



Placental transfusion: may the “force” be with the baby

Judith S. Mercer^{1,2} · Debra A. Erickson-Owens² · Heike Rabe³

Received: 27 August 2020 / Revised: 26 February 2021 / Accepted: 29 March 2021
© The Author(s), under exclusive licence to Springer Nature America, Inc. 2021

Abstract

Placental transfusion results in a significant decrease in the risk of death for extremely preterm infants. With immediate cord clamping (ICC), these infants can leave up to one-half of their normal circulating in utero blood volume in the placenta. Extremely preterm infants are at highest risk of harm from ICC yet are currently the most likely to receive ICC. Receiving a placenta transfusion provides infants with life-saving components and enhanced perfusion. We present some lesser-known but important effects of placental transfusion. New research reveals that enhanced vascular perfusion causes an organ's endothelial cells to release angiocrine responses to guide essential functions. High progesterone levels and pulmonary artery pressure in the first few hours of life assist with neonatal adaptation. We propose that lack of essential blood volume may be a major factor contributing to inflammation, morbidities, and mortality that preterm infants frequently encounter.

Key points

- Placental transfusion provides enhanced vascular perfusion and reduces the risk of death for preterm infants
- Enhanced vascular perfusion stimulates endothelial cells to release vital angiocrine messengers to guide normal function and development of neonatal organs
- High progesterone levels and pulmonary artery pressure in the first 12 h may assist the newborn to adapt to the placental transfusion throughout the body
- Blood volume conservation is important at birth and during the NICU stay for preterm infants

Introduction

“It is not widely appreciated that the dysfunction of the inner lining of blood vessels is the single most common cause of human mortality. Endothelial cells (ECs) control their microenvironments as gate keepers of organ development, homeostasis, and tissue regeneration” [1].

Facilitation of placental transfusion has become the standard of care for newborns [2]. Emerging data from three systematic reviews reveal that the most striking benefit is a 27–30% lower death rate for preterm infants with ~60 s

delay in clamping compared to those with immediate cord clamping (ICC) [3–5]. A recent large ($n = 4680$) retrospective cohort study of extremely low gestational age neonates (≤ 26 weeks' gestational age at birth) reported lower rates of severe neurologic injury as well as death in the infants who had delayed cord clamping (DCC) compared to those with ICC [6]. Placental transfusion has also resulted in better developmental outcomes at 18 and 24 months for preterm infants after a delay in cord clamping and/or milking of the cord [7–9]. No recent studies on DCC have reported adverse outcomes although cord milking of infants under 28 weeks' gestation is under question. Yet, the best time to clamp and cut the umbilical cord at birth remains unknown, and most tiny infants and those infants needing resuscitation still receive ICC.

Placental transfusion at birth provides cardiovascular stability [10, 11], a huge allotment of vital stem cells [12–14], a 50% increase in iron-rich red cell volume [15], a large quantity of neuroprotective progesterone [16–19], and many important messengers including cytokines, chemokines, proteins, growth factors, and other hemostasis

✉ Judith S. Mercer
jmercerc@uri.edu

¹ Neonatal Research Institute at Sharp Mary Birch Hospital for Women and Newborns, San Diego, CA, USA

² University of Rhode Island, Kingston, RI, USA

³ Brighton and Sussex Medical School Royal Alexandra Childrens Hospital, Brighton, UK

facilitators essential for a normal transition [12]. Several excellent reviews about the benefits of placental transfusion are available [20–22].

Although placental transfusion results in a 27–30% decrease in the chance of death for preterm infants, there is no agreement as to how and why receiving some of the residual placental blood volume reduces the incidence of death. In this review we will not focus on the unanswered questions related to the clinical practice concerning placental transfusion. Instead, we will describe and discuss some important but lesser-known effects of placental transfusion in the transition to extra-uterine life of preterm and term infants that may impact the infant's transition and suggest some ideas for addressing some of the most troublesome problems for infants.

Blood as an essential factor for growth and regeneration

Science has not yet revealed exactly how and why placental transfusion reduces the incidence of death for premature infants. We suggest that recent work in biology, physiology,

biometrics, and biophysics offers important new clues about how an enhanced blood volume may benefit newborns. The most plausible reason for the decreased incidence of death may be increased blood volume replete with all of its essential components that infants can receive with placental transfusion [12, 23].

The vascular system, a vast network of capillaries lined by ECs, interconnects arteries and veins creating vascular beds in organs throughout the body—a prerequisite of organ development [24, 25]. It is known that capillary ECs are not merely passive conduits for the delivery of oxygen or nutrients, but also support organ development and organ regeneration through elaboration of tissue-specific paracrine growth factors. These factors, known as angiocrine factors, are secreted by the ECs and are essential players in all physiologic processes [26, 27]. Only recently has research shown that these angiocrine factors can be stimulated by mechanical forces alone [28]. Lorenz et al. demonstrated that shear forces generated by dynamic blood flow appear to induce mechanical signals in ECs causing release of important angiocrine factors essential for homeostasis, and for normal growth and development (Fig. 1) [28].

