Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309

p. 4

ABOUT THE TEST FoundationOne®Heme is a comprehensive genomic profiling test designed to identify genomic alterations within hundreds of cancer-related genes in hematologic malignancies, sarcomas, and pediatric cancers.

PATIENT

DISEASE B-lymphoblastic leukemia-lymphoma (B-ALL)
NAME

DATE OF BIRTH

MEDICAL RECORD #

PHYSICIAN

ORDERING PHYSICIAN MEDICAL FACILITY ADDITIONAL RECIPIENT MEDICAL FACILITY ID PATHOLOGIST

SPECIMEN

SPECIMEN SITE SPECIMEN ID SPECIMEN TYPE DATE OF COLLECTION SPECIMEN RECEIVED

BIOMARKER FINDINGS	ACTIONABILITY		
Microsatellite status - MS-Stable	No therapies or clinical trials. see Biomarker Findings section		
Tumor Mutational Burden - TMB-Low (1 Muts/Mb)	No therapies or clinical trials. see Biomarker Findings section		
GENOMIC FINDINGS	THERAPIES WITH CLINICAL BENEFIT (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL BENEFIT (IN OTHER TUMOR TYPE)	
ABL1 - BCR-ABL1 fusion (p210)	Dasatinib	Bosutinib	
	Imatinib	Nilotinib	
4 Trials see p. 7	Ponatinib		

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

RUNX1 - R166Q

NOTE Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type.

TUMOR TYPE B-lymphoblastic leukemia-lymphoma (B-ALL)

TRF#

REPORT DATE

Biomarker Findings

PATIENT

Microsatellite status - MS-Stable Tumor Mutational Burden - TMB-Low (1 Muts/Mb)

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

ABL1 BCR-ABL1 fusion (p210) RUNX1 R166Q

- 5 Therapies with Clinical Benefit0 Therapies with Lack of Response
- 4 Clinical Trials

BIOMARKER FINDINGS

BIOMARKER Microsatellite status

_{сатедоку} MS-Stable

TRF#

POTENTIAL TREATMENT STRATEGIES

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors ¹⁻³, including approved therapies nivolumab and pembrolizumab ⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases $(70\% \text{ vs. } 12\%, p=0.001)^5$.

FREQUENCY & PROGNOSIS

High MSI (MSI-H) is generally rare in hematologic malignancies compared with solid

tumors. Moreover, reports of MSI in hematologic malignancies in the literature are conflicting and varied due to substantial heterogeneity, lack of consensus on the markers and methods used for MSI assessment, small sample size in most studies, and possible elimination of MSI-positive cells in the bloodstream by immunosurveillance ⁶. MSI at any level has been reported at variable levels in samples from patients with acute lymphoblastic leukemia (ALL), including 12% (3/24) to 38% (6/16) of common ALL ⁷⁻⁸, 4.2% (4/96) to 14% (5/36) of B-cell ALL (B-ALL) 9-10, and 9% (1/11) to 33% (2/6) of T-cell ALL (T-ALL) 10-12. In pediatric patients with ALL, MSI at any level has been observed in 2.5% (1/40) to 12.5% (4/32) of common ALL 13-14, 10% (1/ 10) to 33% (3/6) of T-ALL ^{11,14-15}, and 6% (4/63) of B-ALL 9 or absent in B-ALL 11,14. MSI-H has been reported in 4% (1/24) to 19% (3/16) of ALL 7-8, 8% (3/36) of B-ALL 10, and 9% (1/11) of T-ALL ¹⁰. Several studies have reported an

association between increased MSI and relapsed ALL after chemotherapy ^{13,15-18}; however, one study did not ⁹.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor ¹⁹. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁹⁻²¹. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers ²²⁻²⁴. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins 19,21,23-24.

PATIENT

REPORT DATE

BIOMARKER FINDINGS

BIOMARKER Tumor Mutational Burden

TRF#

category TMB-Low (1 Muts/Mb)

POTENTIAL TREATMENT STRATEGIES

On the basis of emerging clinical evidence, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-CTLA-4²⁵, anti-PD-L1²⁶⁻²⁹, and anti-PD-1 therapies 4,30-31; FDA-approved agents include ipilimumab, atezolizumab, avelumab, durvalumab, pembrolizumab, and nivolumab. In multiple solid tumor types, higher mutational burden has corresponded with response and improved prognosis. Pembrolizumab improved progression-free survival (14.5 vs. 3.4-3.7 months) for patients with non-small cell lung cancer (NSCLC) and higher mutational load (greater than 200 nonsynonymous mutations; hazard ratio = 0.19)30. In studies of patients with either NSCLC or colorectal cancer (CRC), patients whose tumors harbored elevated mutational burden reported higher overall response rates to pembrolizumab ^{4,30-31}. Anti-PD-1 therapies have achieved clinical benefit for certain patients with high mutational burden, including 3 patients with endometrial adenocarcinoma who reported sustained partial responses (PRs) following treatment

with pembrolizumab 32 or nivolumab 33, a patient with hypermutant glioblastoma who obtained clinical benefit from pembrolizumab 34, 2 pediatric patients with biallelic mismatch repair deficiency-associated ultrahypermutant glioblastoma who experienced clinically and radiologically significant responses to nivolumab 35, and 2 patients with microsatellite-stable rectal cancers, 1 who achieved an ongoing PR to pembrolizumab and the other an ongoing complete response to nivolumab 36. For patients with melanoma, mutational load was associated with long-term clinical benefit from ipilimumab 25,37 and anti-PD-1/anti-PD-L1 treatments ²⁷. For patients with metastatic urothelial carcinoma (mUC), those who responded to atezolizumab treatment had a significantly increased mutational load (12.4 mutations [muts] per megabase [Mb]) compared to nonresponders (6.4 muts/Mb)²⁶, and mutational load of 16 muts/Mb or higher was associated with significantly longer overall survival 28. In a retrospective analysis of 17 solid tumor types (comprised of 47% NSCLC, 40% mUC, and 13% encompassing 15 other solid tumors), a TMB of ≥16 muts/Mb associated with an objective response rate to atezolizumab of 30% vs. 14% for chemotherapy alone³⁸.

FREQUENCY & PROGNOSIS

Acute lymphocytic leukemia (ALL) harbors a median TMB of 1.7 mutations per megabase (muts/Mb), and 1.3% of cases have high TMB

(>20 muts/Mb)³⁹. Reports of high TMB are generally rare in leukemia ³⁹. In a study of 92 patients with various hematologic malignancies, elevated TMB levels (>10 muts/ Mb) were not detected in AML (o/5) or ALL (o/1) cases analyzed⁴⁰. Increased TMB was observed for some patients at relapse in a study of pediatric ALL ⁴¹.

FINDING SUMMARY

Tumor mutational burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma 42-43 and cigarette smoke in lung cancer 30,44, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes 45-49, and microsatellite instability (MSI) ^{45,48-49}. This sample harbors a low TMB. Compared to patients with tumors harboring higher TMB levels, patients with tumors harboring low TMB levels have experienced lower rates of clinical benefit from treatment with immune checkpoint inhibitors, including anti-CTLA-4 therapy in melanoma 25, anti-PD-L1 therapy in urothelial carcinoma ²⁶, and anti-PD-1 therapy in non-small cell lung cancer and colorectal cancer 4,30.



PATIENT

TRF#

GENOMIC FINDINGS

GENE ABL1 ALTERATION

BCR-ABL1 fusion (p210)

POTENTIAL TREATMENT STRATEGIES

The BCR-ABL fusion protein is the best studied ABL1 alteration and results in an activated ABL1 alteration and results in an activated ABL1 have focused on BCR-ABLpositive hematological malignancies ⁵⁰⁻⁵¹. Tyrosine kinase inhibitors such as imatinib, nilotinib, dasatinib, ponatinib, and bosutinib are FDA approved for the treatment of hematological malignancies with BCR-ABL fusions. Treatment with these therapies has been correlated with increased responses for patients with ALL or CML, compared to

gene **RUNX1** alteration R166Q

POTENTIAL TREATMENT STRATEGIES

There are no therapies available to directly target inactivating alterations in RUNX1. Limited clinical⁸²⁻⁸³ and preclinical⁸⁴ data suggest that RUNX1 alterations, rearrangements in particular, may be associated with sensitivity to DNMT inhibitors, such as the approved agents azacitidine and decitabine. However, multiple clinical studies have reported that RUNX1 is not a significant biomarker for efficacy of these therapies^{82,85-87}. Similarly, on the basis of limited clinical ⁸⁸ and preclinical ⁸⁹⁻⁹¹ evidence,

treatment regimens lacking tyrosine kinase inhibitors ⁵²⁻⁵⁶.

