed or been ineffective, as defined by standard dressing changes, debridement as necessary, and off-loading of pressure/weight: *and*
- d. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%; *and*
- e. The member must have adequate circulation to the affected extremity, and may be defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).
- f. The wound is between 1cm² and 25cm².
- 5. For venous insufficiency skin ulcers, when the following characteristics are present:
 - a. Ulcer must be chronic, non-infected, partial- or full-thickness and due to venous insufficiency; *and*
 - b. A minimum of *4 weeks* of appropriate conventional therapy has failed or been ineffective; *and*
 - c. Conventional therapy must have included at least 14 days of standard therapeutic compression; *and*
 - d. The member must have adequate circulation to the affected extremity, and may be defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).
 - e. The wound is between 2cm² and 20cm²; and
 - f. If the member has diabetes, the hemoglobin A1C is less than 10%; and
 - g. The ulcer is not on the dorsum of the foot or at least 50% of the ulcer is below the malleolus.

Grafix Core and Grafix PRIME (Q4132-Q4133) or Graftjacket Regenerative Tissue Matrix (Q4107) Grafix Core and Grafix PRIME (Q4132-Q4133) or Graftjacket Regenerative Tissue Matrix (Q4107) is considered medically necessary when the following criteria are met:

- 1. For Grafix Core and Grafix PRIME, the ulcer has no evidence of infection; and
- 2. For Graftjacket Regenerative Tissue Matrix, treatment is limited to ONE application; and
- 3. For diabetic foot ulcers, when the following characteristics are present:
 - a. Full thickness (i.e., extends through the dermis) neuropathic diabetic foot ulcer; and
 - b. The ulcer does not expose bone, tendon, muscle, or joint capsule; and

- c. A minimum of 4 weeks of appropriate conventional therapy has failed or been ineffective, as defined by standard dressing changes, debridement as necessary, and off-loading of pressure/weight: *and*
- d. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%; *and*
- e. The member must have adequate circulation to the affected extremity, and may be defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).

Integra Bilayer Matrix Wound Dressing (Q4104) or Integra Meshed Bilayer Wound Matrix (C9363) or Integra Dermal Regeneration Template (Q4105)

Integra Bilayer Matrix Wound Dressing (Q4104) or Integra Meshed Bilayer Wound Matrix (C9363) or Integra Dermal Regeneration Template (Q4105) are considered medically necessary when the following criteria are met:

- 1. Hemostasis must be obtained prior to application, as blood products may interfere with application; *and*
- 2. Integra Bilayer Matrix Wound Dressing, Integra Meshed Bilayer Wound Matrix, or Integra Dermal Regeneration Templates for deep partial and full-thickness thermal injury (i.e., second and third degree burns), when applied on the day of excision and when ONE of the following characteristics are present:
 - a. There is inadequate remaining skin to perform autografting; or
 - b. The member is too ill for further autografting.
- 3. Integra Dermal Regeneration Template for diabetic foot ulcers when ALL of the following characteristics are present:
 - a. Full thickness (e.g. extends through the dermis) neuropathic diabetic foot ulcer; and
 - b. The ulcer does not expose bone, tendon, muscle, or joint capsule; and
 - c. The member must have adequate circulation to the affected extremity, as defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).
 - d. A minimum of 6 weeks of appropriate conventional therapy has failed or been ineffective, as defined by standard dressing changes, debridement as necessary, and off-loading of pressure/weight; and
 - e. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%.

OrCel (Q4100)

OrCel (Q4100) is considered medically necessary when ONE of the following criteria are met:

- 1. To close and heal wounds in children with dystrophic epidermolysis bullosa who are undergoing hand surgery, including donor sites; *or*
- 2. To be applied to donor sites in members who have been burned and required autografting.

Apligraf (Q4101), Oasis Wound Matrix (Q4102), or Theraskin (Q4121)

Apligraf (Q4101), Oasis Wound Matrix (Q4102), or Theraskin (Q4121) is considered medically necessary when the following criteria are met:

- 1. Apligraf is not being used in conjunction with Dakin's solution, chlorhexidine, polymyxin/nystatin, or any other cytotoxic solution due to breakdown and loss of effectiveness as demonstrated in clinical studies; *and*
- 2. For diabetic foot ulcers, when all of the following characteristics are present:
 - a. Full thickness (e.g., extends through the dermis) neuropathic diabetic foot ulcer; and
 - b. The ulcer does not expose bone, tendon, muscle, or joint capsule; and
 - c. The member must have adequate circulation to the affected extremity, and may be defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg; or
 - iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).
 - d. Appropriate conventional therapy has failed or been ineffective, as defined by standard dressing changes, debridement as necessary, and off-loading of pressure/weight
 - i. For Apligraf, a minimum of 3 weeks; or
 - ii. For Oasis Wound Matrix or Theraskin, a minimum of 4 weeks; and
 - e. The member has a current diagnosis of type 1 or type 2 diabetes, and the hemoglobin A1C is less than 12%.
- 3. For venous insufficiency skin ulcers, when all of the following characteristics are present:
 - a. Ulcer must be chronic, non-infected, partial- or full-thickness and due to venous insufficiency; *and*
 - b. A minimum of *4 weeks* of appropriate conventional therapy has failed or been ineffective; *and*
 - c. The member must have adequate circulation to the affected extremity, and may be defined by:
 - i. Transcutaneous oxygen test (TcPO2) ≥30 mmHg; or
 - ii. Ankle-brachial index (ABI) between 0.7 and 1.2; or
 - iii. Triphasic or biphasic doppler arterial waveforms at the ankle of the affected leg;

- iv. Palpable foot pulses (e.g., dorsalis pedis or posterior tibial pulses).
- d. Conventional therapy must have included standard therapeutic compression.

TransCyte (Q4182)

TransCyte (Q4182) is considered medically necessary when ONE of the following criteria are met:

- 1. For deep partial- and full-thickness thermal injury (i.e., second and third degree burns) after surgical excision has been performed, in members who need temporary wound covering prior to autografting; *or*
- 2. For mid-dermal burns or those of an indeterminate depth that are expected to heal without autografting.

Experimental or Investigational / Not Medically Necessary

Applications beyond 12 weeks are not considered medically necessary. Retreatment of a successfully treated, healed ulcer is generally not covered.

3D bioprinted skin substitutes are considered experimental or investigational, as there is insufficient scientific evidence to evaluate its clinical safety and efficacy.

Skin/tissue substitutes or wound care treatments ordered for any indication not listed in the clinical indications or general criteria are considered investigational and/or experimental. The following skin/tissue substitutes or other wound care treatments are considered investigational and/or experimental (not all inclusive):

- Affinity™
- AlloMend™
- Alloskin AC
- AlloSkin RT
- AlloWrap®
- AlloWrap™ Dry
- AlloWrap™ DS
- Alphaplex[™] with MariGen Omega3[™]
- AmnioArmor™
- AmnioCare®
- AmnioClear®
- AmnioExcel™
- AmnioFix™
- AmnioGraft®
- Amniomatrix™

- Amnio-Maxx[™]
- AmnioMTM™
- AmnioShield®
- Amnio wound
- Aongen™ Collagen
 Matrix
- Architect
 Extracellular Matrix™
- Artacent®
- Artelon®
- ArthroFLEX™
- Atlas Wound Matrix
- Avance® Nerve Graft
- Avaulta Plus™
- AxoGuard® nerve connector

- Axolotl Graft or Axolotl DualGraft
- Belladerm®
- Bio-ConneKt®
- BioDDryFlex®
 Resorbable Adhesion
 Barrier
- BioDExCel™
- BioDfactor™
- BioDfence™
- BioDOptixTM
- BioFiber™
- BioVance®
- CellerateRX®
- Cellesta Cord™
- Cellesta Duo™

- Cellesta Flowable
 Amnion™
- CelluTome™
- CG CryoDerm™
- CLARIX™ 100
 Quick-Peel Wound
 Matrix
- CLARIX Cord 1k
- CLARIX™ FLO
- CollaFix™
- CollaGUARD®
- Collamend™
- CollaSorb™
- CollaWoundTM
- Coll-e-Derm RT™
- Collexa®
- Conexa[™]
- CorMatrix®
- C-QURTM
- CRXaTM
- CryoSkin®
- Cuffpatch™
- Cymetra®
- Cygnus®
- Cytal® Wound & Burn Matrix
- DeNovo® NT Graft
- Dermadapt™
 Wound Dressing
- Derma-Gide™
- DermaMatrix[™] (no longer distributed)
- DermaPure™
- DermaSpan™
- Dermavest[™]
- Dermavest 2™
- DressSkin™
- Duraform™
- Duragen™ Plus
- Duragen® XS
- DuraMatrix™

- Durepair®
 Regeneration Matrix
- Endobon®Xenograft Granules
- Endoform™
- ENDURAgen™
- EpiBurn®
- EpiCord®
- EpiDex®
- EpiFix™, particulate or injectable form
- Excellagen®
- E-Z Derm™
- FloGraft™
- FlowerAmnioFlo™ or FlowerFlo™
- FlowerAmnioPatch™ or FlowerPatch™
- FlowerDerm™
- FortaDerm™ Wound
 Dressing
- Fortiva™Porcine
 Dermis
- Gammagraft™
- Genesis Amniotic
 Membrane
- GORE BIO-A® Fistula Pluq
- Graftjacket[™] Xpress injectable
- GraftRope™
- Hyaluronic acid
 Absorbent Wound
 Dressing
- HelicollTM
- hMatrix®
- Hyalomatrix®
- Inforce®
- InnovaMatrix AC (Triad Life Sciences Inc.)