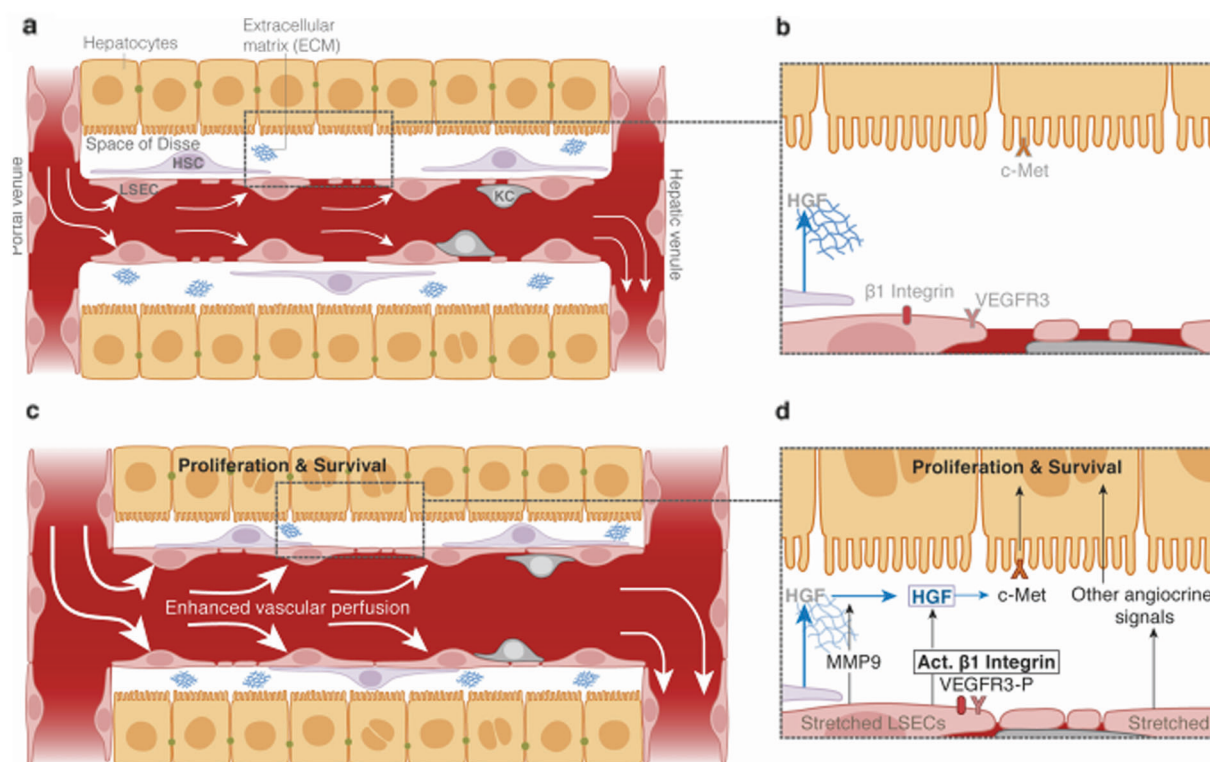


Fig. 1 Model of mechanotransduced angiocrine signals in the liver. Simplified drawing of liver sinusoids with liver sinusoidal endothelial cells (LSECs), hepatic stellate cells (HSCs), extracellular matrix (ECM), Kupffer cells (KCs), and hepatocytes: **a** liver sinusoid under normal blood flow; **b** magnification of space of Disse; **c** liver sinusoid with enhanced blood perfusion; **d** magnification of the space of Disse. When the vascular

lumen widens owing to enhanced blood perfusion, circumferential stretching of liver sinusoidal ECs activates endothelial $\beta 1$ integrin and interaction with VEGFR3. The hepatic ECs (in concert with other cells) subsequently release angiocrine signals (such as HGF, IL-6, and TNF) and activate MMP9 and thus enhance proliferation and survival of the adjacent hepatocytes. Adapted from Lorenz et al. [28] (with permission).

In liver cells, Lorenz et al. demonstrated that enhanced vascular perfusion induces the proliferation of hepatocytes driving liver development. Using murine embryos, they noted that liver expansion was preceded by initial new blood perfusion in a highly controlled manner starting in the periphery and proceeding toward the center of the liver. They found higher levels of activated forms of two important angiocrine growth factors, $\beta 1$ integrin and vascular endothelial growth factor receptor 3, in the periphery of the liver where there was extensive blood perfusion and hyperproliferation of hepatocytes. In subsequent perfusion experiments, they showed that activation of the angiocrine growth factors was reduced when blood perfusion was slowed by decreasing the embryo's heartbeat and accelerated when the heart rate was increased (pharmacologically). Next, they provided evidence (ex vivo and in vitro) that in human liver cells, mechanical stretching alone triggered activation of the same growth factors. Remarkably, culturing human adult liver cells in medium collected from stretched human liver cells resulted in increased proliferation and decreased apoptotic cell death. The process responsible for these changes, known as mechanotransduction, is a mechanism by which cells convert mechanical stimuli into electrochemical activity. This occurs in most if not all organs throughout the body [24, 25]. Lorenz clearly demonstrated that enhanced vascular perfusion provides mechanical stimuli (stretching, shear force) through vasodilation that stimulated organ-specific ECs to release angiocrine factors that guide physiologic function and development of the liver (Fig. 1).

Although Lorenz's work supporting the importance of an ample blood volume was confined to hepatocytes, advances in our understanding of one organ system can inform us about other systems [24, 29, 30]. Many of the intercellular signaling networks that control the morphogenesis of the respiratory system are similarly present in the development of other organs such as the kidney and liver [29]. While angiocrine factors appear to be organ specific, most seem to depend on mechanotransduction force from vasodilation. For example, prolonged or severe hypoperfusion in preterm infants is one of the most common causes of acute intrinsic kidney injury (AKI) often leading to tubular dysfunction and acute tubular necrosis [31]. While damage to tubular epithelial cells is a hallmark of ischemic AKI, “damage to the inner most lining of the renal vascular system, the ECs, has a critical role in the initiation, extension, maintenance, and recovery phases of AKI...in ischemic AKI, with restoration of renal blood flow, glomerular filtration rate can return promptly to normal” [31]. Most organs experience vasodilation—and thus EC stretching—when their physiological function is extensively needed as they start to grow (Fig. 1) [24]. When damaged, ECs produce a systemic

inflammatory response that directly affects the brain, lung, heart, liver, bone marrow, and GI tract [32]. In the presence of hypoperfusion, ECs may be unable to support expected functions including organ growth and development.

The benefits from placental transfusion most likely are related to the autotransfusion of the large volume of residual blood available to the newborn. It is probable that the enhanced blood flow generates mechanical forces within the microcirculation that initiate proper organ health and development [1, 28, 30]. ICC or early cord clamping (ECC) reduces potential blood volume for the neonate that may contribute to loss of organ-specific vascular competence in the lung, gastrointestinal tract, brain, kidney, and other organs.

Evidence of enhanced perfusion from placental transfusion

Classic physiologic studies completed over the past 60 years document that placental transfusion results in improved perfusion in the neonate's hematologic, urinary, gastrointestinal, neurological, and respiratory systems [33–38]. The improved perfusion was demonstrated in a 1968 study in which heel capillaries were biopsied at 2–5 h after birth from 12 neonates who had either ICC or DCC (when pulsations ceased) [39]. As seen in Fig. 2 (electron microscopy), the capillary on the left (Fig. 2a) has a small, irregular lumen with thick endothelium while in the capillary on the right (Fig. 2b) the lumen is large and full and the endothelium is thin with small fenestrations at the top allowing for maximal exchange of gases, nutrients, and waste products. Pietra et al. found significantly more capillaries resembling the undistended capillary in the neonates who received ICC and more resembling the distended capillary in the neonates who experienced DCC, suggesting better blood volume and perfusion with DCC [39].

As heels are usually the last area of the body to be well perfused, it is likely that other more vital areas such as brain and heart are also better perfused. This graphic description of the effect of a full blood volume on vascular perfusion in the heels of newborns could never be repeated today. Yet, it is important because it so clearly demonstrates the increased vascular perfusion that is a consequence of placental transfusion.