FREQUENCY & PROGNOSIS

The p210 BCR-ABL fusion has been predominantly reported in CML patients, as opposed to p190 present in ALL patients, and has been associated with higher BCR-ABL transcript expression than p190 and with transformation of CML to blast crisis; however, p210 Ph+ ALL cases have also been reported ⁵⁷⁻⁶¹. The Philadelphia chromosome (Ph+) is present in 95% of chronic myeloid leukemia (CML) patients, as well as patients with acute lymphoblastic leukemia (ALL) and <5% of acute myelogenous leukemia (AML) cases; Ph+ ALL accounts for 2-5% of pediatric patients, 20-40% of adults, and >50% of individuals over the age of 50 58-59,62-69. Clinical studies have failed to detect consistent

RUNX1 rearrangements may predict sensitivity to HDAC inhibitors. However, further studies are required to establish clinical significance.

FREQUENCY & PROGNOSIS

RUNX1 mutations have been reported in 3% of all acute lymphoblastic leukemias (ALLs) analyzed in COSMIC; they were detected in 1.5% of B-cell ALL cases and in 6% of T-cell ALL cases (COSMIC, Sep 2018). In one study of 90 adult patients with T-cell ALL, RUNX1 mutations were reported in 15.5% of cases ⁹². In patients with T-cell ALL, RUNX1 mutations have been associated with poorer overall survival ⁹².

FINDING SUMMARY

RUNX1 encodes a transcription factor that is involved in developmental gene expression

difference in responses of ALL patients with p190 versus p210 to imatinib or allogeneic transplantation ^{60,66,70-74}, although residual p190 transcript level has been associated with higher risk of relapse after transplantation than residual p210 ⁷⁵⁻⁷⁶.

FINDING SUMMARY

ABL1 encodes the Abelson protein tyrosine kinase, which is involved in cell growth and survival ⁷⁷. Activating alterations in ABL1 kinase have been reported in leukemia, including the BCR-ABL1 translocation carried on the Philadelphia chromosome in chronic myelogenous leukemia (CML) ^{51,78}. The fusion reported here is similar to the oncogenic p210 BCR-ABL fusion that is found in 64% of patients with CML ⁷⁹⁻⁸¹.

programs and hematopoiesis. It is a frequent site of translocation and mutation in myeloid cancers, and it functions as a tumor suppressor in this context 93-94. Reports of RUNX1 translocations and mutations in myelodysplastic syndrome (MDS) and acute myeloid leukemia (AML) are common. RUNX1 plays a context-dependent role in epithelial cells and has been implicated as both a tumor suppressor and oncogene in different types of solid tumors ⁹⁵. Mutations at R166, more frequently described as R139 based on an alternative transcript, including R139Q and R139G, have been characterized as inactivating, and R139Q has been reported as a germline mutation in familial myeloid leukemia predisposition syndromes 96-100. Other changes at this position are predicted to be inactivating as well.



THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

Dasatinib

Assay findings association

ABL1 BCR-ABL1 fusion (p210)

AREAS OF THERAPEUTIC USE

Dasatinib is a kinase inhibitor that targets the BCR-ABL fusion protein, SRC family kinase receptors, KIT, EPHA2, and PDGFR-beta. It is FDA approved for the treatment of certain subtypes of Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) and Ph+ acute lymphoblastic leukemia (ALL).

GENE ASSOCIATION

Activating ABL1 fusions may predict sensitivity to dasatinib.

SUPPORTING DATA

Dasatinib is approved for Ph+ ALL based on a Phase 2 study that demonstrated major hematologic responses (MaHRs) for 42% (15/36) of imatinib-resistant or intolerant patients^{62,101}. As first-line therapy for Ph+ ALL, dasatinib combined with HCVAD (alternating hyperfractionated cyclophosphamide, vincristine, doxorubicin, and dexamethasone and high-dose cytarabine and methotrexate) followed by maintenance therapy achieved a complete remission rate (CRR) of 96% (69/72), median disease-free survival of 31 months, and median OS of 47 months¹⁰²⁻¹⁰³. Analysis of Phase 2 data for patients with newly diagnosed Ph+ ALL suggests that ponatinib plus HCVAD may result in superior outcomes, including longer OS, compared to dasatinib plus HCVAD¹⁰⁴. Patients with relapsed Ph+ ALL or blastphase CML experienced a 3-year OS rate of 26% and 70%, respectively, and a combined CRR of 71% (24/34) on dasatinib plus HCVAD¹⁰⁵. In the Children's Oncology Group (COG) trial AALLo622, newly diagnosed pediatric patients with Ph+ ALL treated with a combination of dasatinib and intensive chemotherapy exhibited 5-year OS and event-free survival (EFS) rates of 86% and 60%, respectively; patients with IKZF1 deletion experienced inferior OS (80% vs. 100%; p= 0.04) and EFS (52% vs. 82%; p=0.04)¹⁰⁶. Five-year OS did not significantly differ between patients with Ph+ ALL receiving chemotherapy in conjunction with dasatinib in COG trial AALLo622 (86%)¹⁰⁶ or imatinib in COG trial AALL0031 (81%)¹⁰⁷. Post-transplant maintenance therapy with tyrosine kinase inhibitors, including dasatinib, is associated with improved outcomes for patients with Ph+ ALL in retrospective studies¹⁰⁸⁻¹⁰⁹. For elderly patients with newly diagnosed Ph+ ALL, dasatinib plus low-intensity chemotherapy achieved a CRR of 96% and a 5-year OS rate of 36%¹¹⁰⁻¹¹¹. A Phase 1 trial of dasatinib for pediatric patients with relapsed or refractory leukemia observed MaHRs for 47% (8/17) of cases with advanced-phase CML or Ph+ ALL and a median MaHR duration of 4.4 months¹¹². Case reports describe pediatric or young adult patients with ABL1-positive T-cell ALL (ABL1 amplification or NUP214-ABL1 fusion) who achieved remissions on dasatinib alone or combined with chemotherapy113-114.

Imatinib

Assay findings association

ABL1 BCR-ABL1 fusion (p210)

AREAS OF THERAPEUTIC USE

Imatinib targets the BCR-ABL fusion protein, PDGFR, and KIT. It is FDA approved for the treatment of KITpositive gastrointestinal stromal tumors (GIST), Ph+ chronic myeloid leukemia (CML) and acute lymphoblastic leukemia (ALL), myelodysplastic syndrome/ myeloproliferative syndrome (MDS/MPS), aggressive systemic mastocytosis without a D816V KIT mutation, hypereosinophilic syndrome and/or chronic eosinophilic leukemia, and dermatofibrosarcoma protuberans.

GENE ASSOCIATION

Activating ABL1 fusions may predict sensitivity to imatinib.

SUPPORTING DATA

In clinical studies, patients with multiple stages of CML or BCR-ABL-positive ALL were able to achieve hematological and sometimes cytological remission following imatinib treatment^{52,115-116}. Responses to imatinib have also been reported in ALL patients harboring ETV6-ABL1 or NUP214-ABL1 fusions¹¹⁷⁻¹¹⁸.

Ponatinib

Assay findings association

ABL1 BCR-ABL1 fusion (p210)

AREAS OF THERAPEUTIC USE

Ponatinib is a multikinase inhibitor targeting BCR-ABL, RET, KIT, FLT-3, PDGFRs, VEGFRs, FGFRs, and other tyrosine kinases. It is FDA approved for the treatment of advanced, T315I-mutated chronic myeloid leukemia (CML) and Philadelphia chromosome-positive (Ph+) acute lymphoblastic leukemia (ALL), as well as for CML and Ph+ ALL patients for whom no other tyrosine kinase inhibitor is indicated.

GENE ASSOCIATION

Activating fusions in ABL1 may predict sensitivity to ponatinib.

SUPPORTING DATA

A Phase 2 study reported major hematologic and cytogenetic responses in 41% (13/32) and 47% (15/32), respectively, of patients with Ph+ ALL¹¹⁹. In particular, among the 22 Ph+ ALL patients with the T315I mutation treated with ponatinib, 36% had a major hematologic response, 41% had a major cytogenetic response, and 32% had a complete cytogenetic response¹¹⁹. In a Phase 2 study of ponatinib in combination with chemotherapy (HCVAD, high-dose MTX/Ara-C) as first-line therapy for patients with Ph+ ALL, the 3-year complete response duration and overall survival rate were 79% and 82%, respectively; at the time of reporting, 96% (50/52) of patients had achieved minimum residual disease¹²⁰.