- Integra[™] Matrix
 Wound Dressing
- Integra Flowable
 Wound Matrix™
- InteguPly™
- Jaloskin®
- Kerecis®
- KeroxxTM
- LiquidGen™
- MariGen Omega3
- Matriderm®
- MatrionTM
- MatriStem® MicroMatrix
- MatriStem/ rebranded to Gentrix Surgical Matrix
- Matrix HD™
- Medeor™
- MediHoney®
- Mediskin®
- Memoderm™
- Menaflex[™] Collagen
 Meniscus Implant
- Meso BioMatrix™
- Microlyte Matrix (Imbed Biosciences)
- Nanofactor™ Flow
- Nanofactor™
 Membrane
- NeoPatch™
- NEOX® 100
 Quick-Peel Wound
 Matrix
- NEOX® 1k Wound Matrix
- NEOX®FLO
- Neuragen®
- NeuraWrap™
- Neuroflex™
- NeuroMatrix[™]

- NeuroMend™
- Novachor™
- NovoSorb® BTM,
 NovoSorb SynPath
- NuCel®
- NuShield®
- OrthADAPT™
- OsseoGuard®
- Ovation®
- OviTex, Reinforced
 Tissue Matrix (TELA
 Bio mesh)
- PalinGen Flow™
- PalinGenHydromembrane
- PalinGen Membrane
- PalinGen
 SportFlow™
- PalinGen XPlus
 Hydromembrane
- PalinGen XPlus
 Membrane
- Pelvicol®
- Pelvisoft®
- Peri-Guard® Repair
 Patch
- Peri-Strips Dry®
- Perlane®
- Permacol™
- Phasix Mesh™
- Preclude® Pericardial
 Membrane
- Preclude® Vessel Guard
- Primatrix™
- PTFF felt
- Puracol®
- Puraply™
- Puros® Dermis
- PX50® and X50®
 Plus

- Repliform®
- Repriza™
- Restore®
 Orthobiologic Soft
 Tissue Implant
- Restorigin Amnion
 Patch, Restorigin
 Amniotic Fluid
 Therapy™
- Restylane®
- Revita
- Revitalon™
- Seamguard®
- SERI® Surgical
 Scaffold
- SIS Wound DressingII
- SJM™ Pericardial
 Patch
- SkinTETM
- SportMatrix
- SportMeshTM
- SS Matrix[™]
- Sterishield II Amnion
 Patch
- Stimulen™ Collagen
- StrataGraft®
- Strattice™
- Stravix® / StravixPL
- Suprathel®
- SurgiGRAFT™,
 SurgiGRAFT nano,
 SurgiGRAFT-Dual
- SurgiMend®
- Surgisis® (including Surgisis® AFP™ Anal Fistula Plug, Surgisis® Gold™ Hernia Repair Grafts, and

- Surgisis®Biodesign TM)
- Symphony (Aroa Biosurgery)
- Talymed ™
- TenoGlide™
- TenSIXTM
- TEXAGEN™
 Amniotic Membrane
 Allograft
- TheraForm™
 Standard/Sheet
- TheraGenesis
 (Bioventus LLC)
- TissueMend®
- Tornier® BioFiber Absorbable
 Biological Scaffold
- TranzGraft®
- TutomeshTM
 Fenestrated Bovine
 Pericardium
- Tutopatch™ Bovine Pericardium
- Unite[™]
- Vascu-Guard®
- Veritas® Collagen
 Matrix
- VIM Amniotic
 Membrane (Cook
 Biotech)
- WoundEx®
- XCelliStem Wound Powder (Stemsys)
- XCM Biologic™
- Xelma®
- XenMatrix[™]
- XenMatrix[™] Surgical Graft
- XenoSure® Biologic
- X-Repair

• XWRAP® (Hydro, DRY, and ECM)

Applicable Billing Codes (CPT/HCPCS/ICD-10 Codes)

CPT/HCPCS Codes considered medically necessary if criteria are met:	
Code	Description
15002-15003	Surgical preparation or creation of recipient site by excision of open wounds, burn eschar, or scar (including subcutaneous tissues), or incisional release of scar contracture, trunk, arms, legs; first 100 sq cm or 1% of body area of infants and children. Or each additional 100 sq cm.
15004-15005	Surgical preparation or creation of recipient site by excision of open wounds, burn eschar, or scar (including subcutaneous tissues), or incisional release of scar contracture, face, scalp, eyelids, mouth, neck, ears, orbits, genitalia, hands, feet and/or multiple digits; first 100 sq cm or 1% of body area of infants and children. Or each additional 100 sq cm.
15100-15101	Split-thickness autograft, trunk, arms, legs; first 100 sq cm or less, or 1% of body area of infants and children (except 15050). Or each additional 100 sq cm.
15120-15121	Split-thickness autograft, face, scalp, eyelids, mouth, neck, ears, orbits, genitalia, hands, feet, and/or multiple digits; first 100 sq cm or less, or 1% of body area of infants and children (except 15050). Or each additional 100 sq cm.
15220-15221	Full thickness graft, free, including direct closure of donor site, scalp, arms, and/or legs; 20 sq cm or less. Or each additional 20 sq cm.
15240-15241	Full thickness graft, free, including direct closure of donor site, forehead, cheeks, chin, mouth, neck, axillae, genitalia, hands, and/or feet; 20 sq cm or less. Or each additional 20 sq cm.
15260-15261	Full thickness graft, free, including direct closure of donor site, nose, ears, eyelids, and/or lips; 20 sq cm or less. Or each additional 20 sq cm.
15271-15278	Application of skin substitute
15760	Graft; composite (eg, full thickness of external ear or nasal ala), including primary closure, donor area
15777	Implantation of biologic implant (eg, acellular dermal matrix) for soft tissue reinforcement (ie, breast, trunk) (List separately in addition to code for primary procedure)
A6196 - A6199	Alginate or other fiber gelling dressing, wound cover, sterile

C5271 - C5274	Application of low cost skin substitute graft to trunk, arms, legs
	Application of low cost skin substitute graft to face, scalp, eyelids, mouth, neck, ears, orbits, genitalia, hands, feet, and/or multiple digits
C9250	Human plasma fibrin sealant, vapor-heated, solvent-detergent (Artiss), 2ml
C9363	Skin substitute, Integra Meshed Bilayer Wound Matrix, per square centimeter
Q4100	Skin substitute, not otherwise specified (Must explain specific use and documentation) - Use for billing: TransCyte™, Orcel®, Biobrane Biosynthetic Dressing®, Epicel®, Cortiva® (AlloMax/NeoForm)
Q4101	Apligraf, per sq cm
Q4102	Oasis Wound Matrix, per sq cm
Q4104	Integra Bilayer Matrix Wound Dressing (BMWD), per sq sm
Q4105	Integra Dermal Regeneration Template (DRT), per sq cm
Q4106	Dermagraft, per sq cm
Q4107	Graftjacket, per sq cm
Q4116	Alloderm, per square centimeter
Q4121	TheraSkin, per sq cm
Q4122	DermACELL, DermACELL AWM or DermACELL AWM Porous, per sq cm
Q4128	FlexHD,Allopatch HD, per sq cm
Q4132	Grafix core, per sq cm
Q4133	Grafix prime, per sq cm
Q4151	Amnioband or guardian, per sq cm
Q4182	Transcyte, per square centimeter
Q4186	Epifix, per square centimeter
ICD-10 codes consideration Transcyte (Q4100):	dered medically necessary if criteria are met for Biobrane/Biobrane-L and
T20.011-T25.799	Burns
ICD-10 codes consi	dered medically necessary if criteria are met for Epicel (Q4100):
T20.011-T25.799, T31.30-T31.99, T32.30-T32.99	Burns
ICD-10 codes considered medically necessary if criteria are met for OrCel (Q4100):	
Q81.2	Epidermolysis bullosa dystrophica

T20.011-T25.799	Burns	
ICD-10 codes considered medically necessary if criteria are met for Cortiva(Allomax/NeoForm) (Q4100), Alloderm/ Alloderm RTU (Q4116), DermACELL(Q4122), or FlexHD (Q4128):		
C50.011 - C50.929	Malignant neoplasm of breast	
C79.81	Secondary malignant neoplasm of breast	
D05.00 - D05.92	Carcinoma in situ of breast	
Z15.01	Genetic susceptibility to malignant neoplasm of breast	
Z15.02	Genetic susceptibility to malignant neoplasm of ovary	
Z80.3	Family history of malignant neoplasm of breast	
Z80.41	Family history of malignant neoplasm of ovary	
Z85.3	Personal history of malignant neoplasm of breast	
Z90.10 - Z90.13	Acquired absence of breast and nipple	
Z92.3	Personal history of irradiation	
ICD-10 codes considered medically necessary if criteria are met for Apligraf (Q4101), Oasis Wound Matrix (Q4102), or Theraskin (Q4121):		
E08.621	Diabetes mellitus due to underlying condition with foot ulcer	
E09.621	Drug or chemical induced diabetes mellitus with foot ulcer	
E10.621	Type 1 diabetes mellitus with foot ulcer	
E11.621	Type 2 diabetes mellitus with foot ulcer	
E13.621	Other specified diabetes mellitus with foot ulcer	
183.001 - 183.029	Varicose veins of lower extremities with ulcer	
183.201 - 183.229	Varicose veins of lower extremities with ulcer and inflammation	
187.311 - 187.319	Chronic venous hypertension (idiopathic) with ulcer	
187.331 - 187.339	Chronic venous hypertension (idiopathic) with ulcer and inflammation	
ICD-10 codes cons	idered medically necessary if criteria are met for Dermagraft (Q4106):	
E08.621	Diabetes mellitus due to underlying condition with foot ulcer	
E09.621	Drug or chemical induced diabetes mellitus with foot ulcer	
E10.621	Type 1 diabetes mellitus with foot ulcer	
E11.621	Type 2 diabetes mellitus with foot ulcer	

E13.621	Other specified diabetes mellitus with foot ulcer
Q81.2	Epidermolysis bullosa dystrophica
ICD-10 codes considered medically necessary if criteria are met for AmnioBand (Q4151), AlloPatch (AllopatchHD) (Q4128), DermACELL (Q4122), Epifix (Q4186), Grafix Core and Grafix PRIME (Q4132-Q4133) or Graftjacket Regenerative Tissue Matrix (Q4107):	
E08.621, E09.621, E10.621, E11.621, E13.621	Diabetes mellitus
	idered medically necessary if criteria are met for Integra Bilayer Matrix Wound or Integra Meshed Bilayer Wound Matrix (C9363) or Integra Dermal Regeneration
E08.621, E09.621, E10.621, E11.621, E13.621	Diabetes mellitus
T20.011-T25.799	Burns

CPT/HCPCS codes considered experimental, investigational	
Code	Description
A2001	InnovaMatrix AC, per sq cm
A2004	XCelliStem, 1 mg
A2005	Microlyte Matrix, per sq cm
A2006	NovoSorb SynPath dermal matrix, per sq cm
A2008	TheraGenesis, per sq cm
A2009	Symphony, per sq cm
A2013	Innovamatrix FS, per sq cm
C9352	Microporous collagen implantable tube (NeuraGen Nerve Guide), per cm length
C9353	Microporous collagen implantable slit tube (NeuraWrap Nerve Protector), per cm length
C9354	Acellular pericardial tissue matrix of nonhuman origin (Veritas), per sq cm
C9356	Tendon, porous matrix of cross-linked collagen and glycosaminoglycan matrix (Tenoglide Tendon Protector Sheet), per square centimeter
C9358	Dermal substitute, native, non-denatured collagen (SurgiMend Collagen Matrix), per 0.5 square centimeters