The impact of enhanced perfusion on fetal–neonatal transition

We propose that Lorenz's work applies to the transition from fetus to neonate with a major role in inducing the

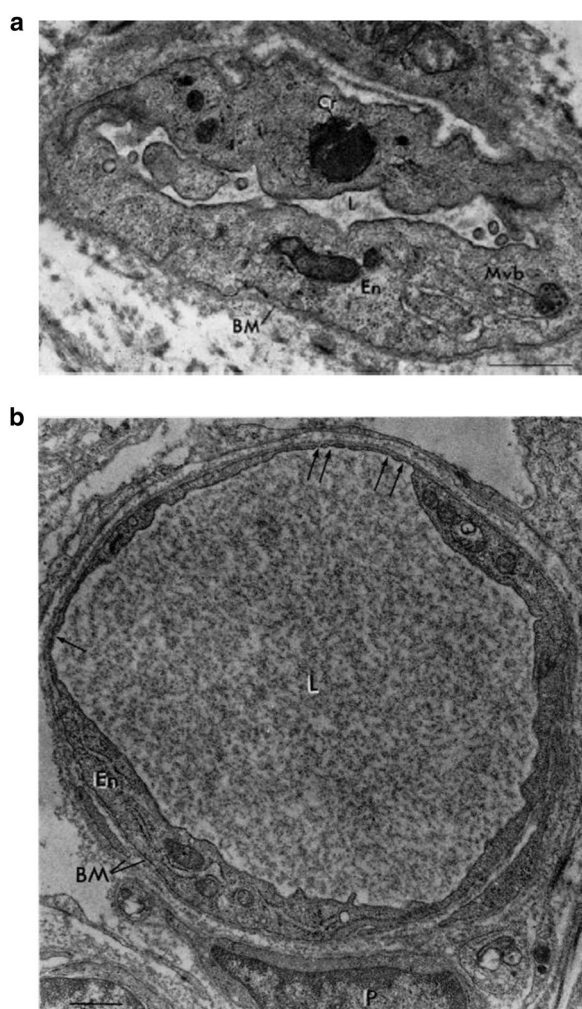


Fig. 2 Electron microscopy photographs of capillaries from heel biopsies of infants with immediate cord clamping (ICC) or cord clamping when pulsations stopped (± 5 min, DCC). There were more capillaries found resembling the specimen on the left (a) in the infants with ICC and more of the distended capillaries in infants with DCC on the right (b). There are small fenestrations at the top of the arrows on the DCC capillaries that would aid in transport of gases and nutrients in b only. Adapted from Pietra et al. [39] (with permission).

beginning of normal organ development from the mechanical force of a large new blood volume auto-transfused over the first few minutes after birth. Our hypothesis is that the residual blood volume obtained by placental transfusion creates increased systemic and regional organ blood flow and vasodilation and is augmented by high initial levels of progesterone and high pulmonary artery pressure (PAP) over the first several hours [16, 35]. Tenets of this hypothesis include initial distension of the lung capillaries, removal of lung liquid, high PAP, and the overall effect of high progesterone levels on transition and are discussed in the following paragraphs.

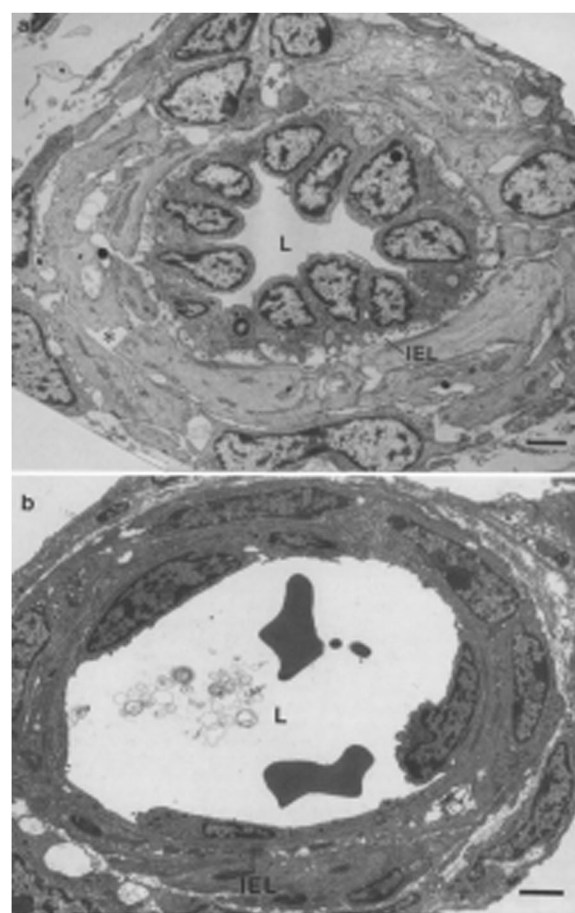


Fig. 3 Electron micrographs of transverse sections through small muscular lung arterioles in naturally born piglets. a Stillborn; b aged 5 min taken at the same magnification. At birth, the endothelial cells of the intra-acinar arteries showed more rapid and greater changes in shape and thickness than did the cells of more proximal vessels. IEL internal elastic lamina, L lumen. Scale bar line on lower right = 2 μ m. Adapted from Haworth et al. [43] (with permission).

Distension of alveolar capillaries

In fetal life, lung growth depends on the lungs (air sacs) being highly distended with a large volume (20–30 mL/kg) of fetal lung liquid [40, 41]. Without this liquid, the fetus will develop hypoplastic lungs as the special liquid creates lung expansion along with mechanical stress. Blood supply to the fetal lung is ~8% of the cardiac output. Placental transfusion can supply ~30 mL/kg of blood volume to the neonate some of which goes to fill the capillaries surrounding each alveolus for the first time.

We propose that, with a full placental transfusion, stretch is applied within the capillary side of the alveolar–capillary membrane instead of by fetal lung liquid within the alveolus [42]. At birth, the alveolar capillaries adapt to the sudden influx of blood by increasing the diameter of the lumen, but do not change the diameter of the capillary itself thus not effecting the overall lung volume (Fig. 3) [43].