THERAPIES WITH CLINICAL BENEFIT IN OTHER TUMOR TYPE

REPORT DATE

Bosutinib

Assay findings association

ABL1 BCR-ABL1 fusion (p210)

Nilotinib

ABL1

Assay findings association

BCR-ABL1 fusion (p210)

AREAS OF THERAPEUTIC USE

Bosutinib is a third-generation tyrosine kinase inhibitor that targets both ABL and SRC kinases. It is FDA approved to treat newly diagnosed chronic phase Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) or chronic, accelerated, or blast phase Ph+ CML with resistance or intolerance to prior therapy.

GENE ASSOCIATION

Activating ABL1 fusions may predict sensitivity to bosutinib.

SUPPORTING DATA

For patients with acute lymphoblastic leukemia (ALL) treated with bosutinib, 9% (2/22) had a hematologic response¹²¹. In a Phase 1 trial combining the CD22 antibody inotuzumab ozogamicin and bosutinib for Ph+, T315I-negative ALL or lymphoid blast phase CML, 11/14 patients achieved a complete response (CR) or CR with incomplete blood count recovery (CRi); of the 11

responders, 10 (91%) achieved complete cytogenetic remission (CCyR), and 6 (55%) had undetectable BCR-ABL¹²². A retrospective analysis reported that a heavily pretreated patient with Ph+ ALL and an ABL1 V299L mutation achieved a CR and eventual minimal residual disease (MRD) negativity from the combination of bosutinib and the CD3/CD19 bispecific antibody blinatumomab¹²³. Bosutinib facilitated remissions in clinical studies of CML, including a 73% response rate for those who had failed prior imatinib or nilotinib therapies¹²⁴⁻¹²⁷. Two Phase 3 studies for patients with newly diagnosed, chronic phase CML (CP-CML) reported significantly higher rates of major molecular response (MMR) at 12 months following treatment with bosutinib versus imatinib (BELA, 41% vs. 27%; and BFORE, 47.2% and 36.9%)128-129 . An elderly patient whose CML transformed to B-ALL achieved a complete hematologic response after 14 months of fourth-line bosutinib treatment¹³⁰.

AREAS OF THERAPEUTIC USE

Nilotinib targets tyrosine kinases such as ABL (including BCR-ABL), PDGFRs, KIT, CSF1R, DDR1, and DDR2. It is FDA approved to treat newly diagnosed pediatric or adult patients with Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) in chronic phase, adults with Ph+ CML in chronic or accelerated phase with resistance or intolerance to prior therapy including imatinib, and pediatric patients with Ph+ CML in chronic phase with resistance or intolerance to prior tyrosine-kinase inhibitor therapy.

GENE ASSOCIATION

Activating fusions in ABL1 may predict sensitivity to nilotinib.

SUPPORTING DATA

In a Phase 2 study of the effects of nilotinib combined with chemotherapy in patients with Ph+ acute lymphoblastic leukemia (ALL), 91% of patients achieved complete hematologic remission (HCR); the 2-year hematologic relapse-free survival (HRFS) rate was 72% for 82 patients that achieved HCR, and the 2-year overall survival rate was 72%¹³¹. Nilotinib combined with chemotherapy was well tolerated and highly effective with a 97% complete response (CR) in elderly patients with newly diagnosed Ph+ ALL in another Phase 2 trial¹³². In a Phase 1 study of nilotinib in patients with imatinib-resistant Ph+ CML or ALL, 15.4% (2/13) patients with Ph+ ALL responded to nilotinib¹³³.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however the agents listed in this report may have little or no evidence in the patient's tumor type.



PATIENT

TUMOR TYPE B-lymphoblastic leukemia-lymphoma (B-ALL) REPORT DATE

TRF#

GENE

ABL1

ALTERATION

CLINICAL TRIALS

IMPORTANT Clinical trials are ordered by gene and
prioritized by: age range inclusion criteria for pediatric
patients, proximity to ordering medical facility, later trial
phase, and verification of trial information within the last
two months. While every effort is made to ensure the
accuracy of the information contained below, the
information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial \rightarrow Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit https://www.foundationmedicine.com/genomictesting#support-services.

RATIONALE

Therapies that inhibit the kinase activity of ABL may be appropriate for a patient with an activating ABL1 fusion.

BCR-ABL1 fusion (p210)	activating replit fusion.	
NCT02081378		PHASE 1
A Phase I, Multicenter, Open-label Study of Oral AB Leukemia (CML) or Philadelphia Chromosome-pos	, .	targets ABL, BCR-ABL, KIT, PDGFRs, DDR2, SRC

LOCATIONS: Paris Cedex 10 (France), Kobe-shi (Japan), Massachusetts, Michigan, New York, Oregon, Roma (Italy), Seoul (Korea, Republic of), Adelaide (Australia), Texas, Utah, Bordeaux (France), Berlin (Germany), Frankfurt (Germany), Jena (Germany), Amsterdam (Netherlands), Singapore (Singapore), Madrid (Spain)

NCT03263572	PHASE 2
Phase II Study of the Combination of Blinatumomab and Ponatinib in Patients With Philadelphia Chromosome (Ph)-Positive and/or BCR-ABL Positive Acute Lymphoblastic Leukemia (ALL)	TARGETS ABL, FGFRs, FLT3, KIT, PDGFRs, RET, VEGFRs

LOCATIONS: Texas

NCT02420717	PHASE 2
A Phase II Study of the Combination of Ruxolitinib or Dasatinib With Chemotherapy in Patients With Philadelphia Chromosome (Ph)-Like Acute Lymphoblastic Leukemia (ALL)	targets CD20, JAK2, JAK1, ABL, DDR2, KIT, PDGFRs, SRC
LOCATIONS: Texas	
NCT01424982	PHASE 2

LOCATIONS: Texas



APPENDIX Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BRCA2	CIITA	CREBBP	ETS1
A2351G	P699L and V642I	F1358S	V13L
GPR124	IKZF3	IRS2	KMT2C (MLL3)
C1196Y	T190M	A512T	S4300P
MSH6	NSD1	NUP98	PDGFRB
K1358fs*2	S822C	P1085L	A366T
РТСН1	PTPRO	STAT4	YY1AP1
К838Т	R480W	L307F	C46_R48del



APPENDIX

Genes Assayed in FoundationOne®Heme

REPORT DATE

FoundationOne Heme is designed to include genes known to be somatically altered in human hematologic malignancies, sarcomas, and pediatric cancers that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay utilizes DNA sequencing to interrogate 406 genes as well as selected introns of 31 genes involved in rearrangements, in addition to RNA sequencing of 265 genes. The assay will be updated periodically to reflect new knowledge about cancer biology.