C9360	Dermal substitute, native, non-denatured collagen, neonatal bovine origin (SurgiMend Collagen Matrix), per 0.5 square centimeters
C9361	Collagen matrix nerve wrap (NeuroMend Collagen Nerve Wrap), per 0.5 cm length
C9364	Porcine implant, Permacol, per square centimeter
G0428	Collagen meniscus implant procedure for filling meniscal defects (e.g., CMI, collagen scaffold, Menaflex)
Q4100	Skin substitute, not otherwise specified (use assigned code for covered product)
Q4103	Oasis Burn Matrix, per sq cm
Q4108	Integra Matrix Wound Dressing
Q4110	PriMatrix, per sq cm
Q4111	GammaGraft, per sq cm
Q4112	Cymetra, injectable, 1 cc
Q4113	GRAFTJACKET XPRESS, injectable, 1cc
Q4114	Integra Flowable Wound Matrix, injectable, 1 cc
Q4115	AlloSkin, per square centimeter
Q4117	HYALOMATRIX, per sq cm
Q4118	MatriStem micromatrix, 1 mg
Q4123	AlloSkin RT, per square centimeter
Q4124	OASIS ultra tri-layer wound matrix, per sq cm
Q4125	Arthroflex, per sq cm
Q4126	MemoDerm, DermaSpan, TranZgraft or InteguPly, per square centimeter
Q4127	Talymed, per sq cm
Q4130	Strattice, per sq cm
Q4134	hMatrix, per sq cm
Q4135	Mediskin, per sq cm
Q4136	E-Z Derm, per sq cm
Q4137	Amnioexcel or biodexcel, per sq cm
Q4138	Biodfence dryflex, per sq cm
Q4139	Amniomatrix or bioDMatrix, injectable, 1 cc
Q4140	Biodfence, per sq cm

Q4141	AlloSkin AC, per square centimeter
Q4142	XCM biologic tissue matrix, per sq cm
Q4143	Repriza, per sq cm
Q4145	Epifix, injectable, 1 mg
Q4147	Architect, architect PX, or architect FX, extracellular matrix, per sq cm
Q4148	Neox Cord 1K, Neox Cord RT, or Clarix Cord 1K, per sq cm
Q4149	Excellagen, 0.1 cc
Q4150	Allowrap DS or dry, per sq cm
Q4152	Dermapure, per sq cm
Q4153	Dermavest and Plurivest, per sq cm
Q4154	Biovance, per sq cm
Q4155	Neoxflo or clarixflo 1 mg
Q4156	Neox 100, per sq cm
Q4157	Revitalon, per sq cm
Q4158	Kerecis Omega3, per sq cm
Q4159	Affinity, per sq cm
Q4160	Nushield, per square centimeter
Q4161	Bio-connekt wound matrix, per square centimeter
Q4162	WoundEx Flow, BioSkin Flow, 0.5 cc
Q4163	Amniopro, Bioskin, Biorenew, Woundex, Amniogen-45, Amniogen-200, per square centimeter
Q4164	Helicoll, per square centimeter
Q4165	Keramatrix, per square centimeter
Q4166	Cytal, per square centimeter[Cytal Burn Matrix, Cytal Wound Matrix]
Q4167	Truskin, per square centimeter
Q4168	Amnioband, 1 mg
Q4169	Artacent wound, per square centimeter
Q4170	Cygnus, per square centimeter
Q4171	Interfyl, 1 mg
Q4173	Palingen or Palingen XPlus, per square centimeter
Q4174	Palingen or Promatrix, 0.36 mg per 0.25 cc

Q4175	Miroderm, per square centimeter
Q4176	NeoPatch, per sq cm
Q4177	FlowerAmnioFlo, 0.1 cc
Q4178	Floweramniopatch, per square centimeter
Q4179	FlowerDerm, per sq cm
Q4180	Revita, per sq cm
Q4181	Amnio wound, per square centimeter
Q4183	Surgigraft, per sq cm
Q4184	Cellesta or Cellesta Duo, per sq cm
Q4185	Cellesta Flowable Amnion (25 mg per cc); per 0.5 cc
Q4187	Epicord, per sq cm
Q4188	AmnioArmor, per sq cm
Q4189 - Q4190	Artacent
Q4191 - Q4192	Restorigin
Q4193	Coll-e-Derm, per sq cm
Q4194	Novachor, per sq cm
Q4195 - Q4197	Puraply
Q4198	Genesis Amniotic Membrane, per sq cm
Q4199	Cygnus matrix, per sq cm
Q4200	SkinTE, per sq cm
Q4201	Matrion, per sq cm
Q4202	Keroxx (2.5 g/cc), 1 cc
Q4203	Derma-Gide, per sq cm
Q4204	XWRAP, per sq cm
Q4210	Axolotl Graft or Axolotl DualGraft, per sq cm
Q4214	Cellesta Cord, per sq cm
Q4219	SurgiGRAFT-Dual, per sq centimeter
Q4239	Amnio-Maxx or Amnio-Maxx Lite, per sq cm
Q4251	Vim, per sq cm
S9055	Procuren or other growth factor preparation to promote wound healing

References

- 1. ACeLL®, Inc. Cytal® Burn Matrix Devices. Accessed May 22, 2023 https://www.acell.com/products/cytal-burn-matrix/
- 2. ACeLL®, Inc. Cytal® Wound Matrix Devices. Accessed May 22, 2023 https://www.acell.com/products/cytal-wound-matrix-devices/
- 3. ACeLL®, Inc. Gentrix® Surgical Matrix Devices. Accessed May 22, 2023 https://www.acell.com/products/gentrix-surgical-matrix/
- 4. ACeLL®, Inc. MicroMatrix®. Accessed May 22, 2023 https://www.acell.com/products/micromatrix-matrix/
- 5. Ahuja N, Jin R, Powers C, Billi A, Bass K. Dehydrated Human Amnion Chorion Membrane as Treatment for Pediatric Burns. Adv Wound Care (New Rochelle). 2020 Nov;9(11):602-611. doi: 10.1089/wound.2019.0983. Epub 2019 Dec 27.
- 6. Alvarez OM, Makowitz L, Patel M. Venous ulcers treated with a hyaluronic acid extracellular matrix and compression therapy: Interim analysis of a randomized controlled trial. Wounds 2017;29(7):E51-E54.
- 7. Aiolfi A, Gagner M, Zappa MA, Lastraioli C, Lombardo F, Panizzo V, Bonitta G, Cavalli M, Campanelli G, Bona D. Staple Line Reinforcement During Laparoscopic Sleeve Gastrectomy: Systematic Review and Network Meta-analysis of Randomized Controlled Trials. Obes Surg. 2022 May;32(5):1466-1478
- 8. Al-Abed YA, Ayers J, Ayantunde A, Praveen BV. Safety and efficacy of Permacol injection in the treatment of fecal incontinence. Ann Coloproctol. 2016;32(2):73-78.
- 9. American Diabetes Association. Standards of Medical Care in Diabetes—2013. Diabetes Care. 2013; 36(Suppl 1):S11-S66.
- 10. Armstrong DG, Meyr AJ. Basic principles of wound management. UpToDate.com. Waltham, MA: UpToDate; Last updated: June 9, 2022.
- 11. Armstrong DG, Tettelbach WH, Chang TJ, et al. (2021). Observed impact of skin substitutes in lower extremity diabetic ulcers: lessons from the Medicare Database (2015–2018). Journal of Wound Care North American Supplement, 30 (7).
- 12. Austin RE, Merchant N, Shahrokhi S, Jeschke MG. A comparison of Biobrane™ and cadaveric allograft for temporizing the acute burn wound: Cost and procedural time. Burns. 2015;41(4):749-753.
- 13. Ball CG, Kirkpatrick AW, Stuleanu T, et al. Is the type of biomesh relevant in the prevention of recurrence following abdominal wall reconstruction? A randomized controlled trial. Can J Surg. 2022; 65(4):E541-E549.
- 14. Ball, J.F., Sheena, Y., Saleh, D.M.T., et al. (2017). A direct comparison of porcine (Strattice™) and bovine (Surgimend™) acellular dermal matrices in implant-based immediate breast reconstruction. *J Plast Reconstr Aesthet Surg, 70* (8),1076-1082. doi: 10.1016/j.bjps.2017.05.015.
- 15. Bakri M, Lovato FC, Diosti GM, Salles YLSG, Moreira PHB, Collaço LM, Czeczko NG, Malafaia O, Kubrusly LF. Comparative Analysis of Tissular Response After AbdominalL Wall Repair Using Polypropylene Mesh and Bovine Pericardium Mesh. Arq Bras Cir Dig. 2022 Jan 5;34(3):e1527.

- 16. Bairagi A, Griffin B, Banani T, McPhail SM, Kimble R, Tyack Z. A systematic review and meta-analysis of randomized trials evaluating the efficacy of autologous skin cell suspensions for re-epithelialization of acute partial thickness burn injuries and split-thickness skin graft donor sites. Burns. 2021 Sep;47(6):1225-1240.
- 17. Barber C, Watt A, Pham C, et al. Influence of bioengineered skin substitutes on diabetic foot ulcer and venous leg ulcer outcomes. J Wound Care. 2008;17(12):517-527.
- 18. Barbul A, Gurtner GC, Gordon H, Bakewell K, Carter MJ. Matched-cohort study comparing bioactive human split-thickness skin allograft plus standard of care to standard of care alone in the treatment of diabetic ulcers: A retrospective analysis across 470 institutions. Wound Repair Regen. 2020 Jan;28(1):81-89.
- 19. Barret JP, Dziewulski P, Ramzy PI, et al. Biobrane versus 1% silver sulfadiazine in second-degree pediatric burns. Plast Reconstr Surg. 2000; 105(1):62-65.
- 20. Barmettler A, Heo M. A Prospective, Randomized Comparison of Lower Eyelid Retraction Repair With Autologous Auricular Cartilage, Bovine Acellular Dermal Matrix (Surgimend), and Porcine Acellular Dermal Matrix (Enduragen) Spacer Grafts. Ophthalmic Plast Reconstr Surg. 2018 May/Jun;34(3):266-273.
- 21. Barski D, Gerullis H, Ecke T, Boros M, Brune J, Beutner U, Tsaur I, Ramon A, Otto T. Application of Dried Human Amnion Graft to Improve Post-Prostatectomy Incontinence and Potency: A Randomized Exploration Study Protocol. Adv Ther. 2020 Jan;37(1):592-602. doi: 10.1007/s12325-019-01158-3. Epub 2019 Nov 28.
- 22. Bowers CA, Brimley C, Cole C, et al. AlloDerm for duraplasty in Chiari malformation: Superior outcomes. Acta Neurochir (Wien). 2015;157(3):507-511
- 23. Brosious JP, Wong N, Fowler G, et al. Evaluation of AlloMax acellular dermal matrix for objective collagen deposition. J Reconstr Microsurg. 2014;30(1):31-34.
- 24. Buchberger B, Follmann M, Freyer D, et al. The evidence for the use of growth factors and active skin substitutes for the treatment of non-infected diabetic foot ulcers (DFU): A health technology assessment (HTA). Exp Clin Endocrinol Diabetes. 2011;119(8):472-479.
- 25. Byrnes MC, Irwin E, Carlson D, et al. Repair of high-risk incisional hernias and traumatic abdominal wall defects with porcine mesh. Am Surg. 2011;77(2):144-150.
- 26. Cahan AC, Palaia DA, Rosenberg M, Bonanno PC. The aesthetic mastectomy utilizing a non-nipple-sparing portal approach. Ann Plast Surg. 2011; 66(5):424-428.
- 27. Caravaggi C, Francesco Grigoletto M, Scuderi N. Wound bed preparation with a dermal substitute (Hyalomatrix® PA) facilitates re-epithelialization and haling: Results of a multicenter, prospective, observational study on complex chronic ulcers: The FAST Study. Wounds. 2011;8(23):228-235.
- 28. Carlson M, Faria K, Shamis Y, et al. Epidermal stem cells are preserved during commercial-scale manufacture of a bilayered, living cellular construct (Apligraf®). Tissue Eng Part A. 2011;17(3-4):487-493.
- 29. Carsin H, Ainaud P, Le Bever H, et al. Cultured epithelial autografts in extensive burn coverage of severely traumatized patients: a five year single-center experience with 30 patients. Burns. 2000;26(4):379-387.