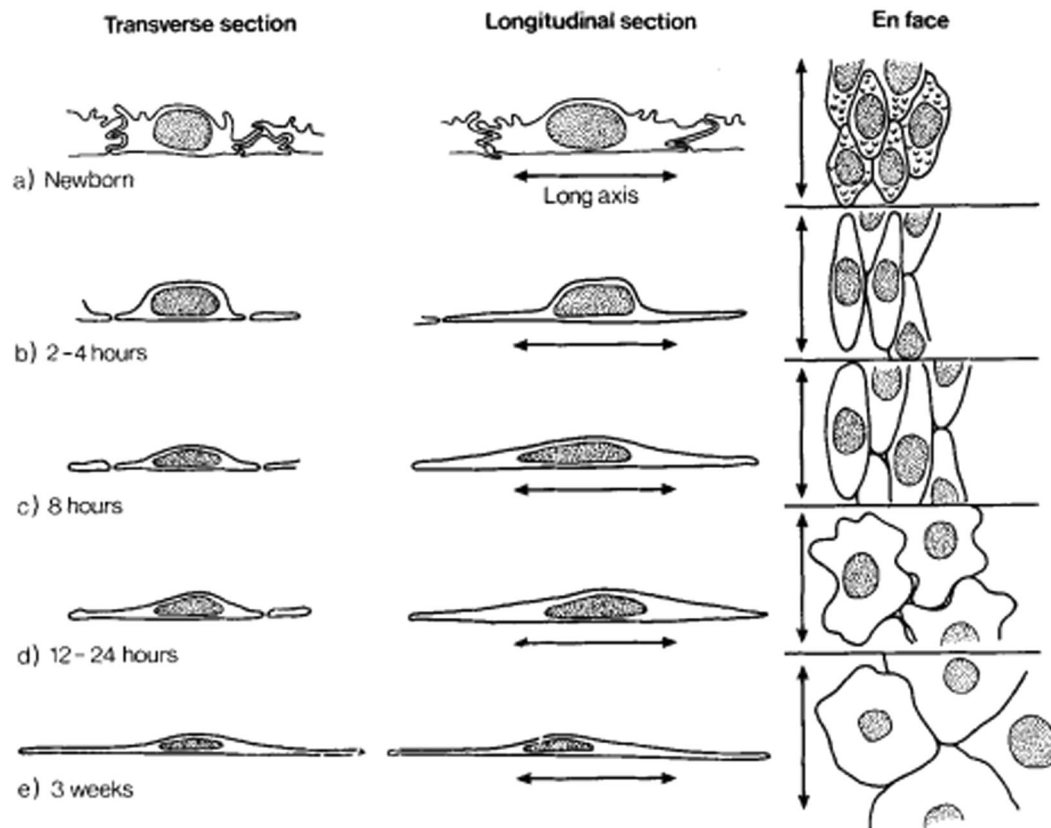


Fig. 4 Diagram illustrating en face shape changes in the endothelial cells of intra-acinar arteries during the first 3 weeks of life (porcine). Two phases of change are shown: a–c stretching during inflation; d, e spreading during dilatation. The endothelial cells of the intra-acinar arteries showed marked changes in cell shape and

relationships after birth while those of large preacinar arteries did not. The first structural changes detected during the first 30 min of life occurred in the endothelial cells lining the intra-acinar arteries. Adapted from Hall and Haworth [44] (with permission).

This allows transfer and inflow of the large volume of blood formerly used by the placenta for respiration in fetal life. Instead of pressure in the alveolus from fluid, pressure is created by new blood flow in the alveolar capillaries. While in small preterm in the canaliculi or saccular stage alveoli are not fully developed, there must be approximation with blood source for oxygenation to occur. This is essential as air exchange following birth must occur in the alveolus itself and rapid growth must continue. It is likely that the enhanced vascular perfusion provides mechanical stimuli (stretching) through vasodilation to continue the distension (stretch; shear force), so important in fetal life, in order to now support new neonatal lung growth [28, 30]. Effects of the essential stretch over time on the vessels surrounding the alveoli are readily seen in the porcine lung over the first minutes (Fig. 3) and hours (Fig. 4) following normal birth [43, 44].

The large volume of blood flow in the alveolar capillaries triggers a mechanical force that causes the capillaries to become erect playing a vital role in recruitment of lung tissue and preventing derecruitment [42, 45]. It has been said that “capillary erection” as described by Jaykka has

been disproven. In fact, the only study that tried used inappropriate subjects (had previously breathed) to try to replicate Jaykka’s work [46]. However, both Jaykka and Avery reported that lungs were easier to inflate if there was fluid in the lung vessels. Thus, waiting to clamp the cord until after pulsations cease allows time for the lungs to fill fully with blood and to augment fetal lung fluid removal.

Removing acidic fetal lung fluid

Fetal lung liquid has a pH of 6.27 and must be removed rapidly from the lung in order for the infant to breathe air [40]. The anatomy of the alveolus and its blood supply informs the process of removal. While approximately 80% of the alveoli develop after birth, ~50 million alveoli are covered by 56 billion capillary segments in the term neonate [47]. This results in a ratio of 1 alveolus to ~900 million capillary segments accounting for the denseness of the capillaries. The pulmonary capillary networks are so dense that an almost contiguous sheet of blood encircles each alveolus [48]. The interstitial space between an alveolus and the surrounding capillary network is extremely thin, with

only a 0.5 μm barrier known as the alveolar–capillary membrane separating air (or fluid) in the alveoli from blood in the pulmonary capillaries [49]. The thinness of this barrier facilitates the exchange of oxygen and carbon dioxide. Filled with rapidly flowing blood, these capillaries present a powerful force for ridding the lung of the acidic fetal lung liquid as quickly as possible. The blood from the placental “lungs” rapidly enters the newborn lung at birth causing alveolar capillary erection and supports rapid removal of fetal lung liquid out of the alveoli through various active and passive transport mechanisms (including sodium channel activation) across the very thin layer of tissues (interstitium) into the capillaries surrounding each alveolus and thus into the general circulation [42, 45].

It is probable that the removal of the fetal lung liquid is responsible for the “hidden acidosis” that Wiberg and others have reported in infants with DCC [50–52]. Wiberg sampled blood in the intact cords and found an increase in acidic blood coming from the intact umbilical arteries at 45 s of life but not until 90 s following birth in the vein. This indicates that the newborn was the source of the acidosis and not the placenta. These findings occurred in infants born either vaginally or by cesarean section. Giovannini reported increased acidosis from birth to 3 min after examining paired arterial–venous blood gas parameters drawn from the unclamped cord and again after a 3-min delay in cord clamping [50]. This occurred in spite of good oxygenation, thereby suggesting that the acidic fetal lung liquid enters the infant’s blood supply rapidly. These reported changes in the acidity of the blood are statistically, but not clinically, significant. However, in an infant with ICC, the acidic lung liquid will be diluted with ~30% less blood volume that may make removal of the acidic lung liquid more difficult.