HEMATOLOGICAL MALIGNANCY DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	S, INSERTION/D	ELETIONS, AND C AKT1	AKT2	AKT3	ALK	AMER1 (FAM123B		APC
ADLI APH1A	ACTB	ARAF	ARFRP1	ARHGAP26 (GRAF)		ARID1A	ARID2	ASMTL
ASXL1	AK ATM	ARAF ATR	ATRX	AURKA	, AURKB	AXIN1 AXIN1	AXL	B2M
BAP1	BARD1	BCL10	BCL11B	BCL2	BCL2L2	BCL6	BCL7A	BZINI BCOR
BCORL1	BIRC3	BLM	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BRSK1
BTG2	BTK	BTLA	C11orf30 (EMSY)	CAD	CALR*	CARD11	CBFB	CBL
CCND1	CCND2	CCND3	CCNE1	ССТ6В	CD22	CD274 (PD-L1)	CD36	CD58
CD70	CD79A	CD79B	CDC73	CDH1	CDK12	CDK4	CDK6	CDK8
CDKN1B	CDKN2A	CDKN2B	CDKN2C	CEBPA	CHD2	CHEK1	CHEK2	CIC
CIITA	CKS1B	CPS1	CREBBP	CRKL	CRLF2	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUX1	CXCR4	DAXX	DDR2	DDX3X	DNM2	DNMT3A
DOT1L	DTX1	DUSP2	DUSP9	EBF1	ECT2L	EED	EGFR	ELP2
EP300	EPHA3	EPHA5	EPHA7	EPHB1	ERBB2	ERBB3	ERBB4	ERG
ESR1	ETS1	ETV6	EXOSC6	EZH2	FAF1	FAM46C	FANCA	FANCC
FANCD2	FANCE	FANCF	FANCG	FANCL	FAS (TNFRSF6)	FBXO11	FBXO31	FBXW7
FGF10	FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2
FGFR3	FGFR4	FHIT	FLCN	FLT1	FLT3	FLT4	FLYWCH1	FOXL2
FOXO1	FOXO3	FOXP1	FRS2	GADD45B	GATA1	GATA2	GATA3	GID4 (C17orf39)
GNA11	GNA12	GNA13	GNAQ	GNAS	GPR124	GRIN2A	GSK3B	GTSE1
HDAC1	HDAC4	HDAC7	HGF	HIST1H1C	HIST1H1D	HIST1H1E	HIST1H2AC	HIST1H2AG
HIST1H2AL	HIST1H2AM	HIST1H2BC	HIST1H2BJ	HIST1H2BK	HIST1H2BO	HIST1H3B	HNF1A	HRAS
HSP90AA1	ICK	ID3	IDH1	IDH2	IGF1R	IKBKE	IKZF1	IKZF2
IKZF3	IL7R	INHBA	INPP4B	INPP5D (SHIP)	IRF1	IRF4	IRF8	IRS2
JAK1	JAK2	JAK3	JARID2	JUN	KAT6A (MYST3)	KDM2B	KDM4C	KDM5A
KDM5C	KDM6A	KDR	KEAP1	KIT	KLHL6	KMT2A (MLL)	KMT2C (MLL3)	KMT2D (MLL2)
KRAS	LEF1	LRP1B	LRRK2	MAF	MAFB	MAGED1	MALT1	MAP2K1
MAP2K2	MAP2K4	MAP3K1	MAP3K14	МАРЗК6	MAP3K7	ΜΑΡΚ1	MCL1	MDM2
MDM4	MED12	MEF2B	MEF2C	MEN1	MET	MIB1	MITF	MKI67
MLH1	MPL	MRE11A	MSH2	MSH3	MSH6	MTOR	МИТҮН	МҮС
MYCL (MYCL1)	MYCN	MYD88	MYO18A	NCOR2	NCSTN	NF1	NF2	NFE2L2
NFKBIA	NKX2-1	NOD1	NOTCH1	NOTCH2	NPM1	NRAS	NT5C2	NTRK1
NTRK2	NTRK3	NUP93	NUP98	P2RY8	PAG1	РАКЗ	PALB2	PASK
PAX5	PBRM1	РС	PCBP1	PCLO	PDCD1	PDCD11	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PHF6	РІКЗСА	PIK3CG	PIK3R1	PIK3R2	PIM1	PLCG2
POT1	PPP2R1A	PRDM1	PRKAR1A	PRKDC	PRSS8	PTCH1	PTEN	PTPN11
PTPN2	PTPN6 (SHP-1)	PTPRO	RAD21	RAD50	RAD51	RAF1	RARA	RASGEF1A
RB1	RELN	RET	RHOA	RICTOR	RNF43	ROS1	RPTOR	RUNX1
S1PR2	SDHA	SDHB	SDHC	SDHD	SERP2	SETBP1	SETD2	SF3B1
SGK1	SMAD2	SMAD4	SMARCA1	SMARCA4	SMARCB1	SMC1A	SMC3	SMO
SOCS1	SOCS2	SOCS3	SOX10	SOX2	SPEN	SPOP	SRC	SRSF2
STAG2	STAT3	STAT4	STAT5A	STAT5B	STAT6	STK11	SUFU	SUZ12
TAF1	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TET2	TGFBR2	TLL2	ТМЕМЗОА	
TMSB4XP8 (TMSI		TNFAIP3	TNFRSF11A	TNFRSF14	TNFRSF17	TOP1	TP53	TP63
TRAF2	TRAF3	TRAF5	TSC1	TSC2	TSHR	TUSC3	TYK2	U2AF1
U2AF2	VHL	WDR90	WHSC1 (MMSET or		WISP3	WT1	XBP1	XPO1
YY1AP1	ZMYM3	ZNF217	ZNF24 (ZSCAN3)		ZRSR2			
	2	2.11217	211127 (230/113)	2.11.7.05	LIGHL			

*Note: the assay was updated on 11/8/2016 to include the detection of alterations in CALR

Electronically signed by Eric Severson, M.D., Ph.D., M.M.Sc. | Jeffrey Ross, M.D., Medical Director, , M.D., Ph.D., M.M.Sc. | 08 February 2019 | Foundation Medicine, Inc. | 1.888.988.3639



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Genes Assayed in FoundationOne®Heme

HEMATOLOGICA	L MALIGNANC	Y DNA GENE LIST	FOR THE DET	ECTION OF SELECT	REARRANGEM	ENTS		
ALK	BCL2	BCL6	BCR	BRAF	CCND1	CRLF2	EGFR	EPOR
ETV1	ETV4	ETV5	ETV6	EWSR1	FGFR2	IGH	IGK	IGL
IAK1	JAK2	KMT2A (MLL)	МҮС	NTRK1	PDGFRA	PDGFRB	RAF1	RARA
RET	ROS1	TMPRSS2	TRG					
HEMATOLOGICA		Y RNA GENE LIST	FOR THE DET		REARRANGEM	ENTS		
ABI1	ABL1	ABL2	ACSL6	AFF1	AFF4	ALK	ARHGAP26 (GR/	AF)
ARHGEF12	ARID1A	ARNT	ASXL1	ATF1	ATG5	ATIC	BCL10	BCL11A
BCL11B	BCL2	BCL3	BCL6	BCL7A	BCL9	BCOR	BCR	BIRC3
BRAF	BTG1	CAMTA1	CARS	CBFA2T3	CBFB	CBL	CCND1	CCND2
CCND3	CD274 (PD-L1)	CDK6	CDX2	CHIC2	CHN1	CIC	CIITA	CLP1
CLTC	CLTCL1	CNTRL (CEP110)	COL1A1	CREB3L1	CREB3L2	CREBBP	CRLF2	CSF1
CTNNB1	DDIT3	DDX10	DDX6	DEK	DUSP22	EGFR	EIF4A2	ELF4
ELL	ELN	EML4	EP300	EPOR	EPS15	ERBB2	ERG	ETS1
TV1	ETV4	ETV5	ETV6	EWSR1	FCGR2B	FCRL4	FEV	FGFR1
GFR1OP	FGFR2	FGFR3	FLI1	FNBP1	FOXO1	FOXO3	FOXO4	FOXP1
FSTL3	FUS	GAS7	GLI1	GMPS	GPHN	HERPUD1	HEY1	HIP1
HIST1H4I	HLF	HMGA1	HMGA2	HOXA11	HOXA13	НОХАЗ	НОХА9	HOXC11
HOXC13	HOXD11	HOXD13	HSP90AA1	HSP90AB1	IGH	IGK	IGL	IKZF1
IL21R	IL3	IRF4	ΙΤΚ	JAK1	JAK2	JAK3	JAZF1	KAT6A (MYST
KDSR	KIF5B	KMT2A (MLL)	LASP1	LCP1	LMO1	LMO2	LPP	LYL1
MAF	MAFB	MALT1	MDS2	MECOM	MKL1	MLF1	MLLT1 (ENL)	MLLT10 (AF10,
MLLT3	MLLT4 (AF6)	MLLT6	MN1	MNX1	MSI2	MSN	MUC1	МҮВ
МҮС	MYH11	МҮН9	NACA	NBEAP1 (BCL8)	NCOA2	NDRG1	NF1	NF2
NFKB2	NIN	NOTCH1	NPM1	NR4A3	NSD1	NTRK1	NTRK2	NTRK3
NUMA1	NUP214	NUP98	NUTM2A	OMD	P2RY8	PAFAH1B2	PAX3	PAX5
PAX7	PBX1	PCM1	PCSK7	PDCD1LG2 (PD-L2)	PDE4DIP	PDGFB	PDGFRA	PDGFRB
PER1	PHF1	PICALM	PIM1	PLAG1	PML	POU2AF1	PPP1CB	PRDM1
PRDM16	PRRX1	PSIP1	PTCH1	PTK7	RABEP1	RAF1	RALGDS	RAP1GDS1
RARA	RBM15	RET	RHOH	RNF213	ROS1	RPL22	RPN1	RUNX1
RUNX1T1 (ETO)	RUNX2	SEC31A	SEPT5	SEPT6	SEPT9	SET	SH3GL1	SLC1A2
5NX29 (RUNDC2A)	SRSF3	SS18	SSX1	SSX2	SSX4	STAT6	STL	SYK
TAF15	TAL1	TAL2	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TEC	TET1	TFE3
TFG	TFPT	TFRC	TLX1	TLX3	TMPRSS2	TNFRSF11A	TOP1	TP63
ТРМЗ	TPM4	TRIM24	TRIP11	TTL	ΤΥΚ2	USP6	WHSC1 (MMSET	or NSD2)
WHSC1L1	YPEL5	ZBTB16	ZMYM2	ZNF384	ZNF521			

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Microsatellite (MS) status

Tumor Mutational Burden (TMB)