- 30. Cazzell S. A Randomized Controlled Trial Comparing a Human Acellular Dermal Matrix Versus Conventional Care for the Treatment of Venous Leg Ulcers. Wounds. 2019 Mar;31(3):68-74. Epub 2019 Jan 31.
- 31. Cazell S, Vayser D, Pham H, et al. A randomized clinical trial of a human acellular dermal matrix demonstrated superior healing rates for chronic diabetic foot ulcers over conventional care and an active acellular dermal matrix comparator. *The International Journal of Tissue Repair and Regeneration.* 2017;25(3):483-497. doi:https://doi.org/10.1111/wrr.12551
- 32. Capella-Monsonís H, Tilbury MA, Wall JG, Zeugolis DI. Porcine mesothelium matrix as a biomaterial for wound healing applications. Mater Today Bio. 2020 May 17;7:100057.
- 33. Cervigni M, Natale F, La Penna C, et al. Collagen-coated polypropylene mesh in vaginal prolapse surgery: An observational study. Eur J Obstet Gynecol Reprod Biol. 2011;156(2):223-227.
- 34. Chadwick P, Acton C. The use of amelogenin protein in the treatment of hard-to-heal wounds. Br J Nurs. 2009;18(6):S22, S24, S26, passim.
- 35. Chang, E.I. & Liu, J. (2017). Prospective unbiased experience with three acellular dermal matrices in breast reconstruction. *J Surg Oncol*, *116*(3):365-370. doi: 10.1002/jso.24656.
- 36. Chalmers RT, Darling Iii RC, Wingard JT, et al. Randomized clinical trial of tranexamic acid-free fibrin sealant during vascular surgical procedures. Br J Surg. 2010;97(12):1784-1789.
- 37. Chauviere MV, Schutter RJ, Steigelman MB, et al. Comparison of AlloDerm and AlloMax tissue incorporation in rats. Ann Plast Surg 2014;73(3):282-285.
- 38. Chavarriaga LF, Lin E, Losken A, et al. Management of complex abdominal wall defects using acellular porcine dermal collagen. Am Surg. 2010;76(1):96-100.
- 39. Chen H, Cheng Y, Tian J, Yang P, Zhang X, Chen Y, Hu Y, Wu J. Dissolved oxygen from microalgae-gel patch promotes chronic wound healing in diabetes. Sci Adv. 2020 May 15;6(20):eaba4311.
- 40. Cheng AW, Abbas MA, Tejirian T. Outcome of abdominal wall hernia repair with biologic mesh: Permacol™ versus Strattice™. Am Surg. 2014;80(10):999-1002.
- 41. Chopra S, Al-Ishaq Z, Vidya R. The Journey of Prepectoral Breast Reconstruction through Time. World J Plast Surg. 2021;10(2):3-13. doi:10.29252/wjps.10.2.3
- 42. Cole PD, Stal D, Sharabi SE, et al. A comparative, long-term assessment of four soft tissue substitutes. Aesthet Surg J. 2011;31(6):674-681.
- 43. Connolly RJ. Evaluation of a unique bovine collagen matrix for soft tissue repair and reinforcement. Int Urogynecol J Pelvic Floor Dysfunct. 2006;17(Suppl 1):S44-S47.
- 44. Cooke M, Tan EK, Mandrycky C, et al. Comparison of cryopreserved amniotic membrane and umbilical cord tissue with dehydrated amniotic membrane/chorion tissue. J Wound Care. 2014;23(10):465-474,
- 45. Costa A, Adamo S, Gossetti F, D'Amore L, Ceci F, Negro P, Bruzzone P. Biological Scaffolds for Abdominal Wall Repair: Future in Clinical Application? Materials (Basel). 2019 Jul 25;12(15):2375.
- 46. Cottler PS, Kang H, Nash V, Salopek L, Bruce AC, Spiller KL, Campbell CA. Immunomodulation of Acellular Dermal Matrix Through Interleukin 4 Enhances Vascular Infiltration. Ann Plast Surg. 2022 Jun 1;88(5 Suppl 5):S466-S472.

- 47. Dahlgren E, Kjølhede P.; RPOP-PELVICOL Study Group. Long-term outcome of porcine skin graft in surgical treatment of recurrent pelvic organ prolapse. An open randomized controlled multicenter study. Acta Obstet Gynecol Scand. 2011; 90(12):1393-1401.
- 48. DeNoto G, 3rd, Ceppa EP, Pacella SJ, et al. 24-month results of the BRAVO study: A prospective, multi-center study evaluating the clinical outcomes of a ventral hernia cohort treated with OviTex® 1S permanent reinforced tissue matrix. Ann Med Surg (Lond). 2022;83:104745.
- 49. Dikmans RE, El Morabit F, Ottenhof MJ, et al. Single-stage breast reconstruction using Strattice™: a retrospective study. J Plast Reconstr Aesthet Surg. 2016; 69(2):227-233.
- 50. Driver VR, Lavery LA, Reyzelman AM, et al. A clinical trial of Integra Template for diabetic foot ulcer treatment. Wound Repair Regen. 2015;23(6):891-900.
- 51. Duisit J, Maistriaux L, Taddeo A, Orlando G, Joris V, Coche E, Behets C, Lerut J, Dessy C, Cossu G, Vögelin E, Rieben R, Gianello P, Lengelé B. Bioengineering a Human Face Graft: The Matrix of Identity. Ann Surg. 2017 Nov;266(5):754-764.
- 52. Edmonds ME, Foster AV, McColgan M. 'Dermagraft': A new treatment for diabetic foot ulcers. Diabet Med 1997;14:1010-1011.
- 53. Egozi D, Allwies TM, Fishel R, Jacobi E, Lemberger M. Free Nipple Grafting and Nipple Sharing in Autologous Breast Reconstruction after Mastectomy. Plast Reconstr Surg Glob Open. 2020 Sep 28;8(9):e3138.
- 54. Elassal AA, Al-Radi OO, Zaher ZF, Dohain AM, Abdelmohsen GA, Mohamed RS, Fatani MA, Abdelmotaleb ME, Noaman NA, Elmeligy MA, Eldib OS. Equine pericardium: a versatile alternative reconstructive material in congenital cardiac surgery. J Cardiothorac Surg. 2021 Apr 23;16(1):110.
- 55. Epstein NE. Dural repair with four spinal sealants: Focused review of the manufacturers' inserts and the current literature. Spine J. 2010;10(12):1065-1068.
- 56. Espinosa-de-Los-Monteros A, Avendano-Peza H, Novitsky YW. Abdominal Closure after TRAM Flap Breast Reconstruction with Transversus Abdominis Muscle Release and Mesh. Plast Reconstr Surg Glob Open. 2016 Sep 21;4(9):
- 57. Everts PA, Knape JT, Weibrich G, et al. Platelet-rich plasma and platelet gel: A review. J Extra Corpor Technol. 2006;38(2):174-187.
- 58. Faga A, Nicoletti G, Brenta F, et al. Hyaluronic acid three-dimensional scaffold for surgical revision of retracting scars: A human experimental study. Int Wound J. 2013;10(3):329-335.
- 59. Finger PT, Jain P, Mukkamala SK. Super-Thick Amniotic Membrane Graft for Ocular Surface Reconstruction. Am J Ophthalmol. 2019 Feb;198:45-53.
- 60. Franceschini G, Masetti R. Acellular dermal matrix as filler in breast-conserving surgery: warnings for a careful use. World J Surg Oncol. 2021 Jan 2;19(1):1.
- 61. Forbes TA, Shaw L, Quinlan C. Topical Honey in the Management of Pediatric Peritoneal Dialysis Exit Sites. Perit Dial Int. 2016;36(6):684-687. doi:10.3747/pdi.2014.00350
- 62. Frykberg RG. Topical Wound Oxygen Therapy in the Treatment of Chronic Diabetic Foot Ulcers. Medicina (Kaunas). 2021 Aug 31;57(9):917.

- 63. Frykberg RG, Cazzell SM, Arroyo-Rivera J, et al. Evaluation of tissue engineering products for the management of neuropathic diabetic foot ulcers: An interim analysis. J Wound Care. 2016;25 Suppl 7:S18-S25.
- 64. Frykberg RG, Gibbons GW, Walters JL, et al. A prospective, multicentre, open-label, single-arm clinical trial for treatment of chronic complex diabetic foot wounds with exposed tendon and/or bone: Positive clinical outcomes of viable cryopreserved human placental membrane. Int Wound J. 2016 Aug 3.
- 65. Frykberg RG, Marston WA, Cardinal M. The incidence of lower-extremity amputation and bone resection in diabetic foot ulcer patients treated with a human fibroblast-derived dermal substitute. Adv Skin Wound Care. 2015;28(1):17-20.
- 66. Galati V, Vonthein R, Stang F, Mailaender P, Kisch T. Split thickness skin graft versus application of the temporary skin substitute suprathel in the treatment of deep dermal hand burns: a retrospective cohort study of scar elasticity and perfusion. Int J Burns Trauma. 2021 Aug 15;11(4):312-320.
- 67. Gerdisch MW, Shea RJ, Barron MD. Clinical experience with CorMatrix extracellular matrix in the surgical treatment of mitral valve disease. J Thorac Cardiovasc Surg. 2014;148(4):1370-1378.
- 68. Gethin G, Cowman S, Kolbach DN. Debridement for venous leg ulcers. Cochrane Database Syst Rev. 2015 Sep 14;2015(9):CD008599.
- 69. Giordano P, Pullan RD, Ystgaard B, et al. The use of an acellular porcine dermal collagen implant in the repair of complex abdominal wall defects: A European multicentre retrospective study. Tech Coloproctol. 2015;19(7):411-417.
- 70. Giordano P, Sileri P, Buntzen S, et al. A prospective multicentre observational study of Permacol collagen paste for anorectal fistula: Preliminary results. Colorectal Dis. 2016;18(3):286-294.
- 71. Glat, P., Orgill, D.P., Galiano, R., et al. (August 2019). Placental Membrane Provides Improved Healing Efficacy and Lower Cost Versus a Tissue-Engineered Human Skin in the Treatment of Diabetic Foot Ulcerations. *Plastic and Reconstructive Surgery Global Open*, 7(8), e2371. doi: 10.1097/GOX.0000000000002371
- 72. Gould LJ. Topical Collagen-Based Biomaterials for Chronic Wounds: Rationale and Clinical Application. Adv Wound Care (New Rochelle). 2016 Jan 1;5(1):19-31.
- 73. Grassi A, Lucidi GA, Filardo G, Agostinone P, Macchiarola L, Bulgheroni P, Bulgheroni E, Zaffagnini S. Minimum 10-Year Clinical Outcome of Lateral Collagen Meniscal Implants for the Replacement of Partial Lateral Meniscal Defects: Further Results From a Prospective Multicenter Study. Orthop J Sports Med. 2021 May 25;9(5):2325967121994919.
- 74. Greenwood JE. A randomized, prospective study of the treatment of superficial partial-thickness burns: AWBAT-S versus Biobrane. Eplasty. 2011;11:e10.
- 75. Greer N, Foman NA, MacDonald R, et al. Advanced wound care therapies for nonhealing diabetic, venous, and arterial ulcers: A systematic review. Ann Intern Med. 2013;159(8):532-542.
- 76. Gossetti F, Zuegel N, Giordano P, Pullan R, Schuld J, Delrio P, Montorsi M, van Kerschaver O, Lemaitre J, Griffiths B, D'Amore L. A Biologic Surgical Implant in Complex Abdominal Wall Repair: 3-Year FollowUp Results of a Multicentric Prospective Study. Med Devices (Auckl). 2021 Aug 25;14:257-264. Doi: 10.2147/MDER.S297897. PMID: 34471389; PMCID: PMC8403569.