Many sources state that the fetal lung liquid moves to the “interstitium” when air is forced into the lungs. The idea of forcing air into this very delicate tissue has been shown to be harmful and resulted in increased death rates in premature infants in the first 48 h [53]. Nair found that sustained inflation, even with DCC, prevented blood flow into the lung via the pulmonary artery and inferior vena cava [54] (Poster Presentation, PAS 4/28/2019). Also, there would not be enough area in the lung “interstitium” for this large volume of lung liquid, even briefly, without severe compromise. The thick interstitium seen in lung samples of infants dying from respiratory complications is a pathologic outcome of their condition and not typical in a healthy lung [54].

Progesterone: the key to incorporation of placental transfusion

The newborn’s body is likely primed for capillary erection and enhanced vascular perfusion by high levels of progesterone, a potent vasodilator [55]. Immediately after a term

birth, the amount of progesterone in the neonate is almost twice as high as it is in the mother at term (~270 versus ~170 ng/mL) [16, 17]. It is likely that this high level of progesterone assists in the vascular modeling that takes place over the first hours of newborn transition because of its effect of relaxation on the vasculature (Figs. 3 and 4) [43, 44, 55]. Progesterone also plays a role in dampening the response to inflammatory cytokines and is highly neuroprotective for the fetus and the newborn [18]. Progesterone levels quickly fall in the mother and the newborn over the first day after birth and thereafter continue to decline over the first week of life [16]. Thus, the essential initial vascular modeling in the lung is likely to be uniquely facilitated by the high levels of circulating progesterone in the infant in the first 12 h of life suggesting this may be a critical period in organ development.

Enhanced perfusion from placental transfusion: the role of initial high pulmonary artery pressure

Placental transfusion results in enhanced perfusion, thus offering a significant mechanical force to drive transformation in the lung tissue and other organs that are now required to begin to function independently from the placenta [35]. Arcilla compared (using cardiac catheterization) PAP in term infants who received a full placental transfusion caused by a 5-min delay in umbilical cord clamping (positioned lower than placenta) with infants who received ICC. The PAP in the DCC group remained high for 10+ h in contrast to those infants with ICC whose PAP fell to 70% of systemic pressure by 2 h of age and to 50% by 4 h [35]. The initial high PAP, along with the high progesterone levels and a full placental transfusion, may be necessary to facilitate the essential stretch and mechanotransduction needed for post-birth lung transition [16, 43, 44]. The combined effect of high progesterone and PAP supports the assimilation of the large placental transfusion at birth and full perfusion to all of the infant’s organs in adapting to extra-uterine life. Together, these factors appear to make the first 10–12 h of an infant’s life a unique critical period for organ development that can be enhanced by a full placental transfusion.

Is over-transfusion from placental transfusion possible?

Concerns have been raised that a full placental transfusion could cause “over-transfusion” in the neonate. Yet, none of the studies on placental transfusion have found evidence of issues of over-transfusion. All fetal respiration has taken place in the placenta requiring a large part of the fetal cardiac output. Immediately at birth, respiration must switch to the newborn lung. To do so, a similar volume of blood, 40–50% of the cardiac output, is required by the newborn

lung for adequate oxygenation. The blood an infant receives from placental transfusion is not “extra”—it is the same volume of blood that the fetus’s heart was pumping in utero out to the placenta and back again to perfuse his body and the placenta (i.e., the fetal lung). The newborn’s body is programmed to receive the transfer or autotransfusion of the residual placental blood as explained above. At no other time, short of a serious hemorrhage, could a human body incorporate such a large volume of blood. Thus, it is highly unlikely that an infant can be over-transfused from his own placental transfusion. Under-perfusion resulting in hypovolemia may be a much larger problem as discussed in the following section. A 1-min delay may not yield an adequate increase in blood volume to impact rates of other morbidities in the smallest infants. Research is ongoing that allows a longer time for the infant to transition (NCT02671305, NCT03019367, NCT02742454).

What is lost due to immediate cord clamping?

Residual placental blood volume is the blood remaining in the placenta after the cord is clamped and cut. This is the blood that is used when stem cells are harvested. Seeking to identify covariates of bronchopulmonary dysplasia (BPD), Chaudhury et al. completed a prospective study of 200 preterm infants <32 weeks by collecting residual placental blood volume for analyses [12]. For infants 29–32 weeks, the amount of whole residual blood collected was 35 mL/kg (IQR 28–41). However, for the 35 infants between 23 and 28 weeks, the amount collected was 46 mL/kg (IQR 37–57) and for the 17 very low birth weight (VLBW) infants <1000 g, it was 57 mL/kg (IQR 47–62). Given a fetal–placental blood volume of 110–115 mL/kg [56], the tiniest infants with ICC lost to cord blood collection half or more of the blood volume potentially available to them with DCC.

Higher levels of stem cells (e.g., CD34+, CD90+, CD105+) were found in the residual placental blood of infants who developed BPD, with the highest stem cell levels for those who developed the worst BPD. In the 74 infants whose cord blood was analyzed to measure 12 cytokines and growth factors, higher levels of angiopoietin, IL-8, and PGF, among others were found in the blood of those developing BPD.

Chaudhury et al. state that their findings may have important implications for those studying stem and progenitor therapies to use for the prevention of BPD. However, when harvested residual umbilical cord blood is processed for storage, the iron-rich red blood cells and the other protective substances in the blood along with the plasma are discarded. When an infant receives the residual placental blood volume at the time of birth, the stem cells

are in the perfect medium along with many cytokines, proangiogenic and antiapoptotic messengers, and growth stimulating factors [12, 23]. Finding the appropriate medium is a major obstacle confronting stem cell therapies especially when attempting proliferation of stem cells and translation of successful animal studies to humans [57].

Baker states that in spite of years of successful animal studies, translation of stem cell therapy to human infants has not been productive in preventing, or in treating, the most damaging diseases such as BPD that preterm infants acquire [57, 58]. Other issues in postnatal transfusion of stem cells include finding the correct dosage, timing of doses, single or multiple doses, cell suspension density, route of administration, infusion protocols, and regulatory issues and good manufacturing procedures [57]. While these factors are important to resolve for use of stem cells in infants already born, the initial autologous auto-transfusion that occurs with placental transfusion at birth preserves all components of the residual placental and cord blood for the infant. This potentially leads to reduced inflammation and conserves the infant’s progenitor cells [23, 57].

The loss of fresh whole fetal blood, and its many components, when ICC or ECC (<30 s) occurs is underappreciated. Not only does the concentration of stem cells increase with decreasing gestational age at birth but preterm stem cells also have higher clonogenic activity [59, 60]. With ICC, larger amounts of these essential substances are unavailable to the tiniest infants. Did the infants in Chaudhury’s study develop BPD because of an intrinsic problem or because they lacked the cells and substances they needed for a successful transition? The large volume of blood lost by the smallest infants with ICC would be considered equivalent to severe hemorrhage and has been shown to result in an inflammatory response in adult humans and rodents [61, 62]. Severe blood volume loss alone has been shown to lead to inflammation without any accompanying infectious process or reperfusion [61, 62]. Makley reports that after hemorrhage, replacing the lost blood volume with fresh whole blood reduces inflammation more quickly and efficiently than replacement with donor blood or other transfusion products [63]. Placental transfusion can supply that fresh whole blood to protect the neonate and is readily available at birth. Given that almost all newborn problems have a large inflammatory component, this information begs further study in human infants.