тимок түре B-lymphoblastic leukemia-lymphoma (B-ALL)

TRF#

APPENDIX

Performance Specifications

The median exon coverage for this sample is 883x				
ACCURACY				
Sensitivity: Base Substitutions	At≥5% Minor Allele Frequency	>99.0%		
Sensitivity: Insertions/Deletions (1-40bp)	At≥10% Minor Allele Frequency	98.0%		
Sensitivity: Focal Copy Number Alterations (Homozygous Deletions or Amplifications)	At≥8% copies	>95.0%		
Sensitivity: Microsatellite status	At≥20% tumor nuclei	97.0%		
Sensitivity: Known Gene Fusions	>95.0%			
Specificity: Base Substitutions, Insertions/Deletions, and Focal Copy Number Alterations	Positive Predictive Value (PPV)	>99.0%		
Specificity: Known Gene Fusions	Positive Predictive Value (PPV)	>95.0%		
Specificity: Microsatellite status	Positive Predictive Value (PPV)	>95.0%		
Accuracy: Tumor Mutation Burden	At≥20% tumor nuclei	>90.0%		
Reproducibility (average concordance between replicates)	97.0% inter-batch precision 97.0% intra-batch precision 95.0% microsatellite status precision 96.0% tumor mutation burden precision			

Assay specifications were determined for pical median exon coverage of approximately 500X. For additional information regarding the validation of FoundationOne, please refer to the article, Frampton, GM. et al. Development and validation of a clinical cancer genomic profiling test based on massively parallel DNA sequencing, Nat Biotechnol (2013 Oct. 20).

Microsatellite status, which is a measure of microsatellite instability (MSI), is determined by assessing indel characteristics at 114 homopolymer repeat loci in or near the targeted gene regions of the FoundationOne Heme test. Microsatellite status is assayed for all FoundationOne Heme samples and may be reported as "MSI-High", "MSI-Intermediate", "MS-Stable", or "Cannot Be Determined". Microsatellite status is reported as "Cannot Be Determined" if the sample is not of sufficient quality to be confidently determined.

Tumor Mutational Burden (TMB) is determined by measuring the number of somatic mutations occurring in sequenced genes on the FoundationOne and FoundationOne Heme tests and extrapolating to the genome as a whole. TMB is assayed for all FoundationOne and FoundationOne Heme samples and may be reported as "TMB-High", "TMB-Intermediate", "TMB-Low", or "Cannot Be Determined". TMB results, which are rounded to the nearest integer, are determined as follows: TMB-High corresponds to greater than or equal to 20 mutations per megabase (Muts/Mb); TMB-Intermediate corresponds to 6-19 Muts/Mb; TMB-Low corresponds to fewer than or equal to 5 Muts/Mb. Tumor Mutational Burden is reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine Tumor Mutational Burden.

For additional information specific to the performance of this specimen, please contact Foundation Medicine, Inc. at 1-888-988-3639.

FOUNDATIONONE®HEME

APPENDIX

About FoundationOne®Heme

TRF#

ABOUT FOUNDATIONONE HEME

FoundationOne Heme is a comprehensive genomic profiling test for hematologic malignancies, sarcomas and pediatric cancers. The test is designed to provide physicians with clinically actionable information to help with diagnostic subclassification, prognosis assessment, and targeted therapeutic selection. Test results provide information about clinically significant alterations, potential targeted therapies, available clinical trials, and quantitative markers that may support immunotherapy clinical trial enrollment. FoundationOne Heme is analytically validated to detect all classes of genomic alterations in more than 400 cancer-related genes. In addition to DNA sequencing, FoundationOne Heme employs RNA sequencing across more than 250 genes to capture a broad range of gene fusions, common drivers of hematologic malignancies and sarcomas, pediatric cancers.

FoundationOne Heme was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Heme has not been cleared or approved by the United States Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. FoundationOne Heme may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high-complexity clinical testing.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne Heme identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

PATIENT

An alteration denoted as "amplification - equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne Heme for identifying a copy number amplification is five (5) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that FoundationOne Heme analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Alterations and Therapies Biomarker Findings Appear at the top of the report, but are not ranked

higher than Genomic Findings.

Genomic Findings

Therapies with Clinical Benefit In Patient's Tumor Type \rightarrow Therapies with Clinical Benefit in Other Tumor Type \rightarrow Clinical Trial Options \rightarrow No Known Options (If multiple findings exist within any of these categories, the results are listed alphabetically by gene name.)

Therapies

Sensitizing therapies → Resistant therapies. (If multiple therapies exist within any of these categories, they are listed alphabetically by therapy name.)

Clinical Trials

Pediatric trial qualification \rightarrow Geographical proximity \rightarrow Later trial phase.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Heme.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. These include: subclonal alterations in heterogeneous samples, low sample quality or with homozygous losses of <3 exons; and deletions and insertions >40bp, or in repetitive/high homology sequences. FoundationOne Heme is performed using DNA and RNA derived from tumor, and as such germline events may not be reported.

The following targets typically have low coverage resulting in a reduction in sensitivity: SDHD exon 4, TNFRSF11A exon1, and TP53 exon 1.

FoundationOne Heme complies with all European Union (EU) requirements of the IVD Directive 98/ 79EC. As such, the FoundationOne Heme Assay has been registered for CE mark by our EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.

CE



APPENDIX

About FoundationOne®Heme

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
ткі	Tyrosine kinase inhibitor



APPENDIX

References

- 1 Gatalica Z, Snyder C, Maney T, et al. (2014) Programmed cell death 1 (PD-1) and its ligand (PD-L1) in common cancers and their correlation with molecular cancer type. Cancer Epidemiol. Biomarkers Prev. ePub Dec 2014
- 2 Kroemer G, Galluzzi L, Zitvogel L, et al. (2015) Colorectal cancer: the first neoplasia found to be under immunosurveillance and the last one to respond to immunotherapy? Oncoimmunology 4 (7):e1058597
- 3 Lal N, Beggs AD, Willcox BE, et al. (2015) An immunogenomic stratification of colorectal cancer: Implications for development of targeted immunotherapy. Oncoimmunology 4 (3):e976052
- 4 Le DT, Uram JN, Wang H, et al. (2015) PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N. Engl. J. Med. ePub Jun 2015
- 5 ASCO-SITC 2016; Abstract P60
- 6 Maletzki C, Stier S, Linnebacher M (2013) Microsatellite instability in hematological malignancies: Hypermutation vs. immune controlwho is challenging who? Oncoimmunology 2 (8):e25419
- 7 Pabst T, Schwaller J, Bellomo MJ, et al. (1996) Frequent clonal loss of heterozygosity but scarcity of microsatellite instability at chromosomal breakpoint cluster regions in adult leukemias. Blood 88 (3):1026-34
- 8 Indraccolo S, Minuzzo S, Nicoletti L, et al. (1999) Mutator phenotype in human hematopoietic neoplasms and its association with deletions disabling DNA repair genes and bcl-2 rearrangements. Blood 94 (7):2424-32
- 9 Molenaar JJ, Gérard B, Chambon-Pautas C, et al. (1998) Microsatellite instability and frameshift mutations in BAX and transforming growth factorbeta RII genes are very uncommon in acute lymphoblastic leukemia in vivo but not in cell lines. Blood 92 (1):230-3
- 10 Nomdedéu JF, Perea G, Estivill C, et al. (2008) Microsatellite instability may involve the pentanucleotide repeat of the PIG3 promoter in bcr/ abl acute lymphoblastic leukemia. Leuk. Res. 32 (1):186-8
- 11 Reato G, Basso G, Putti MC, et al. (1998) Microsatellite analysis in childhood acute lymphoblastic leukemia. Haematologica 83 (5):403-7
- 12 Krsková-Honzátková L, Cermák J, Sajdová J, et al. (2002) Microsatellite instability in hematological malignancies. Leuk. Lymphoma 43 (10):1979-86
- 13 Best A, Matheson E, Minto L, et al. (2010) Mismatch repair and the downstream target genes, PAX5 and Ikaros, in childhood acute lymphoblastic leukemia. Leuk. Res. ePub Aug 2010
- 14 Takeuchi S, Seriu T, Tasaka T, et al. (1997) Microsatellite instability and other molecular abnormalities in childhood acute lymphoblastic leukaemia. Br. J. Haematol. 98 (1):134-9
- 15 Baccichet A, Benachenhou N, Couture F, et al. (1997) Microsatellite instability in childhood T cell acute lymphoblastic leukemia. Leukemia 11 (6):797-802
- 16 Yang JJ, Bhojwani D, Yang W, et al. (2008) Genomewide copy number profiling reveals molecular evolution from diagnosis to relapse in childhood acute lymphoblastic leukemia. Blood ePub Nov 2008