- 77. Guest JF, Weidlich D, Singh H, La Fontaine J, Garrett A, Abularrage CJ, Waycaster CR. Cost-effectiveness of using adjunctive porcine small intestine submucosa tri-layer matrix compared with standard care in managing diabetic foot ulcers in the US. J Wound Care. 2017 Jan 2;26(Sup1):S12-S24.
- 78. Guo Y, Chen G, Tian G, Tapia C. Sensory recovery following decellularized nerve allograft transplantation for digital nerve repair. J Plast Surg Hand Surg. 2013;47(6):451-453.
- 79. Gupta AK, Hug K, Berkoff DJ, et al. Dermal tissue allograft for the repair of massive irreparable rotator cuff tears. Am J Sports Med. 2012; 40(1):141-147.
- 80. Gurtner GC, Garcia AD, Bakewell K, Alarcon JB. A retrospective matched-cohort study of 3994 lower extremity wounds of multiple etiologies across 644 institutions comparing a bioactive human skin allograft, TheraSkin, plus standard of care, to standard of care alone. Int Wound J. 2020 Feb;17(1):55-64.
- 81. Haller HL, Blome-Eberwein SE, Branski LK, Carson JS, Crombie RE, Hickerson WL, Kamolz LP, King BT, Nischwitz SP, Popp D, Shupp JW, Wolf SE. Porcine Xenograft and Epidermal Fully Synthetic Skin Substitutes in the Treatment of Partial-Thickness Burns: A Literature Review. Medicina (Kaunas). 2021 Apr 30;57(5):432.
- 82. Hamilton NA, Porembka MR, Johnston FM, Gao F, Strasberg SM, Linehan DC, Hawkins WG. Mesh reinforcement of pancreatic transection decreases incidence of pancreatic occlusion failure for left pancreatectomy: a single-blinded, randomized controlled trial. Ann Surg. 2012 Jun;255(6):1037-42.
- 83. Han HH, Jun D, Moon SH, Kang IS, Kim MC. Fixation of split-thickness skin graft using fast-clotting fibrin glue containing undiluted high-concentration thrombin or sutures: a comparison study. Springerplus. 2016 Nov 2;5(1):1902.
- 84. Hankin CS, Knispel J, Lopes M, et al. Clinical and cost efficacy of advanced wound care matrices for venous ulcers. J Manag Care Pharm. 2012;18(5):375-384.
- 85. Harth KC, Rosen MJ. Major complications associated with xenograft biologic mesh implantation in abdominal wall reconstruction. Surg Innov. 2009;16(4):324-329.
- 86. Hayes, Inc. 3D Bioprinted Skin Substitutes for Treatment of Burns. Search & Summary. Prepared by Hayes, Inc. Publication Date: May 13, 2019.
- 87. Hayes, Inc., Evidence Analysis Research Brief. bio-ConneKt Wound Matrix (MLM Biologics Inc.) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., Jan 20, 2023.
- 88. Hayes, Inc., Evidence Analysis Research Brief. InnovaMatrix AC (Triad Life Sciences Inc.) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., Jan 20, 2023.
- 89. Hayes, Inc., Evidence Analysis Research Brief. NovoSorb (PolyNovo Pty. Ltd.) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., Feb 13, 2023.
- 90. Hayes, Inc., Evidence Analysis Research Brief. Microlyte Matrix (Imbed Biosciences) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., March 20, 2023.
- 91. Hayes, Inc., Evidence Analysis Research Brief. SurgiMend (Integra Life Sciences) for Postmastectomy Breast Reconstruction. Lansdale, PA: Hayes, Inc., March 24, 2023.
- 92. Hayes, Inc., Evidence Analysis Research Brief. Symphony (Aroa Biosurgery Ltd.) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., March 21, 2023.

- 93. Hayes, Inc. Evidence Analysis Research Brief. TheraGenesis (Bioventus LLC) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., Mar 21, 2023.
- 94. Hayes, Inc., Evidence Analysis Research Brief. Vim Amniotic Membrane (Cook Biotech) for Treatment of Surgical and Nonsurgical Wounds. Lansdale, PA: Hayes, Inc., Feb 13, 2023.
- 95. Hayes, Inc. Evidence Analysis Research Brief. XCelliStem Wound Powder (Stemsys) for Treatment of Wounds. Lansdale, PA: Hayes, Inc., Jan 19, 2023.
- 96. Hayes Inc., Evidence Analysis Research Brief. *Stravix and Stravix PL (Osiris Therapeutics) for the Treatment of Nonhealing Wounds.* Lansdale, PA: Hayes, Inc.,: September 2019.
- 97. Hayes Inc., Evolving Evidence Review. *Kerecis Omega3 Wound (Kerecis Limited) for the Management of Chronic Lower Extremity Wounds.* Lansdale, PA: Hayes, Inc.,: Feb 2022.
- 98. Hayes, Inc. Evolving Evidence Review. PuraPly AM Antimicrobial Wound Matrix (Organogenesis) for Treatment of Wounds. Lansdale, PA: Hayes, Inc.,: Dec 12, 2022
- 99. Hayes, Inc. Health Technology Assessment. Amniotic Allografts for Tendon and Ligament Injuries. Lansdale, PA: Hayes, Inc.,: Sept 2022.
- 100. Hayes, Inc. Health Technology Assessment. Human Amniotic Membrane Injections for Treatment of Chronic Plantar Fasciitis. Lansdale, PA: Hayes, Inc.,: Dec 27, 2022.
- 101. Hayes, Inc. Health Technology Assessment. Comparative Effectiveness Review Of Human Acellular Dermal Matrix For Breast Reconstruction. Lansdale, PA: Hayes, Inc.,: Feb 28, 2022
- 102. Hayes, Inc. Health Technology Assessment. Processed Nerve Allografts with the Avance Nerve Graft (Axogen Corporation) for Peripheral Nerve Discontinuities. Lansdale, PA: Hayes, Inc.,: May 18, 2023.
- 103. HerniaSurge Group. International guidelines for groin hernia management. Hernia. 2018 Feb;22(1):1-165. doi: 10.1007/s10029-017-1668-x. Epub 2018 Jan 12.
- 104. Ho G, Nguyen TJ, Shahabi A, et al. A systematic review and meta-analysis of complications associated with acellular dermal matrix-assisted breast reconstruction. Ann Plast Surg. 2012; 68(4):346-356.
- 105. Hoogewerf CJ, Hop MJ, Nieuwenhuis MK, Oen IM, Middelkoop E, Van Baar ME. Topical treatment for facial burns. Cochrane Database Syst Rev. 2020 Jul 29;7(7):CD008058.
- 106. Huang W, Chen Y, Wang N, Yin G, Wei C, Xu W. Effectiveness and safety of human amnion/chorion membrane therapy for diabetic foot ulcers: An updated meta-analysis of randomized clinical trials. Wound Repair Regen. 2020 Nov;28(6):739-750. doi: 10.1111/wrr.12851. Epub 2020 Aug 11.
- 107. Hyland EJ, D'Cruz R, Menon S, Harvey JG, La Hei E, Lawrence T, Waddell K, Nash M, Holland AJ. Biobrane™ versus acticoat™ for the treatment of mid-dermal pediatric burns: a prospective randomized controlled pilot study. Int J Burns Trauma. 2018 Jun 20;8(3):63-67.
- 108. Hicks KE, Huynh MN, Jeschke M, Malic C. Dermal regenerative matrix use in burn patients: A systematic review. J Plast Reconstr Aesthet Surg. 2019 Nov;72(11):1741-1751. doi: 10.1016/j.bjps.2019.07.021. Epub 2019 Aug 8.
- 109. Ibrahim AM, Shuster M, Koolen PG, et al. Analysis of the National Surgical Quality Improvement Program database in 19,100 patients undergoing implant-based breast