Premature infants are at the highest risk from harm due to ICC or ECC

Preterm infants have about 50% of their fetal–placental blood volume in the placenta at any point in time due to the 1:1 placenta to body ratio in contrast to term infants whose

placentas are approximately 1/3 the size of the fetus at the time of birth. With ICC, Chaudhury et al. demonstrated that the smallest infants receive a lower percentage of essential residual cord blood with reduced stem cell levels and other important factors essential for building their hematologic and immune systems after birth [12]. In addition to ICC, the tiniest infants have a large blood sample for admission laboratory studies and frequent multiple phlebotomies for testing throughout the NICU stay [64]. It is likely that these actions contribute to susceptibility to hypoperfusion and the devastating and too common illnesses of prematurity [65]. Hellstrom et al. reported that preterm infants lose 58% of their own newborn blood volume in the first 2 weeks due to phlebotomy. While this is most often replaced with adult blood, many of the components unique to fetal/newborn blood are lost. We suggest that new approaches are needed to protect the smallest infants: a safer, more physiological way to get more of the available residual placental blood volume into the preterm infant at the time of birth; admission blood sampling from the umbilical cord or placental veins; and micro-sampling techniques for NICU blood testing.

Placental transfusion at birth

If one thinks of death, disease, and health on a continuum, it is not unreasonable to suggest that if a DCC of ~60 s reduces the rate of death, obtaining an even greater placental transfusion may further reduce death and illnesses. We have provided evidence suggesting that birth is the time when the infant's body is best prepared to receive the residual placental blood. There is likely greater receptivity and plasticity of the vascular system due to the high levels of progesterone that will decrease within ~12 h. Longer DCC times than the current recommended 30–60 s may provide additional benefits and be needed to accomplish a greater transfusion for preterm infants. Katheria provided a review of current and ongoing studies on longer delays [66].

Blood conservation during transition and NICU stay

Two other methods to protect the premature infant's blood volume included collection of blood for initial admission studies from the umbilical cord after delivery and use of microsystems for ongoing testing in the NICU. Using cord samples for admission blood work has been shown to result in higher 24-h hematocrit and hemoglobin and can lower risk of hypotension [67–69]. Phlebotomy losses following birth lead to nearly universal anemia in VLBW infants [64, 65, 70]. Rabe reported that some tiny infants had over 30 mL/kg withdrawn during the NICU stay [71]. More microsystems that can reduce the volume of blood removed from the infant for testing are needed [64, 65, 72].

Conclusion

The importance of the microvascular system among very preterm infants in disease progression is only beginning to be appreciated for its significant clinical implications. Enhanced vascular perfusion provides mechanical stimuli causing electrochemical activity that seems increasingly likely to be essential for normal function, development, and maintenance of newborn organs, especially the lungs. It is probable that immediate or ECC denies the infant the enhanced vascular perfusion needed to provide the mechanical force to stimulate organ-specific ECs to direct their growth, maintenance, and repair. Methods to enhance and protect neonatal blood volume via placental transfusion at birth and conserve blood in the NICU are under study and may help prevent or lessen the burden some of the morbidities observed in very preterm infants.

Acknowledgements The authors wish to thank John (Jack) Widness, MD, for his generous and insightful reviews of this article.

Author contributions JSM conceived of the idea for the paper; all authors made substantial contributions, revised and reviewed the article, and finally approved the paper. All authors agree to be accountable for the information presented in the paper.

Compliance with ethical standards

Conflict of interest The authors declare no competing interests.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

References

1. Augustin HG, Koh GY. Organotypic vasculature: from descriptive heterogeneity to functional pathophysiology. *Science*. 2017;357:6353.
2. Committee on Obstetric Practice. Committee Opinion No. 684. Delayed umbilical cord clamping after birth. *Obstet Gynecol*. 2017;129:e5–10.
3. Fogarty M, Osborn DA, Askie L, Seidler AL, Hunter K, Lui K, et al. Delayed vs early umbilical cord clamping for preterm infants: a systematic review and meta-analysis. *Am J Obstet Gynecol*. 2018;218:1–18.
4. Backes CH, Rivera BK, Haque U, Bridge JA, Smith CV, Hutchon DJR, et al. Placental transfusion strategies in very preterm neonates: a systematic review and meta-analysis. *Obstet Gynecol*. 2014;124:47–56.
5. Rabe H, Gyte GM, Díaz-Rossello JL, Duley L. Effect of timing of umbilical cord clamping and other strategies to influence placental transfusion at preterm birth on maternal and infant outcomes. *Cochrane Pregnancy and Childbirth Group, editor. Cochrane Database Syst Rev*. 17 Sep 2019. <http://doi.wiley.com/10.1002/14651858.CD003248.pub4>. Accessed 10 Nov 2019.
6. Lodha A, Shah PS, Soraisham AS, Rabi Y, Abou Mehrem A, Singhal N, et al. Association of deferred vs immediate cord clamping with severe neurological injury and survival in