- 17 Matheson FC, Hall AG (2003) Assessment of mismatch repair function in leukaemic cell lines and blasts from children with acute lymphoblastic leukaemia. Carcinogenesis 24 (1):31-8
- 18 Kendall HE, Vacek PM, Finette BA (2004) Analysis of microsatellite instability in children treated for acute lymphocytic leukemia with elevated HPRT mutant frequencies. Mutagenesis 19 (5):409-12
- 19 Kocarnik JM, Shiovitz S, Phipps AI (2015) Molecular phenotypes of colorectal cancer and potential clinical applications. Gastroenterol Rep (Oxf) 3 (4):269-76
- 20 You JF, Buhard O, Ligtenberg MJ, et al. (2010) Tumours with loss of MSH6 expression are MSI-H when screened with a pentaplex of five mononucleotide repeats. Br. J. Cancer ePub Dec 2010
- 21 Bairwa NK, Saha A, Gochhait S, et al. (2014) Microsatellite instability: an indirect assay to detect defects in the cellular mismatch repair machinery. Methods Mol. Biol. ePub 2014
- 22 Boland CR, Thibodeau SN, Hamilton SR, et al. (1998) A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. Cancer Res. 58 (22):5248-57
- 23 Pawlik TM, Raut CP, Rodriguez-Bigas MA (2004) Colorectal carcinogenesis: MSI-H versus MSI-L. Dis. Markers 20 (4-5):199-206
- 24 Boland CR, Goel A (2010) Microsatellite instability in colorectal cancer. Gastroenterology ePub Jun 2010
- 25 Snyder A, Makarov V, Merghoub T, et al. (2014) Genetic basis for clinical response to CTLA-4 blockade in melanoma. N. Engl. J. Med. ePub Dec 2014
- 26 Rosenberg JE, Hoffman-Censits J, Powles T, et al. (2016) Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinumbased chemotherapy: a single-arm, multicentre, phase 2 trial. Lancet ePub May 2016
- 27 Johnson DB, Frampton GM, Rioth MJ, et al. (2016) **Targeted Next Generation Sequencing Identifies** Markers of Response to PD-1 Blockade. Cancer Immunol Res ePub Nov 2016
- 28 Balar AV, Galsky MD, Rosenberg JE, et al. (2017) Atezolizumab as first-line treatment in cisplatinineligible patients with locally advanced and metastatic urothelial carcinoma: a single-arm, multicentre, phase 2 trial. Lancet ePub 01 2017
- 29 Miao D, Margolis CA, Vokes NI, et al. (2018) Genomic correlates of response to immune checkpoint blockade in microsatellite-stable solid tumors. Nat. Genet. ePub Sep 2018
- 30 Rizvi NA, Hellmann MD, Snyder A, et al. (2015) Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science ePub Apr 2015
- 31 Dong ZY, Zhong WZ, Zhang XC, et al. (2017) Clin. Cancer Res. 23 (12):3012-3024
- 32 Mehnert JM, Panda A, Zhong H, et al. (2016) Immune activation and response to pembrolizumab in POLEmutant endometrial cancer. J. Clin. Invest. ePub Jun 2016

- 33 Santin AD, Bellone S, Buza N, et al. (2016) Regression of Chemotherapy-Resistant Polymerase ε (POLE) Ultra-Mutated and MSH6 Hyper-Mutated Endometrial Tumors with Nivolumab. Clin. Cancer Res. 22 (23):5682-5687
- 34 Johanns TM, Miller CA, Dorward IG, et al. (2016) Immunogenomics of Hypermutated Glioblastoma: A Patient with Germline POLE Deficiency Treated with Checkpoint Blockade Immunotherapy. Cancer Discov ePub 11 2016
- 35 Bouffet E, Larouche V, Campbell BB, et al. (2016) Immune Checkpoint Inhibition for Hypermutant Glioblastoma Multiforme Resulting From Germline Biallelic Mismatch Repair Deficiency. J. Clin. Oncol. ePub Jul 2016
- 36 Fabrizio DA, George TJ, Dunne RF, et al. (2018) Beyond microsatellite testing: assessment of tumor mutational burden identifies subsets of colorectal cancer who may respond to immune checkpoint inhibition. J Gastrointest Oncol 9 (4):610-617
- 37 Van Allen EM, Miao D, Schilling B, et al. (2015) Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. Science ePub Oct 2015
- 38 Legrand et al., 2018; ASCO Abstract 12000
- 39 Chalmers ZR, Connelly CF, Fabrizio D, et al. (2017) Analysis of 100,000 human cancer genomes reveals the landscape of tumor mutational burden. Genome Med ePub 04 2017
- 40 Karim et al., 2017; AACR Abstract 3724
- 41 Ding LW, Sun QY, Tan KT, et al. (2017) Mutational Landscape of Pediatric Acute Lymphoblastic Leukemia. Cancer Res. ePub 01 2017
- 42 Pfeifer GP, You YH, Besaratinia A (2005) Mutations induced by ultraviolet light. Mutat. Res. 571 (1-2):19-31
- 43 Hill VK, Gartner JJ, Samuels Y, et al. (2013) The genetics of melanoma: recent advances. Annu Rev Genomics Hum Genet ePub 2013
- 44 Pfeifer GP, Denissenko MF, Olivier M, et al. (2002) Tobacco smoke carcinogens, DNA damage and p53 mutations in smoking-associated cancers. Oncogene 21 (48):7435-51
- 45 Cancer Genome Atlas Research Network, Kandoth C. Schultz N, et al. (2013) Integrated genomic characterization of endometrial carcinoma. Nature ePub May 2013
- 46 Briggs S, Tomlinson I (2013) Germline and somatic polymerase ϵ and δ mutations define a new class of hypermutated colorectal and endometrial cancers. J. Pathol. ePub Jun 2013
- 47 Heitzer E, Tomlinson I (2014) Replicative DNA polymerase mutations in cancer. Curr. Opin. Genet. Dev. ePub Feb 2014
- 48 (2012) Comprehensive molecular characterization of human colon and rectal cancer. Nature ePub Jul 2012
- 49 Roberts SA, Gordenin DA (2014) Hypermutation in human cancer genomes: footprints and mechanisms. Nat. Rev. Cancer ePub 12 2014
- 50 Lim TH, Tien SL, Lim P, et al. (2005) The incidence and patterns of BCR/ABL rearrangements in chronic myeloid leukaemia (CML) using fluorescence in situ hybridisation (FISH). Ann. Acad. Med. Singap. 34 (9):533-8