- reconstruction: complication rates with acellular dermal matrix. Plast Reconstr Surg. 2013; 132(5):1057-1066.
- 110. James, WD, Elston D, Treat JR, et al. *Andrews' Diseases of the Skin, 13th Edition.* USA: Elsevier; 2019.
- 111. Janfaza M, Martin M, Skinner R. A preliminary comparison study of two noncrosslinked biologic meshes used in complex ventral hernia repairs. World J Surg. 2012;36(8):1760-1764.
- 112. Jaseem M, Alungal S, Dhiyaneswaran, Shamsudeen J. Effectiveness of autologous PRP therapy in chronic nonhealing ulcer: A 2-year retrospective descriptive study. J Family Med Prim Care. 2020 Jun 30;9(6):2818-2822.
- 113. Johnston M, McBride M, Dahiya D, Owusu-Apenten R, Nigam PS. Antibacterial activity of Manuka honey and its components: An overview. AIMS Microbiol. 2018 Nov 27;4(4):655-664
- 114. Kalaiselvan R, Carlson GL, Hayes S, et al. Recurrent intestinal fistulation after porcine acellular dermal matrix reinforcement in enteric fistula takedown and simultaneous abdominal wall reconstruction. Hernia. 2020: 24(3):537-543.
- 115. Kashimura T, Nagasaki K, Horigome M, Yoshida K, Soejima K. Selection of Artificial Dermis for Shortening Treatment Period: Integra versus Pelnac. Plast Reconstr Surg Glob Open. 2021 Jun 10;9(6):e3599.
- 116. Karr JC, Taddei AR, Picchietti S, et al. A morphological and biochemical analysis comparative study of the collagen products Biopad, Promogram, Puracol, and Colactive. Adv Skin Wound Care. 2011;24(5):208-216.
- 117. Kavros S, Dutra T, Gonzalez-Cruz R, et al. The use of PriMatrix, a fetal bovine acellular dermal matrix, in healing chronic diabetic foot ulcers: A prospective multicenter study. Adv Skin Wound Care. 2014;27(8):356-362.
- 118. Kavros SJ. Acellular fetal bovine dermal matrix for treatment of chronic ulcerations of the midfoot associated with Charcot neuroarthropathy. Foot Ankle Spec. 2012; 5(4):230-234.
- 119. Khan MA, Javed AA, Rao DJ, Corner JA, Rosenfield P. Pediatric Traumatic Limb Amputation: The Principles of Management and Optimal Residual Limb Lengths. World J Plast Surg. 2016 Jan;5(1):7-14.
- 120. Khan ZA, Nambiar A, Morley R, et al. Long-term follow-up of a multicentre randomised controlled trial comparing tension-free vaginal tape, xenograft and autologous fascial slings for the treatment of stress urinary incontinence in women. BJU Int. 2015; 115(6):968-977.
- 121. Kim JH, Kim DH, Lee YP. Long-term comparison of physiologic anorectal changes and recurrence between transanal repair and transanal repair with posterior colporrhaphy in rectocele. Asian J Surg. 2020 Jan;43(1):265-271.
- 122. Kim JY, Davila AA, Persing S, et al. A meta-analysis of human acellular dermis and submuscular tissue expander breast reconstruction. Plast Reconstr Surg. 2012; 129(1):28-41.
- 123. Kirsner RS, Sabolinski ML, Parsons NB, et al. Comparative effectiveness of a bioengineered living cellular construct vs. a dehydrated human amniotic membrane allograft for the treatment of diabetic foot ulcers in a real world setting. *Wound Repair and Regeneration*. 2015; 23(5): p. 737-744. Doi: https://doi.org/10.1111/wrr.12332

- 124. Kirsner RS, Bohn G, Driver VR, et al. Human acellular dermal wound matrix: Evidence and experience. Int Wound J. 2015;12(6):646-654.
- 125. Kirsner RS, Warriner R, Michela M, et al. Advanced biological therapies for diabetic foot ulcers. Arch Dermatol. 2010;146(8):857-862.
- 126. Kogan S, Sood A, Granick MS. Amniotic Membrane Adjuncts and Clinical Applications in Wound Healing: A Review of the Literature. Wounds. 2018 Jun;30(6):168-173.
- 127. Krezdorn N, Könneker S, Paprottka FJ, et al. Biobrane versus topical agents in the treatment of adult scald burns. Burns. 2016 Aug 6.
- 128. Kulig KM, Luo X, Finkelstein EB, et al. Biologic properties of surgical scaffold materials derived from dermal ECM. Biomaterials. 2013;34(23):5776-5784.
- 129. Kumar, N.G., Berlin, N.L., Kim, H.M., et al. (Jan 2021). Development of an evidence-based approach to the use of acellular dermal matrix in immediate expander-implant-based breast reconstruction. *J Plast Reconstr Aesthet Surg*, 74(1), 30-40. doi: 10.1016/j.bjps.2020.10.005.
- 130. Lakmal K, Basnayake O, Hettiarachchi D. Systematic review on the rational use of amniotic membrane allografts in diabetic foot ulcer treatment. BMC Surg. 2021 Feb 15;21(1):87.
- 131. Lagus H, Sarlomo-Rikala M, Böhling T, Vuola J. Prospective study on burns treated with Integra®, a cellulose sponge and split thickness skin graft: Comparative clinical and histological study--randomized controlled trial. Burns. 2013;39(8):1577-1587.
- 132. Laimer, M., Bauer, J., & Murrell, D.F. (Jan 2021). *Epidermolysis bullosa: Epidemiology, pathogenesis, classification, and clinical features.* UpToDate.com. https://www.uptodate.com/contents/epidermolysis-bullosa-epidemiology-pathogenesis-classific ation-and-clinical-features?search=Epidermolysis%20Bullosa&source=search_result&selectedTitle=1~73&usage_type=default&display_rank=1#H2900707
- 133. Lakmal K, Basnayake O, Hettiarachchi D. Systematic review on the rational use of amniotic membrane allografts in diabetic foot ulcer treatment. BMC Surg. 2021 Feb 15;21(1):87.
- 134. Landsman A, Rosines E, Houck A, et al. Characterization of a cryopreserved split-thickness human skin allograft TheraSkin. Adv Skin Wound Care. 2016;29(9):399-406.
- 135. Lavery LA, Fulmer J, Shebetka KA, et al.; Grafix Diabetic Foot Ulcer Study Group. The efficacy and safety of Grafix(®) for the treatment of chronic diabetic foot ulcers: Results of a multi-centre, controlled, randomised, blinded, clinical trial. Int Wound J. 2014;11(5):554-560.
- 136. Lim JZ, Ng NS, Thomas C. Prevention and treatment of diabetic foot ulcers. J R Soc Med. 2017;110(3):104-109. Doi: 10.1177/0141076816688346.
- 137. Lipsky BA, Berendt AR, Cornia PB, et al.; Infectious Diseases Society of America. 2012 Infectious Diseases Society of America clinical practice guideline for the diagnosis and treatment of diabetic foot infections. Clin Infect Dis. 2012;54:e132-e173.
- 138. Liu DZ, Mathes DW, Neligan PC, et al. Comparison of outcomes using AlloDerm versus FlexHD for implant-based breast reconstruction. Ann Plast Surg. 2014; 72(5):503-507.
- 139. Lineaweaver W, Bush K, James K. Suppression of α Smooth Muscle Actin Accumulation by Bovine Fetal Dermal Collagen Matrix in Full Thickness Skin Wounds. Ann Plast Surg. 2015 Jun;74 Suppl 4(Suppl 4):S255-8.

- 140. Lullove E. Acellular fetal bovine dermal matrix in the treatment of nonhealing wounds in patients with complex comorbidities. J Am Podiatr Med Assoc. 2012;102(3):233-239.
- 141. Martinez-Zapata MJ, Martí-Carvajal AJ, Solà I, et al. Autologous platelet-rich plasma for treating chronic wounds. Cochrane Database Syst Rev. 2012;(10):CD006899.
- 142. Martinson M, Martinson N. A comparative analysis of skin substitutes used in the management of diabetic foot ulcers. J Wound Care. 2016;25(Sup10):S8-S17.
- 143. Massee M, Chinn K, Lei J, et al. Dehydrated human amnion/chorion membrane regulates stem cell activity in vitro. J Biomed Mater Res B Appl Biomater. 2016;104(7):1495-1503.
- 144. Mayer, T & Glat, P.M. The Use of Dehydrated Human Amnion/Chorion Membranes in the Treatment of Burns and Complex Wounds. Annals of Plastic Surgery. 2017; 78 (2):pS11-S13. doi: 10.1097/SAP.000000000000983
- 145. Mazari FAK, Wattoo GM, Kazzazi NH, et al. The comparison of Strattice and SurgiMend in acellular dermal matrix-assisted, implant-based immediate breast reconstruction. Plast Reconstr Surg. 2018; 141(2):283-293.
- 146. Mendenhall SD, Anderson LA, Ying J, et al. The BREASTrial: stage I. Outcomes from the time of tissue expander and acellular dermal matrix placement to definitive reconstruction. Plast Reconstr Surg. 2015; 135(1):29e-42e.
- 147. Mendenhall SD, Anderson LA, Ying J, Boucher KM, Neumayer LA, Agarwal JP. The BREASTrial Stage II: ADM Breast Reconstruction Outcomes from Definitive Reconstruction to 3 Months Postoperative. Plast Reconstr Surg Glob Open. 2017 Jan 25;5(1):e1209.
- 148. Michelotti BF, Brooke S, Mesa J, et al. Analysis of clinically significant seroma formation in breast reconstruction using acellular dermal grafts. Ann Plast Surg. 2013; 71(3):274-177.
- 149. Min JH, Yun IS, Lew DH, et al. The use of matriderm and autologous skin graft in the treatment of full thickness skin defects. Arch Plast Surg. 2014;41(4):330-336.
- 150. Mohebbi A, Hosseinzadeh F, Mohebbi S, Dehghani A. Determining the effect of platelet-rich plasma (PRP) on improving endoscopic sinus surgery: A randomized clinical trial study (RCT). Med J Islam Repub Iran. 2019 Dec 25;33:150.
- 151. Moravvej H, Hormozi AK, Hosseini SN, et al. Comparison of the application of allogeneic fibroblast and autologous mesh grafting with the conventional method in the treatment of third-degree burns. J Burn Care Res. 2016;37(1):e90-e95.
- 152. Motolese A, Vignati F, Brambilla R, et al. Interaction between a regenerative matrix and wound bed in nonhealing ulcers: Results with 16 cases. Biomed Res Int. 2013;2013:849321.
- 153. Nahabedian MY. Secondary nipple reconstruction using local flaps and AlloDerm. Plast Reconstr Surg. 2005; 115(7):2056-2061.
- 154. Narang SK, Alam NN, Köckerling F, Daniels IR, Smart NJ. Repair of Perineal Hernia Following Abdominoperineal Excision with Biological Mesh: A Systematic Review. Front Surg. 2016 Sep 5:3:49.
- 155. Nasso G, Di Bari N, Moscarelli M, Fiore F, Condello I, Santarpino G, Speziale G. A modified technique for aortic prosthesis implantation after prosthetic valve endocarditis complicated by complex paraannular aortic abscess. Rev Cardiovasc Med. 2021 Dec 22;22(4):1621-1627.