- extremely low-gestational-age neonates. *JAMA Netw Open*. 2019;2:e191286.
7. Katheria A, Garey D, Truong G, Akshoomoff N, Steen J, Maldonado M, et al. A randomized clinical trial of umbilical cord milking vs delayed cord clamping in preterm infants: neurodevelopmental outcomes at 22–26 months of corrected age. *J Pediatr*. 2018;194:76–80.
 8. Mercer JS, Erickson-Owens DA, Vohr BR, Tucker RJ, Parker AB, Oh W, et al. Effects of placental transfusion on neonatal and 18 month outcomes in preterm infants: a randomized controlled trial. *J Pediatr*. 2016;168:50–55. e1.
 9. Rabe H, Sawyer A, Amess P, Ayers S. Brighton Perinatal Study Group Neurodevelopmental outcomes at 2 and 3.5 years for very preterm babies enrolled in a randomized trial of milking the umbilical cord versus delayed cord clamping. *Neonatology*. 2016;109:113–9.
 10. Bhatt S, Polglase GR, Wallace EM, te Pas AB, Hooper SB. Ventilation before umbilical cord clamping improves the physiological transition at birth. *Front Pediatr*. 20 Oct 2014. <http://journal.frontiersin.org/article/10.3389/fped.2014.00113/abstract>. Accessed 4 Oct 2019.
 11. Cashore W. Hypovolemia resulting from a tight nuchal cord at birth. *Pediatr Res*. 1973;7:399.
 12. Chaudhury S, Saqibuddin J, Birkett R, Falcon-Girard K, Kraus M, Ernst LM, et al. Variations in umbilical cord hematopoietic and mesenchymal stem cells with bronchopulmonary dysplasia. *Front Pediatr*. 2019;7:475.
 13. Cotten CM, Murtha AP, Goldberg RN, Grotegut CA, Smith PB, Goldstein RF, et al. Feasibility of autologous cord blood cells for infants with hypoxic-ischemic encephalopathy. *J Pediatr*. 2014;164:973. e1.
 14. Liao Y, Cotten M, Tan S, Kurtzberg J, Cairo MS. Rescuing the neonatal brain from hypoxic injury with autologous cord blood. *Bone Marrow Transpl*. 2013;48:890–900.
 15. Yao AC, Moinian M, Lind J. Distribution of blood between infant and placenta after birth. *Lancet Lond Engl*. 1969;2:871–3.
 16. Sippell WG, Becker H, Versmold HT, Bidlingmaier F, Knorr D. Longitudinal studies of plasma aldosterone, corticosterone, deoxycorticosterone, progesterone, 17-hydroxyprogesterone, cortisol, and cortisone determined simultaneously in mother and child at birth and during the early neonatal period. I. Spontaneous delivery. *J Clin Endocrinol Metab*. 1978;46:971–85.
 17. Trotter A, Maier L, Kron M, Pohlandt F. Effect of oestradiol and progesterone replacement on bronchopulmonary dysplasia in extremely preterm infants. *Arch Dis Child Fetal Neonatal Ed*. 2007;92:F94–98.
 18. González-Orozco JC, Camacho-Arroyo I. Progesterone actions during central nervous system development. *Front Neurosci*. 2019;13:503.
 19. Berger R, Söder S. Neuroprotection in preterm infants. *BioMed Res Int*. 2015;2015:1–14.
 20. Katheria AC, Lakshminrusimha S, Rabe H, McAdams R, Mercer JS. Placental transfusion: a review. *J Perinatol*. 2017;37:105–11.
 21. Kresch M. Management of the third stage of labor: how delayed umbilical cord clamping can affect neonatal outcome. *Am J Perinatol*. 2017;34:1375–81.
 22. Mercer JS, Erickson-Owens DA. Rethinking placental transfusion and cord clamping issues. *J Perinat Neonatal Nurs*. 2012;26:202–17. quiz 218–9.
 23. Tolosa JN, Park D-H, Eve DJ, Klasko SK, Borlongan CV, Sanberg PR. Mankind's first natural stem cell transplant. *J Cell Mol Med*. 2010;14:488–95.
 24. Li J, Wang Z, Chu Q, Jiang K, Li J, Tang N. The strength of mechanical forces determines the differentiation of alveolar epithelial cells. *Dev Cell*. 2018;44:297–312. e5.
 25. Peng T, Morrissey EE. Development of the pulmonary vasculature: current understanding and concepts for the future. *Pulm Circ*. 2013;3:176–8.
 26. Rafii S, Butler JM, Ding B-S. Angiocrine functions of organ-specific endothelial cells. *Nature*. 2016;529:316–25.
 27. Pasquier J, Ghiabi P, Chouchane L, Razzouk K, Rafii S, Rafii A. Angiocrine endothelium: from physiology to cancer. *J Transl Med*. 2020;18:52.
 28. Lorenz L, Axnick J, Buschmann T, Henning C, Uner S, Fang S, et al. Mechanosensing by $\beta 1$ integrin induces angiocrine signals for liver growth and survival. *Nature*. 2018;562:128–32.
 29. Morrissey EE, Hogan BLM. Preparing for the first breath: genetic and cellular mechanisms in lung development. *Dev Cell*. 2010;18:8–23.
 30. Li J, Tang N. May the force be with you. *Dev Cell*. 2018;47:673–4.
 31. Askenazi D, Selewski D, Willig L, Warady B. Chapter 90. Acute kidney injury and chronic kidney disease. In: Gleason C, Juul S (eds). *Avery's disease of the newborn*. 10th ed. Philadelphia, PA: Elsevier; 2018. p. 1280–5.
 32. Awad AS, Okusa MD. Distant organ injury following acute kidney injury. *Am J Physiol-Ren Physiol*. 2007;293:F28–9.
 33. Oh WOM, Lind J. Renal function and blood volume in newborn infant related to placental transfusion. *Acta Paediatr Scand*. 1967;55:197–210.
 34. Arcilla RA, Oh W, Wallgren G, Hanson JS, Gessner IH, Lind J. Quantitative studies of the human neonatal circulation. II. Hemodynamic findings in early and late clamping of the umbilical cord. *Acta Paediatr Scand* 1967;179(Suppl):25.
 35. Arcilla RA, Oh W, Lind J, Gessner IH. Pulmonary arterial pressures of newborn infants born with early and late clamping of the cord. *Acta Paediatr*. 1966;55:305–15.
 36. Nelle M, Zilow EP, Bastert G, Linderkamp O. Effect of Leboyer childbirth on cardiac output, cerebral and gastrointestinal blood flow velocities in full-term neonates. *Am J Perinatol*. 1995;12:212–6.
 37. Oh W, Lind J. Body temperature of the newborn infant in relation to placental transfusion. *Acta Paediatr Scand*. 1967;172(Suppl):135.
 38. Oh W, Lind J, Gessner IH. The circulatory and respiratory adaptation to early and late cord clamping in newborn infants. *Acta Paediatr Scand*. 1966;55:17–25.
 39. Pietra GG, D'Amodio MD, Leventhal MM, Oh W, Braudo JL. Electron microscopy of cutaneous capillaries of newborn infants: effects of placental transfusion. *Pediatrics*. 1968;42:678–83.
 40. Plosa E, Guttentag SH. Lung development. In: Gleason C, Juul S (eds). *Avery's diseases of the newborn*. 10th ed. Philadelphia, PA: Elsevier; 2018. p. 586–99.
 41. Hooper SB, Harding R. Fetal lung liquid: a major determinant of the growth and functional development of the fetal lung. *Clin Exp Pharm Physiol*. 1995;22:235–47.
 42. Jaykka S. Capillary erection and the structural appearance of fetal and neonatal lungs. *Acta Paediatr*. 1958;47:484–500.
 43. Haworth SG, Hall SM, Chew M, Allen K. Thinning of fetal pulmonary arterial wall and postnatal remodelling: ultrastructural studies on the respiratory unit arteries of the pig. *Virchows Arch A Pathol Anat Histopathol*. 1987;411:161–71.
 44. Hall SM, Haworth SG. Normal adaptation of pulmonary arterial intima to extrauterine life in the pig: ultrastructural studies. *J Pathol*. 1986;149:55–66.
 45. Mercer JS, Skovgaard RL. Neonatal transitional physiology: a new paradigm. *J Perinat Neonatal Nurs*. 2002;15:56–75.
 46. Avery ME, Frank NR, Gribetz I. The inflationary force produced by pulmonary vascular distension in excised lungs. *J Clin Invest*. 1959;38:456–62.