REPORT DATE

```
APPENDIX References
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- 51 Konopka JB, Watanabe SM, Singer JW, et al. (1985) Cell lines and clinical isolates derived from Ph1-positive chronic myelogenous leukemia patients express c-abl proteins with a common structural alteration. Proc. Natl. Acad. Sci. U.S.A. 82 (6):1810-4
- 52 (2011) Current treatment of Philadelphia chromosome-positive acute lymphoblastic leukemia. Hematology Am Soc Hematol Educ Program ePub 2011
- 53 Santos FP, Quintás-Cardama A (2011) New drugs for chronic myelogenous leukemia. Curr Hematol Malig Rep ePub Jun 2011
- 54 Reddy EP, Aggarwal AK (2012) The ins and outs of bcr-abl inhibition. Genes Cancer 3 (5-6):447-54
- 55 Soverini S, Hochhaus A, Nicolini FE, et al. (2011) BCR-ABL kinase domain mutation analysis in chronic myeloid leukemia patients treated with tyrosine kinase inhibitors: recommendations from an expert panel on behalf of European LeukemiaNet. Blood ePub Aug 2011
- 56 Keller-V Amsberg G, Brümmendorf TH (2012) Novel aspects of therapy with the dual Src and Abl kinase inhibitor bosutinib in chronic myeloid leukemia. Expert Rev Anticancer Ther ePub Sep 2012
- 57 Jones D, Luthra R, Cortes J, et al. (2008) BCR-ABL fusion transcript types and levels and their interaction with secondary genetic changes in determining the phenotype of Philadelphia chromosome-positive leukemias. Blood ePub Dec 2008
- 58 Faderl S, Garcia-Manero G, Thomas DA, et al. (2002) Philadelphia chromosome-positive acute lymphoblastic leukemia- current concepts and future perspectives. Rev Clin Exp Hematol 6 (2):142-60; discussion 200-2
- 59 (1997) BCR-ABL gene variants. Baillieres Clin. Haematol. 10 (2):203-22
- 60 Jaso J, Thomas DA, Cunningham K, et al. (2011) Prognostic significance of immunophenotypic and karyotypic features of Philadelphia positive Blymphoblastic leukemia in the era of tyrosine kinase inhibitors. Cancer ePub Sep 2011
- 61 Voncken JW, Kaartinen V, Pattengale PK, et al. (1995) BCR/ABL P210 and P190 cause distinct leukemia in transgenic mice. Blood 86 (12):4603-11
- 62 Talpaz M, Shah NP, Kantarjian H, et al. (2006) Dasatinib in imatinib-resistant Philadelphia chromosome-positive leukemias. N. Engl. J. Med. ePub Jun 2006
- 63 Pakakasama S, Kajanachumpol S, Kanjanapongkul S, et al. (2008) Simple multiplex RT-PCR for identifying common fusion transcripts in childhood acute leukemia. Int J Lab Hematol 30 (4):286-91
- 64 van der Veer A, Waanders E, Pieters R, et al. (2013) Independent prognostic value of BCR-ABL1-like signature and IKZF1 deletion, but not high CRLF2 expression, in children with B-cell precursor ALL. Blood ePub Oct 2013
- 65 (2006) Management of acute lymphoblastic leukemia in older patients. Semin. Hematol. 43 (2):126-33

- 66 Li Y, Zou D, Zhao Y, et al. (2010) Clinical characteristics and outcomes of adults with Philadelphia chromosome positive and/or bcr-abl positive acute lymphoblastic leukemia: a single center study from China. Leuk. Lymphoma ePub Mar 2010
- 67 Mullighan CG, Su X, Zhang J, et al. (2009) Deletion of IKZF1 and prognosis in acute lymphoblastic leukemia. N. Engl. J. Med. ePub Jan 2009
- 68 Geng H, Brennan S, Milne TA, et al. (2012) Integrative epigenomic analysis identifies biomarkers and therapeutic targets in adult B-acute lymphoblastic leukemia. Cancer Discov ePub Nov 2012
- 69 Kantarjian HM, Talpaz M, Dhingra K, et al. (1991) Significance of the P210 versus P190 molecular abnormalities in adults with Philadelphia chromosome-positive acute leukemia. Blood 78 (9):2411-8
- 70 Secker-Walker LM, Craig JM, Hawkins JM, et al. (1991) Philadelphia positive acute lymphoblastic leukemia in adults: age distribution, BCR breakpoint and prognostic significance. Leukemia 5 (3):196-9
- 71 Wang Y, Gu M, Mi Y, et al. (2011) Clinical characteristics and outcomes of mixed phenotype acute leukemia with Philadelphia chromosome positive and/or bcr-abl positive in adult. Int. J. Hematol. ePub Dec 2011
- 72 Cimino G, Pane F, Elia L, et al. (2006) The role of BCR/ ABL isoforms in the presentation and outcome of patients with Philadelphia-positive acute lymphoblastic leukemia: a seven-year update of the GIMEMA 0496 trial. Haematologica ePub Mar 2006
- 73 Fagioli F, Zecca M, Rognoni C, et al. (2012) Allogeneic hematopoietic stem cell transplantation for Philadelphia-positive acute lymphoblastic leukemia in children and adolescents: a retrospective multicenter study of the Italian Association of Pediatric Hematology and Oncology (AIEOP). Biol. Blood Marrow Transplant. ePub Jun 2012
- 74 Vignetti M, Fazi P, Cimino G, et al. (2007) Imatinib plus steroids induces complete remissions and prolonged survival in elderly Philadelphia chromosome-positive patients with acute lymphoblastic leukemia without additional chemotherapy: results of the Gruppo Italiano Malattie Ematologiche dell'Adulto (GIMEMA) LAL0201-B protocol. Blood 109 (9):3676-8
- 75 Stirewalt DL, Guthrie KA, Beppu L, et al. (2003) Predictors of relapse and overall survival in Philadelphia chromosome-positive acute lymphoblastic leukemia after transplantation. Biol. Blood Marrow Transplant. 9 (3):206-12
- 76 Radich J, Gehly G, Lee A, et al. (1997) Detection of bcr-abl transcripts in Philadelphia chromosomepositive acute lymphoblastic leukemia after marrow transplantation. Blood 89 (7):2602-9
- 77 (2010) ABL tyrosine kinases: evolution of function, regulation, and specificity. Sci Signal ePub Sep 2010
- 78 Chissoe SL, Bodenteich A, Wang YF, et al. (1995) Sequence and analysis of the human ABL gene, the BCR gene, and regions involved in the Philadelphia chromosomal translocation. Genomics 27 (1):67-82
- 79 Bennour A, Ouahchi I, Achour B, et al. (2013) Analysis of the clinico-hematological relevance of the breakpoint location within M-BCR in chronic myeloid leukemia. Med. Oncol. ePub Mar 2013

- 80 Lugo TG, Pendergast AM, Muller AJ, et al. (1990) Tyrosine kinase activity and transformation potency of bcr-abl oncogene products. Science 247 (4946):1079-82
- 81 Laurent E, Talpaz M, Kantarjian H, et al. (2001) The BCR gene and philadelphia chromosome-positive leukemogenesis. Cancer Res. 61 (6):2343-55
- 82 Kuendgen et al., 2013; ASH Abstract 2757
- 83 Inoue A, Kawakami C, Takitani K, et al. (2014) Azacitidine in the treatment of pediatric therapyrelated myelodysplastic syndrome after allogeneic hematopoietic stem cell transplantation. J. Pediatr. Hematol. Oncol. ePub Jul 2014
- 84 Buchi F, Masala E, Rossi A, et al. (2014) Redistribution of H3K27me3 and acetylated histone H4 upon exposure to azacitidine and decitabine results in derepression of the AML1/ETO target gene IL3. Epigenetics ePub Mar 2014
- 85 Guadagnuolo et al., 2014; ASH Abstract 1030
- 86 Braun T, Itzykson R, Renneville A, et al. (2011) Molecular predictors of response to decitabine in advanced chronic myelomonocytic leukemia: a phase 2 trial. Blood ePub Oct 2011
- 87 Tobiasson M, McLornan DP, Karimi M, et al. (2016) Mutations in histone modulators are associated with prolonged survival during azacitidine therapy. Oncotarget ePub Apr 2016
- 88 Odenike OM, Alkan S, Sher D, et al. (2008) Histone deacetylase inhibitor romidepsin has differential activity in core binding factor acute myeloid leukemia. Clin. Cancer Res. 14 (21):7095-101
- 89 Barbetti V, Gozzini A, Rovida E, et al. (2008) Selective anti-leukaemic activity of low-dose histone deacetylase inhibitor ITF2357 on AML1/ETO-positive cells. Oncogene ePub Mar 2008
- 90 Hu Z, Gu X, Baraoidan K, et al. (2011) RUNX1 regulates corepressor interactions of PU.1. Blood ePub Jun 2011
- 91 Bots M, Verbrugge I, Martin BP, et al. (2014) Differentiation therapy for the treatment of t(8;21) acute myeloid leukemia using histone deacetylase inhibitors. Blood ePub Feb 2014
- 92 Grossmann V, Haferlach C, Weissmann S, et al. (2013) The molecular profile of adult T-cell acute lymphoblastic leukemia: mutations in RUNX1 and DNMT3A are associated with poor prognosis in T-ALL. Genes Chromosomes Cancer ePub Apr 2013
- 93 Rio-Machín A, Menezes J, Maiques-Diaz A, et al. (2012) Abrogation of RUNX1 gene expression in de novo myelodysplastic syndrome with t(4;21)(q21;q22). Haematologica ePub Apr 2012