- 156. National Institute for Health and Care Excellence. *Diabetic foot problems: prevention and management. NICE guideline [NG19].* Published: 26 August 2015. Last updated: 11 October 2019. Retrieved from
 - https://www.nice.org.uk/guidance/ng19/chapter/Recommendations#assessing-the-risk-of-developing-a-diabetic-foot-problem
- 157. National Institute for Health and Care Excellence. *Peripheral arterial disease: diagnosis and management. Clinical guideline [CG147].* Published: August 8, 2012. Last updated: December 11, 2020. Retrieved from
 - https://www.nice.org.uk/guidance/cg147/chapter/Recommendations#diagnosis
- 158. Noël J, Mascarenhas A, Patel E, Reddy S, Sandri M, Bhat S, Moschovas M, Rogers T, Ahmed S, Stirt D, Patel V. Nerve spare robot assisted laparoscopic prostatectomy with amniotic membranes: medium term outcomes. J Robot Surg. 2022 Oct;16(5):1219-1224. doi: 10.1007/s11701-022-01370-4. Epub 2022 Jan 11.
- 159. Noordenbos J, Doré C, Hansbrough JF. Safety and efficacy of TransCyte for the treatment of partial-thickness burns. J Burn Care Rehabil. 1999; 20(4):275-281.
- 160. O'Donnell TF Jr, Lau J. A systematic review of randomized controlled trials of wound dressings for chronic venous ulcer. J Vasc Surg. 2006;44(5):1118-1125.
- 161. Ogaya-Pinies G, Palayapalam-Ganapathi H, Rogers T, Hernandez-Cardona E, Rocco B, Coelho RF, Jenson C, Patel VR. Can dehydrated human amnion/chorion membrane accelerate the return to potency after a nerve-sparing robotic-assisted radical prostatectomy? Propensity score-matched analysis. J Robot Surg. 2018 Jun;12(2):235-243. doi: 10.1007/s11701-017-0719-8. Epub 2017 Jun 27. PMID: 28656504.
- 162. Ontario Health (Quality). Skin Substitutes for Adults With Diabetic Foot Ulcers and Venous Leg Ulcers: A Health Technology Assessment. Ont Health Technol Assess Ser. 2021 Jun 4;21(7):1-165.
- 163. Osorio CC, Escobar LM, González MC, Gamboa LF, Chambrone L. Evaluation of density, volume, height and rate of bone resorption of substitutes of autologous bone grafts for the repair of alveolar clefts in humans: A systematic review. Heliyon. 2020 Sep 4;6(9):e04646.
- 164. Osugi I, Inagawa K, Ebisudani S, Hara N. Usefulness of a Skin Graft Obtained from the Bilateral Nasolabial Folds for a Skin Defect following Resection of a Malignant Tumor at the Nasal Tip. Plast Reconstr Surg Glob Open. 2021 Mar 26;9(3):e3481.
- 165. Palaia DA, Arthur KS, Cahan AC, Rosenberg MH. Incidence of Seromas and Infections Using Fenestrated versus Nonfenestrated Acellular Dermal Matrix in Breast Reconstructions. Plast Reconstr Surg Glob Open. 2015; 3(11):e569.
- 166. Patel KM, Nahabedian MY, Gatti M, Bhanot P. Indications and outcomes following complex abdominal reconstruction with component separation combined with porcine acellular dermal matrix reinforcement. Ann Plast Surg. 2012 Oct;69(4):394-398.
- 167. Patel VR, Samavedi S, Bates AS, et al. Dehydrated human amnion/chorion membrane allograft nerve wrap around the prostatic neurovascular bundle accelerates early return to continence and potency following robot-assisted radical prostatectomy: propensity score-matched analysis. Eur Urol. 2015; 67(6):977-980.

- 168. Peled ZM. Treatment of a patient with small fiber pathology using nerve biopsy and grafting: A case report. J Reconstr Microsurg. 2013;29(8):551-554.
- 169. Pittman, T.A., Fan, K.L., Knapp, A., et al. (March 2017). Comparison of Different Acellular Dermal Matrices in Breast Reconstruction: The 50/50 Study. Plast Reconstr Surg,139(3), 521-528. doi: 10.1097/PRS.0000000000003048.
- 170. Piejko M, Radziun K, Bobis-Wozowicz S, Waligórska A, Zimoląg E, Nessler M, Chrapusta A, Madeja Z, Drukała J. Adipose-Derived Stromal Cells Seeded on Integra® Dermal Regeneration Template Improve Post-Burn Wound Reconstruction. Bioengineering (Basel). 2020 Jul 2;7(3):67.
- 171. Poppler LH, Mundschenk MB, Linkugel A, Zubovic E, Dolen UC, Myckatyn TM. Tissue Expander Complications Do Not Preclude a Second Successful Implant-Based Breast Reconstruction. Plast Reconstr Surg. 2019 Jan;143(1):24-34.
- 172. Quarti A, Nardone S, Colaneri M, et al. Preliminary experience in the use of an extracellular matrix to repair congenital heart diseases. Interact Cardiovasc Thorac Surg. 2011; 13(6):569-572
- 173. Reilly, DA, Hickey S, Glat P, et al. Using Dehydrated Human Amnion/Chorion Membrane Allografts for Acute and Reconstructive Burn Care. Annals of Plastic Surgery. 2017; 78(1):S19-S26. DOI: 10.1097/SAP.0000000000000981
- 174. Regulski M, Jacobstein DA, Petranto RD, et al. A retrospective analysis of a human cellular repair matrix for the treatment of chronic wounds. Ostomy Wound Manage. 2013; 59(12):38-43.
- 175. Reyzelman AM, Bazarov I. Human acellular dermal wound matrix for treatment of DFU: Literature review and analysis. J Wound Care. 2015;24(3):128; 129-134.
- 176. Rice JB, Desai U, Ristovska L, et al. Economic outcomes among Medicare patients receiving bioengineered cellular technologies for treatment of diabetic foot ulcers. J Med Econ. 2015;18(8):586-595.
- 177. Robb GL, Gurtner GC. Letter to the editor. Healing rates in a multicenter assessment of a sterile, room temperature, acellular dermal matrix versus conventional care wound management and an active comparator in the treatment of full-thickness diabetic foot ulcers. ePlasty. 2016;16:229.
- 178. Romain, B., Story, F., Meyer, N., et al. (Jun 2016). Comparative study between biologic porcine dermal meshes: risk factors of postoperative morbidity and recurrence. *J Wound Care, 25*(6), 320-5. doi: 10.12968/jowc.2016.25.6.320.
- 179. Roshangar L, Soleimani Rad J, Kheirjou R, Reza Ranjkesh M, Ferdowsi Khosroshahi A. Skin Burns: Review of Molecular Mechanisms and Therapeutic Approaches. Wounds. 2019 Dec;31(12):308-315.
- 180. Roth JS, Brathwaite C, Hacker K, et al. Complex ventral hernia repair with a human acellular dermal matrix. Hernia.2015;19(2):247-252.
- 181. Royal Biologics. Amnio-Maxx™. 2022. Accessed May 18, 2023. Available at URL address: https://royalbiologics.com/amnio-maxx-dual-layer-patch
- 182. Rundell VL, Beck RT, Wang CE, et al. Complication prevalence following use of tutoplast-derived human acellular dermal matrix in prosthetic breast reconstruction: a retrospective review of 203 patients. J Plast Reconstr Aesthet Surg. 2014; 67(10):1345-1351.

- 183. Sanders L, Landsman AS, Landsman A, et al. A prospective, multicenter, randomized, controlled clinical trial comparing a bioengineered skin substitute to a human skin allograft. Ostomy Wound Manage. 2014; 60(9):26-38.
- 184. Santema TB, Poyck PP, Ubbink DT. Skin grafting and tissue replacement for treating foot ulcers in people with diabetes. Cochrane Database Syst Rev. 2016 Feb 11;2(2):CD011255.
- 185. Serena TE, Carter MJ, Le LT, et al.; EpiFix VLU Study Group. A multi-center randomized controlled clinical trial evaluating the use of dehydrated human amnion/chorion membrane allografts and multilayer compression therapy vs. multilayer compression therapy alone in the treatment of venous leg ulcers. Wound Repair Regen. 2014; 22(6):688-693
- 186. Selvarajah D, Bollu BK, Harvey J, Jacques M, Jehangir S, Fuller ES, La Hei E, Bertinetti M, Lawrence T, Holland AJ. Acticoat versus biobrane: a retrospective review on the treatment of paediatric mid-dermal torso burns. Int J Burns Trauma. 2019 Aug 15;9(4):82-87.
- 187. Selvarajah D, Bollu BK, Harvey J, Jacques M, Jehangir S, Fuller ES, La Hei E, Bertinetti M, Lawrence T, Holland AJ. Acticoat versus biobrane: a retrospective review on the treatment of paediatric mid-dermal torso burns. Int J Burns Trauma. 2019 Aug 15;9(4):82-87.
- 188. Shah BC, Tiwari MM, Goede MR, et al. Not all biologics are equal! Hernia. 2011;15(2):165-171.
- 189. Shah SS, Todkar JS, Shah PS. Buttressing the staple line: A randomized comparison between staple-line reinforcement versus no reinforcement during sleeve gastrectomy. Obes Surg. 2014;24(12):2014-2020.
- 190. Sheikh ES, Sheikh ES, Fetterolf DE. Use of dehydrated human amniotic membrane allografts to promote healing in patients with refractory non healing wounds. Int Wound J. 2014; 11(6):711-717.
- 191. Shikora SA, Mahoney CB. Clinical Benefit of Gastric Staple Line Reinforcement (SLR) in Gastrointestinal Surgery: a Meta-analysis. Obes Surg. 2015 Jul;25(7):1133-41.
- 192. Singh DP, Zahiri HR, Gastman B, et al. A modified approach to component separation using biologic graft as a load-sharing onlay reinforcement for the repair of complex ventral hernia. Surg Innov. 2014;21(2):137-146.
- 193. Skovgaard C, Holm B, Troelsen A, et al. No effect of fibrin sealant on drain output or functional recovery following simultaneous bilateral total knee arthroplasty: A randomized, double-blind, placebo-controlled study. Acta Orthop. 2013;84(2):153-158.
- 194. Sobti N, Liao EC. Surgeon-controlled study and meta-analysis comparing FlexHD and AlloDerm in immediate breast reconstruction outcomes. Plast Reconstr Surg. 2016;138(5):959-967.
- 195. Solli P, Brandolini J, Pardolesi A, Nardini M, Lacava N, Parri SF, Kawamukai K, Bonfanti B, Bertolaccini L. Diaphragmatic and pericardial reconstruction after surgery for malignant pleural mesothelioma. J Thorac Dis. 2018 Jan;10(Suppl 2):S298-S303.
- 196. Solomon MP, Komlo C, Defrain M. Allograft materials in phalloplasty: A comparative analysis. Ann Plast Surg. 2013;71(3):297.