47. Weibel ER, Gomez DM. Architecture of the human lung: use of quantitative methods establishes fundamental relations between size and number of lung structures. *Science*. 1962;137:577–85.
48. Sherwood L. Human physiology: from cells to systems. 9th ed. Boston, MA, USA: Cengage Learning; 2016. 1 p.
49. Sherwood L. Human physiology: from cells to systems. 8th ed. Belmont, CA: Brooks/Cole, Cengage Learning; 2013. 1 p.
50. Giovannini N, Crippa B, Denaro E, Raffaeli G, Cortesi V, Consonni D, et al. The effect of delayed umbilical cord clamping on cord blood gas analysis in vaginal and caesarean-delivered term newborns without fetal distress: a prospective observational study. *BJOG Int J Obstet Gynaecol*. 2020;127:405–13.
51. Wiberg N, Källén K, Olofsson P. Delayed umbilical cord clamping at birth has effects on arterial and venous blood gases and lactate concentrations. *BJOG Int J Obstet Gynaecol*. 2008;115:697–703.
52. Mokarami P, Wiberg N, Olofsson P. Hidden acidosis: an explanation of acid-base and lactate changes occurring in umbilical cord blood after delayed sampling. *BJOG Int J Obstet Gynaecol*. 2013;120:996–1002.
53. Kirpalani H, Ratcliffe SJ, Keszler M, Davis PG, Foglia EE, Te Pas A, et al. Effect of sustained inflations vs intermittent positive pressure ventilation on bronchopulmonary dysplasia or death among extremely preterm infants: the SAIL randomized clinical trial. *JAMA* 2019;321:1165–75.
54. Jackson JC, Truog WE, Standaert TA, Juul SE, Murphy JH, Chi EY, et al. Effect of high-frequency ventilation on the development of alveolar edema in premature monkeys at risk for hyaline membrane disease. *Am Rev Respir Dis*. 1991;143:865–71.
55. Barbagallo M, Dominguez LJ, Licata G, Shan J, Bing L, Karpinski E, et al. Vascular effects of progesterone: role of cellular calcium regulation. *Hypertension*. 2001;37:142–7.
56. Linderkamp O. Placental transfusion: determinants and effects. *Clin Perinatol*. 1982;9:559–92.
57. Baker EK, Jacobs SE, Lim R, Wallace EM, Davis PG. Cell therapy for the preterm infant: promise and practicalities. *Arch Dis Child Fetal Neonatal Ed*. 2020;105:563–8.
58. Thébaud B, Goss KN, Laughon M, Whitsett JA, Abman SH, Steinhorn RH, et al. Bronchopulmonary dysplasia. *Nat Rev Dis Prim*. 2019;5:78.
59. Haneline LS, Marshall KP, Clapp DW. The highest concentration of primitive hematopoietic progenitor cells in cord blood is found in extremely premature infants. *Pediatr Res*. 1996;39:820–5.
60. Wisgrill L, Schüller S, Bammer M, Berger A, Pollak A, Radke TF, et al. Hematopoietic stem cells in neonates: any differences between very preterm and term neonates? *PLoS One*. 2014;9:e106717.
61. Makley AT, Goodman MD, Belizaire RM, Friend LAW, Johannigman JA, Dorlac WC, et al. Damage control resuscitation decreases systemic inflammation after hemorrhage. *J Surg Res*. 2012;175:e75–82.
62. Rajnik M, Salkowski CA, Thomas KE, Li Y-Y, Rollwagen FM, Vogel SN. Induction of early inflammatory gene expression in a murine model of nonresuscitated, fixed-volume hemorrhage. *Shock Augusta Ga*. 2002;17:322–8.
63. Makley AT, Goodman MD, Friend LAW, Deters JS, Johannigman JA, Dorlac WC, et al. Resuscitation with fresh whole blood ameliorates the inflammatory response after hemorrhagic shock. *J Trauma*. 2010;68:305–11.
64. Carroll PD, Widness JA. Nonpharmacological, blood conservation techniques for preventing neonatal anemia—effective and promising strategies for reducing transfusion. *Semin Perinatol*. 2012;36:232–43.
65. Hellström W, Forssell L, Morsing E, Sävman K, Ley D. Neonatal clinical blood sampling led to major blood loss and was associated with bronchopulmonary dysplasia. *Acta Paediatr*. 2020;109:679–87.
66. Katheria AC. Neonatal resuscitation with an intact cord: current and ongoing trials. *Children* 2019;6:60.
67. Kuehne B, Kirchgaessner C, Becker I, Kuckelkorn M, Valter M, Kribs A, et al. Mask continuous positive airway pressure therapy with simultaneous extrauterine placental transfusion for resuscitation of preterm infants—a preliminary study. *Biomed Hub*. 2018;3:1–10.
68. Christensen RD, Lambert DK, Baer VL, Montgomery DP, Barney CK, Coulter DM, et al. Postponing or eliminating red blood cell transfusions of very low birth weight neonates by obtaining all baseline laboratory blood tests from otherwise discarded fetal blood in the placenta. *Transfus (Paris)*. 2011;51:253–8.
69. Carroll PD, Christensen RD. New and underutilized uses of umbilical cord blood in neonatal care. *Matern Health Neonatol Perinatol*. 2015;1:16.
70. Rosebraugh MR, Widness JA, Nalbant D, Veng-Pedersen P. A mathematical modeling approach to quantify the role of phlebotomy losses and need for transfusions in neonatal anemia. *Transfus (Paris)*. 2013;53:1353–60.
71. Rabe H, Wacker A, Hülskamp G, Hörnig-Franz I, Schülze-Everding A, Harms E, et al. A randomised controlled trial of delayed cord clamping in very low birth weight preterm infants. *Eur J Pediatr*. 2000;159:775–7.
72. Rabe H, Alvarez J, Lawn C, Seddon P, Amess P. A management guideline to reduce the frequency of blood transfusion in very-low-birth-weight infants. *Am J Perinatol*. 2009;26:179–83.