- 94 Silva FP, Morolli B, Storlazzi CT, et al. (2003) Identification of RUNX1/AML1 as a classical tumor suppressor gene. Oncogene 22 (4):538-47
- 95 Scheitz CJ, Tumbar T (2013) New insights into the role of Runx1 in epithelial stem cell biology and pathology. J. Cell. Biochem. ePub May 2013
- 96 Song WJ, Sullivan MG, Legare RD, et al. (1999) Haploinsufficiency of CBFA2 causes familial thrombocytopenia with propensity to develop acute myelogenous leukaemia. Nat. Genet. 23 (2):166-75

REPORT DATE

APPENDIX

References

- 97 Walker LC, Stevens J, Campbell H, et al. (2002) A novel inherited mutation of the transcription factor RUNX1 causes thrombocytopenia and may predispose to acute myeloid leukaemia. Br. J. Haematol. 117 (4):878-81
- 98 Osato M, Yanagida M, Shigesada K, et al. (2001) Point mutations of the RUNx1/AML1 gene in sporadic and familial myeloid leukemias. Int. J. Hematol. 74 (3):245-51
- 99 Kim JH, Jang JW, Lee YS, et al. (2014) RUNX family members are covalently modified and regulated by PIAS1-mediated sumoylation. Oncogenesis 3 :e101
- 100 Zhao LJ, Wang YY, Li G, et al. (2012) Functional features of RUNX1 mutants in acute transformation of chronic myeloid leukemia and their contribution to inducing murine full-blown leukemia. Blood ePub Mar 2012
- 101 Ottmann O, Dombret H, Martinelli G, et al. (2007) Dasatinib induces rapid hematologic and cytogenetic responses in adult patients with Philadelphia chromosome positive acute lymphoblastic leukemia with resistance or intolerance to imatinib: interim results of a phase 2 study. Blood 110 (7):2309-15
- 102 Ravandi F, O'Brien SM, Cortes JE, et al. (2015) Longterm follow-up of a phase 2 study of chemotherapy plus dasatinib for the initial treatment of patients with Philadelphia chromosome-positive acute lymphoblastic leukemia. Cancer ePub Dec 2015
- 103 Ravandi F, O'Brien S, Thomas D, et al. (2010) First report of phase 2 study of dasatinib with hyper-CVAD for the frontline treatment of patients with Philadelphia chromosome-positive (Ph+) acute lymphoblastic leukemia. Blood ePub Sep 2010
- 104 Sasaki K, Jabbour EJ, Ravandi F, et al. (2016) Hyper-CVAD plus ponatinib versus hyper-CVAD plus dasatinib as frontline therapy for patients with Philadelphia chromosome-positive acute lymphoblastic leukemia: A propensity score analysis. Cancer ePub Dec 2016
- 105 Benjamini O, Dumlao TL, Kantarjian H, et al. (2014) Phase II trial of hyper CVAD and dasatinib in patients with relapsed Philadelphia chromosome positive acute lymphoblastic leukemia or blast phase chronic myeloid leukemia. Am. J. Hematol. ePub Mar 2014
- 106 Slayton WB, Schultz KR, Kairalla JA, et al. (2018) Dasatinib Plus Intensive Chemotherapy in Children, Adolescents, and Young Adults With Philadelphia Chromosome-Positive Acute Lymphoblastic Leukemia: Results of Children's Oncology Group Trial AALL0622. J. Clin. Oncol. ePub Aug 2018
- 107 Schultz KR, Carroll A, Heerema NA, et al. (2014) Longterm follow-up of imatinib in pediatric Philadelphia chromosome-positive acute lymphoblastic leukemia: Children's Oncology Group study AALL0031. Leukemia ePub Jul 2014

- 108 Bachanova V, Marks DJ, Zhang MJ, et al. (2014) Ph+ ALL patients in first complete remission have similar survival after reduced intensity and myeloablative allogeneic transplantation: impact of tyrosine kinase inhibitor and minimal residual disease. Leukemia ePub Mar 2014
- 109 Giebel S, Czyz A, Ottmann O, et al. (2016) Use of tyrosine kinase inhibitors to prevent relapse after allogeneic hematopoietic stem cell transplantation for patients with Philadelphia chromosome-positive acute lymphoblastic leukemia: A position statement of the Acute Leukemia Working Party of the European Society for Blood and Marrow Transplantation. Cancer ePub 10 2016
- 110 Rousselot P, Coudé MM, Gokbuget N, et al. (2016) Dasatinib and low-intensity chemotherapy in elderly patients with Philadelphia chromosome-positive ALL, Blood ePub 08 2016
- 111 Foà R, Vitale A, Vignetti M, et al. (2011) Dasatinib as first-line treatment for adult patients with Philadelphia chromosome-positive acute lymphoblastic leukemia. Blood ePub Dec 2011
- 112 Zwaan CM, Rizzari C, Mechinaud F, et al. (2013) Dasatinib in children and adolescents with relapsed or refractory leukemia: results of the CA180-018 phase I dose-escalation study of the Innovative Therapies for Children with Cancer Consortium. J. Clin. Oncol. ePub Jul 2013
- 113 Deenik W, Beverloo HB, van der Poel-van de Luytgaarde SC, et al. (2009) Rapid complete cytogenetic remission after upfront dasatinib monotherapy in a patient with a NUP214-ABL1-positive T-cell acute lymphoblastic leukemia. Leukemia ePub Mar 2009
- 114 Crombet O. Lastrapes K. Zieske A. et al. (2012) Complete morphologic and molecular remission after introduction of dasatinib in the treatment of a pediatric patient with t-cell acute lymphoblastic leukemia and ABL1 amplification. Pediatr Blood Cancer ePub Aug 2012
- 115 Druker BJ, Guilhot F, O'Brien SG, et al. (2006) Fiveyear follow-up of patients receiving imatinib for chronic myeloid leukemia. N. Engl. J. Med. ePub Dec 2006
- 116 Cohen MH, Williams G, Johnson JR, et al. (2002) Approval summary for imatinib mesylate capsules in the treatment of chronic myelogenous leukemia. Clin. Cancer Res. 8 (5):935-42
- 117 Clarke S, O'Reilly J, Romeo G, et al. (2011) NUP214-ABL1 positive T-cell acute lymphoblastic leukemia patient shows an initial favorable response to imatinib therapy post relapse. Leuk. Res. ePub Jul 2011
- 118 Malone A, Langabeer S, O'Marcaigh A, et al. (2010) A doctor(s) dilemma: ETV6-ABL1 positive acute lymphoblastic leukaemia. Br. J. Haematol. ePub Oct 2010

- 119 Cortes IF, Kim DW, Pinilla-Ibarz I, et al. (2013) A phase 2 trial of ponatinib in Philadelphia chromosome-positive leukemias. N. Engl. J. Med. ePub Nov 2013
- 120 Sasaki et al., 2016; ASCO Abstract 7036
- 121 Gambacorti-Passerini C, Kantarjian HM, Kim DW, et al. (2015) Long-term efficacy and safety of bosutinib in patients with advanced leukemia following resistance/intolerance to imatinib and other tyrosine kinase inhibitors. Am. J. Hematol. ePub Sep 2015
- 122 Jain et al., 2017; ASH Abstract 143
- 123 Assi R, Kantarjian H, Short NJ, et al. (2017) Safety and Efficacy of Blinatumomab in Combination With a Tyrosine Kinase Inhibitor for the Treatment of Relapsed Philadelphia Chromosome-positive Leukemia. Clin Lymphoma Myeloma Leuk ePub Dec 2017
- 124 Khoury HJ, Cortes JE, Kantarjian HM, et al. (2012) Bosutinib is active in chronic phase chronic myeloid leukemia after imatinib and dasatinib and/or nilotinib therapy failure. Blood ePub Apr 2012
- 125 Amsberg GK, Schafhausen P (2013) Bosutinib in the management of chronic myelogenous leukemia. Biologics 7 :115-22
- 126 Komeno Y, Uchida N, Satoh Y, et al. (2017) Bosutinib as a fourth-line therapy for a patient with T315Ipositive lymphoid blastic phase chronic myeloid leukemia: A case report. Oncol Lett 13 (6):4285-4289
- 127 Valent P, Herndlhofer S, Schneeweiß M, et al. (2017) TKI rotation-induced persistent deep molecular response in multi-resistant blast crisis of Ph+ CML. Oncotarget ePub Apr 2017
- 128 Cortes JE, Kim DW, Kantarjian HM, et al. (2012) Bosutinib versus imatinib in newly diagnosed chronic-phase chronic myeloid leukemia: results from the BELA trial. J. Clin. Oncol. ePub Oct 2012
- 129 Cortes JE, Gambacorti-Passerini C, Deininger MW, et al. (2018) Bosutinib Versus Imatinib for Newly Diagnosed Chronic Myeloid Leukemia: Results From the Randomized BFORE Trial. J. Clin. Oncol. ePub Jan 2018
- 130 Atilla E, Ataca P, Ozyurek E, et al. (2015) Successful Bosutinib Experience in an Elderly Acute Lymphoblastic Leukemia Patient with Suspected Central Nervous System Involvement Transformed from Chronic Myeloid Leukemia. Case Rep Hematol 2015 :689423
- 131 Kim DY, Joo YD, Lim SN, et al. (2015) Nilotinib combined with multiagent chemotherapy for newly diagnosed Philadelphia-positive acute lymphoblastic leukemia. Blood ePub Aug 2015
- 132 Ottmann et al., 2014; ASH Abstract 798
- 133 Kantarjian H, Giles F, Wunderle L, et al. (2006) Nilotinib in imatinib-resistant CML and Philadelphia chromosome-positive ALL. N. Engl. J. Med. ePub Jun 2006