- 197. Swan J. Use of cryopreserved, particulate human amniotic membrane and umbilical cord (AM/UC) tissue: A case series study for application in the healing of chronic wounds. Surg Technol Int. 2014;25:73-78.
- 198. Tettelbach W, Cazzell S, Reyzelman AM, et al. A confirmatory study on the efficacy of dehydrated human amnion/chorion membrane dHACM allograft in the management of diabetic foot ulcers: A prospective, multicentre, randomised, controlled study of 110 patients from 14 wound clinics. *International Wound Journal*. 2018; 16 (1): 19-29. Doi: https://doi.org/10.1111/iwj.12976
- 199. U.S. Food & Drug Administration. Acellular Dermal Matrix (ADM) Products Used in Implant-Based Breast Reconstruction Differ in Complication Rates: FDA Safety Communication. FDA.gov. Issued March 31, 2021. https://www.fda.gov/medical-devices/safety-communications/acellular-dermal-matrix-adm-products-used-implant-based-breast-reconstruction-differ-complication?utm_medium=email&utm_source=qovdelivery
- 200. Vaishya R, Agarwal AK, Tiwari M, Vaish A, Vijay V, Nigam Y. Medical textiles in orthopedics: An overview. J Clin Orthop Trauma. 2018 Mar;9(Suppl 1):S26-S33. doi: 10.1016/j.jcot.2017.10.016.
- 201. van Doormaal TPC, Germans MR, Sie M, Brouwers B, Fierstra J, Depauw PRAM, Robe PA, Regli L. Single-Arm, Open-Label, Multicenter Study to Evaluate the Safety and Performance of Dura Sealant Patch in Reducing Cerebrospinal Fluid Leakage Following Elective Cranial Surgery: The ENCASE Trial Study Protocol. Neurosurgery. 2020 Feb 1;86(2):E203-E208.
- 202. Venturi ML, Mesbahi AN, Boehmler JH 4th, Marrogi AJ. Evaluating sterile human acellular dermal matrix in immediate expander-based breast reconstruction: a multicenter, prospective, cohort study. Plast Reconstr Surg. 2013; 131(1):9e-18e.
- 203. Ventris Medical. Cellesta™. 2020. Accessed May 15, 2023. Available at URL address: https://www.ventrismedical.com/products/amniotic-tissue/
- 204. Veit-Rubin N, Cartwright R. Sacrocolpopexy tends to be superior to transvaginal mesh surgery. BJOG. 2021 Jan;128(1):24.
- 205. Vloemans AF, Hermans MH, van der Wal MB, et al. Optimal treatment of partial thickness burns in children: A systematic review. Burns. 2014;40(2):177-190.
- 206. Walters J, Cazzell S, Pham H, et al. Healing rates in a multicenter assessment of a sterile, room temperature, acellular dermal matrix versus conventional care wound management and an active comparator in the treatment of full-thickness diabetic foot ulcers. *Eplasty.* 2016;16(10).
- 207. Wang L, Yan X, Zhao J, Chen C, et al. Expert consensus on resection of chest wall tumors and chest wall reconstruction. Transl Lung Cancer Res. 2021 Nov;10(11):4057-4083.
- 208. Ward KC, Costello KP, Baalman S, et al. Effect of acellular human dermis buttress on laparoscopic hiatal hernia repair. Surg Endosc. 2015;29(8):2291-2297.
- 209. Wazir U, Mokbel K. The Evolving Role of Pre-pectoral ADM-assisted Approach in Implant-based Immediate Breast Reconstruction Following Conservative Mastectomy: An Overview of the Literature and Description of Technique. In Vivo. 2018 Nov-Dec;32(6):1477-1480.

- 210. Wen Z, Lu T, Wang Y, Liang H, Gao Z, He X. Anterior Cervical Corpectomy and Fusion and Anterior Cervical Discectomy and Fusion Using Titanium Mesh Cages for Treatment of Degenerative Cervical Pathologies: A Literature Review. Med Sci Monit. 2018 Sep 12;24:6398-6404.
- 211. Willett NJ, Thote T, Lin AS, et al. Intra-articular injection of micronized dehydrated human amnion/chorion membrane attenuates osteoarthritis development. Arthritis Res Ther. 2014;16(1):R47.
- 212. Williams ML, Holewinski JE. Use of a human acellular dermal wound matrix in patients with complex wounds and comorbidities. J Wound Care. 2015;24(6):261-262, 264-267.
- 213. Woo KY, Coutts PM, Sibbald RG. Continuous topical oxygen for the treatment of chronic wounds: A pilot study. Adv Skin Wound Care. 2012;25(12):543-547.
- 214. Xu R, Xie ME, Jackson CM. Trigeminal Neuralgia: Current Approaches and Emerging Interventions. J Pain Res. 2021 Nov 3;14:3437-3463.
- 215. Yanagawa B, Rao V, Yau TM, Cusimano RJ. Initial experience with intraventricular repair using CorMatrix extracellular matrix. Innovations (Phila). 2013;8(5):348-352.
- 216. Yanagawa B, Rao V, Yau TM, Cusimano RJ. Potential myocardial regeneration with CorMatrix ECM: A case report. J Thorac Cardiovasc Surg. 2014;147(4):e41-e43.
- 217. Yao M, Attalla K, Ren Y, et al. Ease of use, safety, and efficacy of integra bilayer wound matrix in the treatment of diabetic foot ulcers in an outpatient clinical setting: A prospective pilot study. J Am Podiatr Med Assoc. 2013;103(4):274-280.
- 218. Yeen WC, Faber C, Caldeira C, et al. Reconstruction of pulmonary venous conduit with CorMatrix in lung transplant. Asian Cardiovasc Thorac Ann. 2013;21(3):360-362.
- 219. Zelen, C.M., Orgill, D.P., Serena T.E., et al. (2018). An aseptically processed, acellular, reticular, allogenic human dermis improves healing in diabetic foot ulcers: A prospective, randomised, controlled, multicentre follow-up trial. *International Wound Journal*, 15(5), 731-739. doi: https://doi.org/10.1111/iwj.12920
- 220. Zelen CM, Serena TE, Gould L, Le L, Carter MJ, Keller J, Li WW. Treatment of chronic diabetic lower extremity ulcers with advanced therapies: a prospective, randomised, controlled, multi-centre comparative study examining clinical efficacy and cost. Int Wound J. 2016 Apr;13(2):272-82.
- 221. Zelen CM, Gould L, Serena TE, et al. A prospective, randomised, controlled, multi-centre comparative effectiveness study of healing using dehydrated human amnion/chorion membrane allograft, bioengineered skin substitute or standard of care for treatment of chronic lower extremity diabetic ulcers. Int Wound J. 2015;12(6):724-732.
- 222. Zelen CM, Poka A, Andrews J. Prospective, randomized, blinded, comparative study of injectable micronized dehydrated amniotic/chorionic membrane allograft for plantar fasciitis--a feasibility study. Foot Ankle Int. 2013;34(10):1332-1339.
- 223. Zelen CM, Serena TE, Snyder RJ. A prospective, randomised comparative study of weekly versus biweekly application of dehydrated human amnion/chorion membrane allograft in the management of diabetic foot ulcers. Int Wound J. 2014;11(2):122-128.

- 224. Zenn, M.R., & Salzberg, C.A., (Aug 2016). A Direct Comparison of Alloderm-Ready to Use (RTU) and DermACELL in Immediate Breast Implant Reconstruction. *Eplasty, 16*: e23.
- 225. Zhong T, Temple-Oberle C, Hofer S, et al.; MCCAT Study Group. The Multi Centre Canadian Acellular Dermal Matrix Trial (MCCAT): study protocol for a randomized controlled trial in implant-based breast reconstruction. Trials. 2013; 14:356.
- 226. Zhu T, Wang H, Jing Z, Fan D, Liu Z, Wang X, Tian Y. High efficacy of tetra-PEG hydrogel sealants for sutureless dural closure. Bioact Mater. 2021 Jun 27;8:12-19.
- 227. Zografakis J, Johnston G, Haas J, Berbiglia L, Bedford T, Spear J, Dan A, Pozsgay M. Urinary Bladder Matrix Reinforcement for Laparoscopic Hiatal Hernia Repair. JSLS. 2018

 Apr-Jun;22(2):e2017.00060.
- 228. Baldursson BT, et al. Healing rate and autoimmune safety of full-thickness wounds treated with fish skin acellular dermal matrix versus porcine small-intestine submucosa: a noninferiority study. Int J Low Extrem Wounds. 2015 Mar;14(1):37-43.
- 229. Yang CK, Polanco TO, Lantis JC. A Prospective, Postmarket, Compassionate Clinical Evaluation of a Novel Acellular Fish-skin Graft Which Contains Omega-3 Fatty Acids for the Closure of Hard-to-heal Lower Extremity Chronic Ulcers. Wounds. 2016 Apr;28(4):112-8.
- 230. Magnusson S, et al. Regenerative and Antibacterial Properties of Acellular Fish Skin Grafts and Human Amnion/Chorion Membrane: Implications for Tissue Preservation in Combat Casualty Care. Mil Med. 2017 Mar;182(S1):383-388.
- 231. Woodrow T, Chant T, Chant H. Treatment of diabetic foot wounds with acellular fish skin graft rich in omega-3: a prospective evaluation. J Wound Care. 2019 Feb 2;28(2):76-80.
- 232. Alam K, Jeffery SLA. Acellular Fish Skin Grafts for Management of Split Thickness Donor Sites and PartialThickness Burns: A Case Series. Mil Med. 2019 Mar 1;184(Supplement_1):16-20.
- 233. Chattopadhyay S, Raines RT. Review collagen-based biomaterials for wound healing. Biopolymers. 2014 Aug;101(8):821-33.
- 234. Hu MS, et al. Tissue engineering and regenerative repair in wound healing. Ann Biomed Eng. 2014 Jul;42(7):1494-507.
- 235. Lintzeris D, et al. Effect of a New Purified Collagen Matrix With Polyhexamethylene Biguanide on Recalcitrant Wounds of Various Etiologies: A Case Series. Wounds. 2018 Mar;30(3):72-78.
- 236. Bervoets A, Aerts O. Polyhexamethylene biguanide in wound care products: a non-negligible cause of peri-ulcer dermatitis. Contact Dermatitis. 2016 Jan;74(1):53-5.
- 237. Kanapathy M, et al. Epidermal grafting versus split-thickness skin grafting for wound healing (EPIGRAAFT): study protocol for a randomised controlled trial. Trials. 2016 May 17;17(1):245.
- 238. Cai SS, et al. A Case Series of Complex Recalcitrant Wounds Treated with Epidermal Grafts Harvested from an Automated Device. Cureus. 2016 Oct 30;8(10):e853.
- 239. Smith OJ, et al. The CelluTome epidermal graft-harvesting system: a patient-reported outcome measure and cost evaluation study. Int Wound J. 2016 Aug 4.
- 240. Fearmonti RM. Efficacy of Epidermal Skin Grafts Over Complex, Chronic Wounds in Patients With Multiple Comorbidities. Wounds. 2016 Jul;28(7):226-32.

- 241. Serena TE. Use of epidermal grafts in wounds: a review of an automated epidermal harvesting system. J Wound Care. 2015 Apr;24(4 Suppl):30-4.
- 242. MiMedx. (2019). EpiCord. Retrieved from https://mimedx.com/epicord/

Clinical Guideline Revision / History Information

Original Date: 7/31/2017

Reviewed/Revised: 1/18/2018, 7/31/2018, 7/23/2019, 07/21/2020, 08/04/2021, 10/21/2021,

12/01/2021, 07/26/2022, 07/19/2023